

Research Article

Open Access

Immunopharmacological Activity of Saponin from *Terminalia arjuna* and *Prosopis spicigera*

Amit Gupta* and Sushama R Chaphalkar

Department of Immunology and Virology, Vidya Pratishthan's School of Biotechnology, Maharashtra, India

Abstract

The aim of the present work was to examine the immunopharmacological (anti-microbial and anti-inflammatory) effect of saponin on human peripheral blood mononuclear cells (PBMC) that are isolated as well as purified from infected (virally) and non-infected (normal) human whole blood samples. However, there is little information available concerning the effect of crude saponin on human PBMC in case of infected and non-infected human PBMCs. Saponin, one of the members of secondary metabolites that are isolated from aqueous leaves extract of *Terminalia arjuna* and *Prosopis spicigera*. Our group evaluated the immunopharmacological activity of saponin (0.5–30 mg/ml, 100 μ l) in human PBMC were determined and measured cell proliferation using Concanavalin (Con A, 2.5 μ g/ml, 50 μ l) and estimate Th1 (IFN-gamma and TNF alpha) type of cytokines from cell culture supernatant including nitric oxide production.

The results showed that crude saponin from *Terminalia arjuna* and *Prosopis spicigera* showed its inhibitory activity at higher doses (30 mg/ml, 100 μ l) in Con A induced PBMC (infected and non-infected) proliferation. In addition, these saponin also inhibits its Th1 (IFN-gamma and TNF alpha) type of cytokines from Con A stimulated cell culture supernatant of PBMC and also inhibits nitric oxide production in a dose dependent manner. In conclusion, saponin appears to have potential as an anti-microbial and anti-inflammatory agent.

Keywords: *Terminalia arjuna*; *Prosopis spicigera*; Saponin; Cytokines; Nitric oxide; Anti-inflammatory; Anti-microbial

Introduction

Medicinal plants have been used extensively as a source for numerous bioactive constituents for treating human diseases [1]. There are number of phytochemicals that are present in many medicinal plant products and showed its prophylactic as well as therapeutic value [2,3]. These are widely used in the veterinary, agriculture, scientific research etc. Out of these phytochemicals, saponins are generally predominant especially triterpenoids in wild plants and cultivated crops whereas steroid saponins are common in plants used as herbs or for their health-promoting properties [4,5].

Saponins are steroid or triterpene glycoside compounds that are present in a variety of medicinal plants e.g. *Azadirachta indica* [6], *Ficus religiosa* [7], *Ficus benghalensis* [8], *Prosopis spicigera* [9], etc. Several preclinical and clinical studies related to Saponins have already been done with respect to various immunopharmacological activities, i.e., anti-inflammatory [10], vaccine adjuvants [11], cytotoxic [12] etc. Some *in vitro* and *in vivo* experiments in animal model studies that demonstrate the beneficial or harmful effects of saponin are reported or some still under investigation. In view of this, there is an urgent need for systematic evaluation of the most active structural components of the saponins and their interaction with the specific protein antigen including microbial community, the host animal, etc. Along with these studies, the direct effects of saponins on human whole blood must be examined in order to get the full understanding of its metabolism and beneficial effects of saponins on human. Knowledge of the chemical constituents of plant is desirable for the synthesis of complex chemical substances.

Terminalia arjuna (evergreen tree; family Combretaceae), versatile medicinal plant in India having a wide spectrum of immunobiological activity, e.g. anti-microbial, anti-coagulant, antipyretic, astringent, anti-inflammatory, cardiogenic, etc. [13,14]. In spite of these activities, many useful phytoconstituents that are isolated from *Terminalia arjuna* which included triterpenoids for cardiovascular properties; flavonoids

including tannins for anticancer and antimicrobial properties and so on [13,14].

