

**Cloning and Transgenesis** 

## Improving AAV Gene Therapy: Graduating From Transgene Expression "Everywhere, All The Time" To "Disease-Specific"

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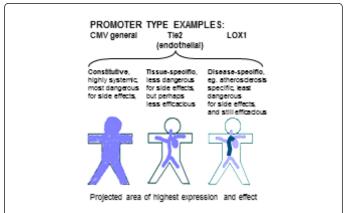
## Introduction

Adeno-associated virus (AAV) is now one of the top gene delivery/ gene therapy vectors in use today [1-6]. The hallmark goal of gene therapy is to deliver a therapeutic gene which then counteracts a negative phenotype or disease within the patient or animal model. While there are many types of diseases to treat, each disease is somewhat different and there are a variety of delivery/expression strategies which can be undertaken. AAV capsid type and tropism clearly play a valuable role in gene therapy; however the strategy for expressing the transgene is even more important. Here we discuss the three most prominent types of gene expression approaches, that is, which transcriptional promoter should be chosen for expressing the therapeutic gene. The issue is that many, perhaps all, therapeutic genes will likely have consequences, adverse reactions, if expressed at high levels. Yet the gene therapy agents must be safe, and not induce widespread unintended damage.

The first major expression approach is the "constitutive" approach, such as using the cytomegalovirus (CMV) immediate early promoter (pr) (Figure 1).

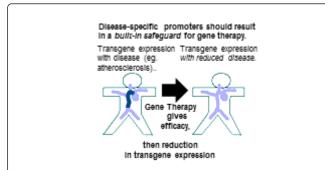
However there are a number of competitors in this category [8]. The treatment of genetic syndromes might be the most appropriate for this approach. Genetic syndromes result from a faulty protein which is important within a tissue or organ for normal function. For genetic syndromes the goal of gene therapy is simple; get that therapeutic gene and its protein delivered and expressed into as many cells of the patient, organ or tissue as possible to give back normal function. The strategy then is usually for maximum gene delivery and gene expression. A similar approach might also be needed for the treatment of cancer which is a malignant tissue and requires maximum gene delivery of the cancer-killing or cancer-altering gene.

Many, perhaps most, therapeutic genes actually have a down-side because of their inherent function and give adverse reactions when expressed at high levels. An example of this is in the treatment of inflammatory diseases, such as cardiovascular disease. Therapeutic genes that we have used in the treatment of atherosclerosis, in particular interleukin 10 (IL10), which, while strongly immunosuppressive (needed for its efficacy against atherosclerosis), is also associated with a number of adverse reactions which are manifested in the clinical setting and in some animal models. These adverse reactions include increased bacterial, fungal and viral infections, cancer, headache, and anemia [9-16]. Thus, as a generalization, the strongest therapeutic genes are also likely to be the most dangerous and they must be tightly regulated. We call this strong constitutive transcriptional promoter use, introducing and expressing the gene in many cell types, as the "everywhere, all the time" approach. We consider this approach to be somewhat dangerous. Only in the treatment of genetic syndromes does this strategy seem appropriate.



**Figure 1:** The three general approaches in therapeutic gene delivery. CMVpr represents the constitutive "everywhere, all the time" approach. It is dangerous. Tie2pr represents the somewhat better tissue specific approach. The LOX1pr represents the atherosclerosis-specific, disease-specific approach. It is the safest approach, yet still gives efficacy [7]. This proposal intends to further verify and make further advances in the latter, the disease-specific approach. This cartoon is not intended to be completely accurate in showing specific expression patterns, but rather to show the differences between the approaches.