Prosopis spicigera Linn. (Shami; Family Fabaceae; xerophytic and draught resistant plant) showed several immunopharmacological activities, i.e., anti-inflammatory, antimicrobial, etc. [9]. One of the novel alkaloid is reported in this medicinal plant, i.e., piperidine-3-ol alkaloid called as spicigerin [9]. The purpose of the study was to find out the anti-inflammatory and antimicrobial activity of crude saponin extracted from the leaves of *Terminalia arjuna* and *Prosopis spicigera*.

Materials and Method**Collection of plant material**

Fresh mature plant leaves of *Terminalia arjuna* and *Prosopis spicigera* were collected from Vidya Pratishthan's, garden at Baramati. First of all, leaves were washed with tap water in order to remove the pollutants that are present on the surface and then the leaves were dried in a shady area.

Preparation of plant (crude saponin) material

Fresh dried shady leaves of *Terminalia arjuna* and *Prosopis spicigera*, medicinal plants were collected in order to isolate the crude

*Corresponding author: Amit Gupta, Assistant Professor (Immunology), Department of Immunology and Virology, Vidya Pratishthan's School of Biotechnology (VSBT, Research Centre affiliated to Savitribai Phule Pune University) Baramati, Maharashtra, India, Tel: +918308881506, E-mail: amitvsbt@gmail.com; amitgupta@vsbt.res.in; amitri@yahoo.com

Received December 07, 2015; Accepted December 29, 2015; Published December 31, 2015

Citation: Gupta A, Chaphalkar SR (2015) Immunopharmacological Activity of Saponin from *Terminalia arjuna* and *Prosopis spicigera*. J Pharma Reports 1: 102.

Copyright: © 2015 Gupta A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

saponins using this technique, i.e., liquid-liquid extraction method. Dried plant (6 g) leaves of both the medicinal plants were macerated in liquid nitrogen and finally macerated and dissolved in phosphate buffered saline (60 ml, pH 7.2) and then filtered it. For the preparation of crude saponin from leaves aqueous extract using diethyl ether, one third volume dissolved in aqueous extract and incubated for 5–10 min. Later on, upper layer containing diethyl ether was discarded (process repeated two times). At the bottom, aqueous layer was retained and add double volume amount of n-butanol and incubate for 5–10 min. Finally, extracts (nbutanol) were bulked together and washed two times using 10 ml of 5% NaCl. Afterwards, washed aqueous extract was concentrated at <70°C in an oven and air dried at room temperature to yield mg of crude saponin residue [4,5]. Overall, crude residue of saponin was screened using foaming test.

Immunopharmacological studies (anti-inflammatory and anti-microbial studies), i.e., PBMC proliferation and estimation of Th1 type of cytokines including nitric oxide production from cell culture supernatant

Virally Infected (major symptoms such as cough, common cold, fever, etc.) and non-infected (normal) EDTA blood samples of human were collected from Mangal Pathology laboratory, Baramati for anti-inflammatory and anti-microbial studies. The inclusion criteria for these studies i.e. informed consent letters were collected from potential volunteers in order to separate the human peripheral blood mononuclear cells (PBMC) from healthy volunteers and also from infected person showed such type of symptoms as mentioned above where as exclusion of this study is that non-healthy person was not included in non-infected cases. In spite of this, our study has been thoroughly explained to all the volunteers.

For immunopharmacological studies, using human PBMC (10^6 cells/ml, 100 μ l) were separated from human whole blood (infected and non-infected blood) by means of Ficoll–Hypaque gradient centrifugation and plated in 96 well plate in presence of Concanavalin A (Con A, 2.5 μ g/ml, 50 μ l) for 48 h in carbon dioxide incubator (37°C, 5% CO₂). After incubation, tissue culture plates were centrifuged at 1800 rpm for 15 minutes and then the supernatant (100 μ l) was collected for analyzing Th1 (IFN-gamma and TNF alpha) type of cytokines including nitric oxide production and then add equal volume of fresh medium. Afterwards, MTT solution (5 mg/ml, 20 μ l) were added to each well and then incubated for 4 h. Again, the 96 well flat bottom plates were spinned or centrifuged at 1500 rpm for 4 minutes and the supernatant was discarded [3,8-10]. Add DMSO (100 μ l) solution to the formazon crystals and the absorbance was evaluated in an ELISA reader at 570 nm.

Th1 (IFN-gamma and TNF alpha) were measured with an enzyme linked immunosorbent assay (BD optia kit, ELISA) according to the instructions of the manufacturer. This assay employs the quantitative sandwich enzyme immunoassay technique. Human monoclonal antibody for Th1 (IFN-gamma and TNF alpha) has been pre-coated on to a microplate (96 well, Elisa plate). Standards, control including cell supernatant samples of saponin treated with human PBMC are pipetted into the wells and any human IFN-gamma and TNF alpha is present is bound by the immobilized antibody. After washing away any unbound substances, an enzyme linked polyclonal antibody specific for human IFN-gamma and TNF alpha is added to the wells. The enzyme reaction yields a blue product that turns yellow when the stop solution is added. The intensity of the colour measured is in proportion to the amount of IFN-gamma and TNF alpha bound in the initial step. The sample values are then read off the standard curve.

For nitric oxide production, cell culture supernatant of PBMC along with variable doses of crude saponin from *Terminalia arjuna* and *Prosopis spicigera* were collected [13]. Briefly, 50 μ l of cell culture supernatant of PBMC along with variable doses of saponin was mixed with equal volume of Griess reagent [1% sulfanilamide and 0.1% naphthylethylenediamine dihydrochloride (NED) in 2–2.5% phosphoric acid]. Incubate 96 well flat bottom plates at room temperature for 7–8 minutes and the absorbance or optical density was measured at 540 nm in a microplate reader. The culture medium (phosphate buffered saline containing 10% fetal bovine serum) was used as a blank. The quantity of nitrite (μ M) was evaluated using sodium nitrite in order to generate standard curve.

Statistical analysis

Data represented as Mean \pm S.E using one way ANOVA test (Boniferroni multiple comparison test).

Results

PBMC proliferation assay and Th1 cytokines

Saponin extracted from the aqueous extract of *Terminalia arjuna* and *Prosopis spicigera* used in the traditional system of medicine were tested for its ability to inhibit human PBMC proliferation at a concentration between 0.5–30 mg/ml, 100 μ l. In order to determine its effect after 48 h of crude saponin on Con A stimulated human PBMC (infected and non-infected) proliferation and estimate Th1 (IFN-gamma and TNF alpha) cytokines from cell culture supernatant as shown in Figures 1 and 2. The results showed that crude saponin at higher doses (30 mg/ml, 100 μ l) showed decline in PBMC proliferation and Th1 (IFN-gamma and TNF alpha) cytokines from cell culture supernatant as compared to control. On the other hand, Con A used as mitogen for T cell proliferation and the results showed significant enhancement in proliferation and Th1 (IFN-gamma and TNF alpha) cytokines as compared to control. Overall, saponin from both the plants showed its anti-inflammatory and anti-microbial activity.