A major refinement of the constitutive approach is the "tissuespecific" approach, the second general approach. In this scheme the delivered transgene is to be expressed only within a specific cell type, thereby limiting its overall expression [17]. Clearly this is an improvement over the constitutive approach, yet we consider this approach as only a modified version of the constitutive approach, limited to a specific cell type. This is still rather dangerous as one still changes, wholesale, an entire tissue. This expression strategy may be, in fact, superior for most genetic syndromes than the constitutive approach as usually one tissue is overly affected and limited expression to only that one tissue can give efficacy without the chance of unintended consequences in other tissues. However, the genemodified tissue possibly then becomes essentially a new tissue type, with perhaps a new, still abnormal, phenotype.



**Figure 2:** The disease-specific expression approach provides a builtin safeguard for the gene therapy. As planned and envisioned the use of the disease-specific transcriptional promoter within the AAV vector will give targeted expression at the site of disease, and little expression elsewhere. Additionally, when the level of disease is reduced then so too should the expression of the therapeutic transgene. This should result in a built-in safeguard for the gene therapy and prevent the appearance of adverse reactions due to transgene overexpression

Here, however, we wish to highlight the third approach, the "disease-specific" approach [4]. The goal of the disease-specific approach is to limit the expression of the therapeutic transgene to the site of disease, and in the cells which are changing towards the diseaseassociated phenotype. We view this disease-specific approach as highly desirable in particular for diseases which are related to inflammation, which encompasses a long list of major diseases which increase in prevalence as we age (atherosclerosis, dementia, arthritis and so on). The disease-specific approach gives the gene therapy an important safety feature against adverse reactions from the over expression of a powerful therapeutic transgene such as IL10. The prototype diseasespecific promoter which we have recently used to express IL10 for the treatment of atherosclerosis in a mouse model is the LOX1 promoter (LOX1pr) [4]. A variety of stimuli up- and down-regulate LOX1pr, so it is a broadly responsive promoter which is also active in a variety of cell types [17]. Our recent study on the use of the LOX1pr to drive expression of the IL10 cytokine gene within the AAV vector to inhibit atherogenesis in the low density lipoprotein receptor knockout mouse on high cholesterol diet (LDLR KO-HCD) demonstrates that this disease-specific strategy is viable [4]. While mice do not readily demonstrate adverse reactions for IL10 expression as has been documented in humans [9-16], it was observed that the overall level of human IL10 gene expression was much lower when the LOX1pr was used compared to that when the CMVpr was used. In spite of this lower overall expression of hIL10 the disease-specific approach (using LOX1pr) resulted in aortic measurements which were statistically not significant, therapeutically the same, as those resulting from the use of the constitutive approach (CMVpr)[4]. Thus, the disease-specific nature of promoters such as LOX1 should minimize the overall transgene expression, and thereby should limit adverse reactions. Importantly, the therapeutic gene is expressed where it is needed most, at the site of disease. Moreover, if the disease-stimulus is eliminated or reduced (after the disease-specific gene therapy treatment is giving a reduced level of disease), then, in response, the transgene expression, from the disease-specific promoter, itself, should be down-regulated.

That promoters should function as envisioned within the AAV vector can be expected due to the lack of strong promoter elements within the AAV inverted terminal repeats [17,18].

In conclusion, the disease-specific approach, using promoters which respond to disease-stimuli, is an under-studied area of gene therapy. Yet we consider the disease-specific approach to be critical to the success of the field of gene therapy. We believe that the diseasespecific approach will be grow in use and popularity because of its advantages, most importantly because it provides a built-in safeguard against the dangers of transgene overexpression (Figure 2).

Now the quest starts for the identification of appropriate diseasespecific promoters for use in AAV vectorology.