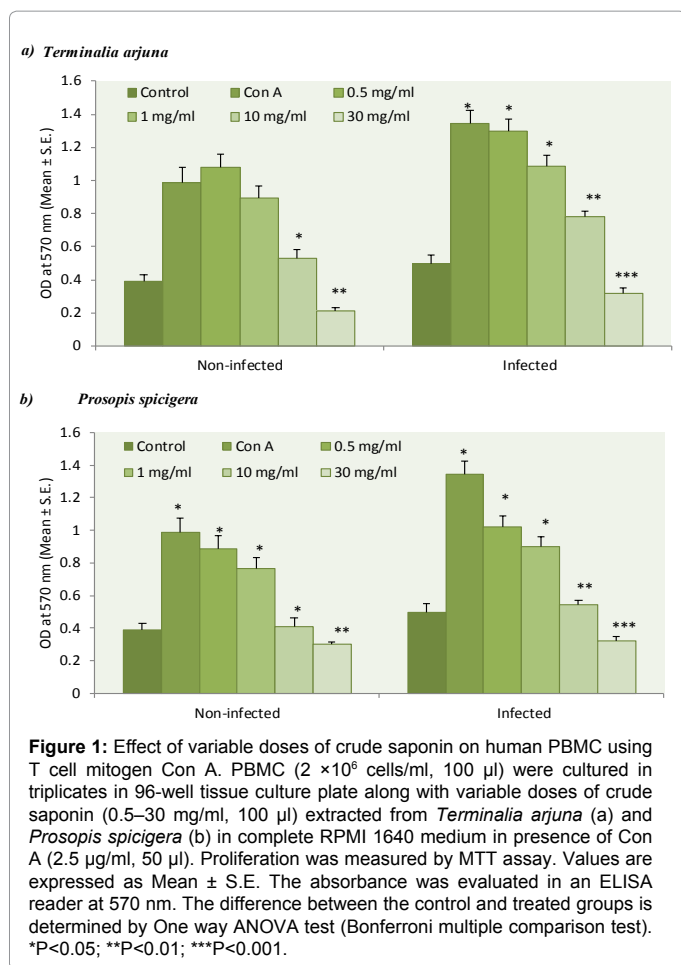
Nitric oxide production

For the estimation of nitric oxide production from PBMC (infected and non-infected) cell culture supernatant as shown in Figure 3. The results showed that crude saponin showed similar results i.e. decline in nitric production at higher doses. Overall, crude saponin data represents and showed its anti-microbial and anti-inflammatory activity.

Discussion

In the present study, human PBMCs (monocytes and T cells including lesser extent of killer and dendritic cells) are used as a screening assay which is already established in clinical immunology for the last so many years and constitutes one of the important component of peripheral immune system [15,16]. The objective of our study is to provide the relationship between proliferation, estimation of Th1 type of cytokines from cell culture supernatant including nitric oxide production in order to evaluate this *in vitro* test system for screening of potential anti-inflammatory and anti-microbial properties of saponin extracted from aqueous leaves extract of *Terminalia arjuna* and *Prosopis spicigera*.

In the present study, T cells were the major proliferating cells in Con A-induced activated PBMCs. Activation and clonal expansion of human PBMCs from infected and non-infected human whole blood which contained mostly T lymphocytes that played a vital role in the



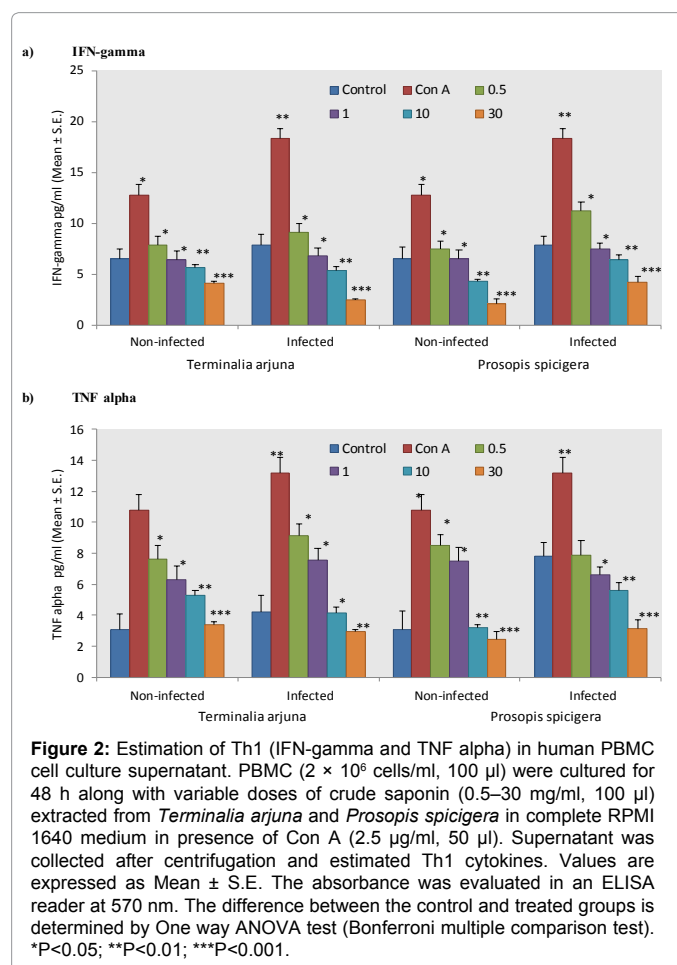
generation of immune responses [17]. Number of studies has shown that Th1 (IFN-gamma and TNF alpha) cytokines are important in the growth of antigen or Con A-induced PBMCs. Thus, the inhibitory effect of saponin from *Terminalia arjuna* and *Prosopis spicigera* on Con A-activated PBMC proliferation in case of infected and non-infected PBMC might have been caused by the suppression of T cell proliferation. The present study indicated that proliferation of Con A-activated PBMC was significantly inhibited by 30 mg/ml, 100 μ l in both these Saponins extracted from *Terminalia arjuna* and *Prosopis spicigera*. On the other hand, this study also revealed its inhibitory activity of crude saponin on nonspecific cellular immune response. Although the exact mechanism of this crude mechanism extracted from the leaves of *Terminalia arjuna* and *Prosopis spicigera* is not known, it may be mediated by the interaction between the most active components of saponin from these medicinal plants and cell-surface molecules or growth factors involving mitogen (Con A) activation, and it is possible that identification and elucidation of the active constituents in this medicinal plants may provide useful leads to the development of new and effective immunosuppressant drugs.

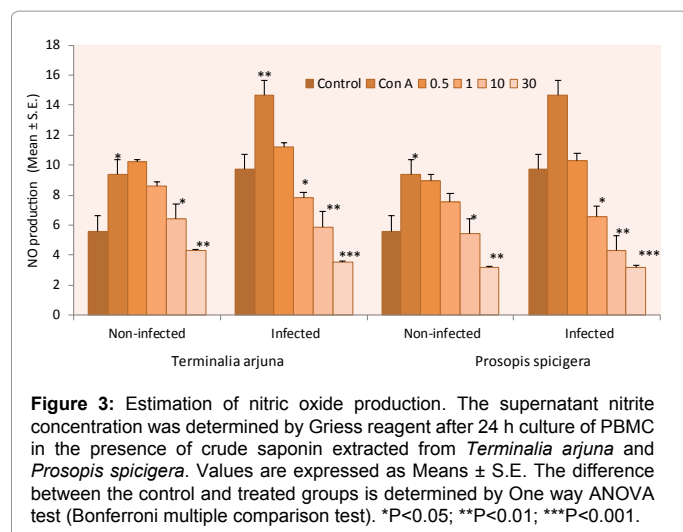
Immunopharmacological activity of saponin can be measured by detecting the changes in the production of immune molecules such as cytokines and nitric oxide production. To elucidate the saponin biological effect on human PBMCs, our group measured the levels of Th1 (IFN-gamma and TNF alpha) cytokines using ELISA. Pro-inflammatory cytokines (IFN-gamma and TNF alpha) are involved in the regulation of PBMC proliferation [18]. Our results showed that