## References

- Hermonat PL and Muzyczka N (1984) Use of adeno-associated virus as a mammalian DNA cloning vector, transduction of neomycin resistance into mammalian tissue culture cells. Proc Natl Acad Sci USA 81: 6466-6470
- Hermonat PL (2014) The first adeno-associated virus gene transfer experiment, (1983) Human Gene Therapy 9: e104596
- Buchlis G, Podsakoff GM, Radu A, Hawk SM, Flake AW, et al. (2012) Factor IX expression in skeletal muscle of a severe hemophilia B patient 10 years after AAV-mediated gene transfer. Blood 119: 3038-3041
- 4. Zhu H, Cao M, Chriva-Internati M and Hermonat PL (2014) Comparison of efficacy of human interleukin 10, expressed from the disease-specific LOX1 or constitutive cytomegalovirus promoters, against atherosclerosis in mice using adeno-associated virus 2/8 delivery. PLoS ONE 9: e94665
- Zhu H, Cao M, Figueroa JA, Cobos E, Uretsky BF, Chiriva-Internati M, et al. (2014) AAV2/8-hSMAD3 gene delivery attenuates aortic atherogenesis, enhances Th2 response, without inducing COL1A2/2A1 and CTGF (fibrosis) in LDLR-KO mice on high cholesterol diet. J Transl Med 12: 252
- Zhu H, Cao M, Straub KD and Hermonat PL (2013) Systemic delivery of thiol-specific antioxidant hPRDX6 gene by AAV2/8 inhibits atherogenesis in LDLR KO mice on HCD. Genetic Syndromes and Gene Therapy 4: 3
- Brooks DG, A. M. Lee, H. Elsaesser, D. B. McGavern and M. B. Oldstone (2008) IL-10 blockade facilitates DNA vaccine-induced T cell responses and enhances clearance of persistent virus infection. J. Exp Med 205: 533-541
- Damdindorj L, Karnan S, Ota A, Hossain E, Konishi Y, et al. (2014) A comparative analysis of constitutive promoters located in adenoassociated viral vectors. PLoS One 9: e106472
- 9. Zobel, Katrin. Martus, Peter. Pletz, Mathias W. Ewig, Santiago. Prediger, et al. (2012) Interleukin 6, lipopolysaccharide-binding protein and interleukin 10 in the prediction of risk and etiologic patterns in patients with community-acquired pneumonia: results from the German competence network CAPNETZ. BMC Pulmonary Med 12: 6
- Clemons KV, Grunig G, Sobel RA, Mirels LF, Rennick DM et al. (2000) Role of IL-10 in invasive aspergillosis: increased resistance of IL-10 gene knockout mice to lethal systemic aspergillosis. Clin Exp Imm 122: 186-91
- 11. Filippi CM and Von Herrath MG (2008) IL-10 and the resolution of infections. High levels of IL10 are associated with increased viral, bacterial, and fungal infections, as well as cancer. J Pathol 214: 224-230
- Brooks DG, Trifilo MJ, Edelmann KH, Teyton L, DB, et al. (2006) Interleukin-10 determines viral clearance or persistence in vivo. Nat Med 12: 1301-1309
- Ejrnaes M, Filippi CM, Martinic MM, Ling EM, Togher LM et al. (2006) Resolution of a chronic viral infection after interleukin-10 receptor blockade. J Exp Med 203: 2461-2472

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- Zeni E, Mazzetti L, Miotto D, Lo Cascio N, Maestrelli P, et al. (2007) Macrophage expression of interleukin-10 is a prognostic factoir in nonsmall cell lung cancer. Eur Respir J 30: 627-632
- 15. Maris CH, Chappell CP and Jacob J (2007) Interleukin-10 plays an early role in generating virus-specific T cell anergy. BMC Immunol 8: 8
- Herfarth H and Schölmerich J (2002) IL-10 therapy in Crohn's disease: at the crossroads. Gut 50: 146–147
- 17. Hermonat PL, Zhou H, Cao M and Mehta JL. LOX-1 transcription (2011) Cardiovasc Drugs Ther 25: 393-400
- Flotte TR, Afione SA, Solow R, Drumm ML, Markakis D, et al. (1993) Expression of the cystic fibrosis transmembrane conductance regulator from a novel adeno-associated virus promoter. J Biol Chem 268: 3781-3790