addition of saponin from *Terminalia arjuna* and *Prosopis spicigera* significantly declined in the production in Con A-stimulated PBMCs in case of infected and non-infected human PBMC. Overall, the data represents that saponin from *Terminalia arjuna* and *Prosopis spicigera* showed its anti-microbial and anti-inflammatory activity in case of infected and non-infected human blood samples because PBMCs fail to proliferate due to the suppression of Th1 (IFN-gamma and TNF alpha) pro-inflammatory cytokines. In contrast, saponin showed less toxicity in case of normal human PBMCs as evaluated by the Trypan blue exclusion method (data not shown). Most importantly, in human PBMC the production of nitric oxide from saponin extracted from *Terminalia arjuna* and *Prosopis spicigera* is tightly correlated with the extent of cytokine production. In this study, saponin has the potential to inhibit the nitric oxide production in case of infected and non-infected human PBMC stimulated with Con A. For these studies, cell culture supernatant collected from treated group of variable doses of saponin in human PBMC inhibited nitric oxide production at higher doses when activated *in vitro* with Con A at 48 h after stimulation as compared to control. Overall, nitric oxide assay may be regarded as an efficient, economical and relatively reliable tool in primary screening for anti-inflammatory and anti-microbial activities of saponin in human PBMC. Overall, the results showed its anti-inflammatory and anti-microbial activities.

Conclusion

The present study indicates that crude saponin from *Terminalia arjuna* and *Prosopis spicigera* got profound anti-inflammatory and





antimicrobial effect and may have potential use in medicine. This novel finding will aid us to conduct bioactivity guided isolation of saponin derivatives and characterized the compounds in due course of time.

References

- Gupta A, Khamkar PR, Chaphalkar SR (2014) Review on medicinal plants to target and inhibit the epidermal growth factor receptor signaling in cancer and tissue repair therapy. International Journal of Advances in Pharmacy, Biology and chemistry 3: 210- 213.
- Gupta A, Khamkar PR, Chaphalkar SR (2014) Applications and uses of active ingredients from medicinal plants. Indian Journal of novel drug delivery 6: 106-111.
- Gupta A, Chaphalkar SR (2015) Inhibitory potential of aqueous leaves extract of *Messua ferrea* and *Mimusops elengi* on antigen specific immune response using human whole blood. Asian Journal of medical and Pharmaceutical researches 5: 22-26.
- Gupta A, Chaphalkar SR (2015) Immunosuppressive activity of crude saponins from the leaves of *Calotropis gigantean*, *Calamus rotang* and *Artocarpus integrifolia*. International Journal of Pharma sciences and research 5: 1-5.
- Gupta A, Chaphalkar SR (2015) Flow cytometric analysis of crude saponin from the leaves of *Mangifera indica* and *Anthocephalus cadamba* for its anti-inflammatory activity. European Journal of Biomedical and Pharmaceutical Sciences 2: 163-173.
- Gupta A, Chaphalkar SR (2015) Immunoadjuvant potential of *Azadirachta indica* against rabies, hepatitis and DPT vaccine antigen. International Journal of Medical and pharmaceutical sciences 5: 1-5.
- Gupta A, Khamkar PR, Chaphalkar SR (2014) Immunomodulatory and anti-inflammatory activity of aqueous extract of leaf, stem and root of *Ficus religiosa* on human whole blood and peripheral blood mononuclear cells. International Journal of Medicine and Pharmaceutical research 2: 732-739.
- Gupta A, Chaphalkar SR (2014) Inhibitory potential of aqueous extract of *Ficus bengalensis* on human peripheral blood mononuclear cells. Journal of Pharma research 3: 250-253.
- Gupta A, Chaphalkar SR (2015) Anti-inflammatory activity of aqueous extract of leaves of *Prosopis spicigera*. International Journal of Research in pharmacy and life sciences 3: 829-834.
- Gupta A, Khamkar PR, Chaphalkar SR (2014) Anti-inflammatory activity of aqueous extract of *Mimusops elengi* on human whole blood and peripheral blood mononuclear cells. International Journal of Current trends in Pharmaceutical research 2: 638-645.
- Gupta A, Chaphalkar SR (2015) Flow cytometric analysis of immunoadjuvant activity of *Emblica officinalis* on human whole blood. World Journal of Pharmaceutical research 4: 1063-1071.
- Gupta A, Chaphalkar SR (2015) Cytotoxic and anti-inflammatory activity of aqueous extract of *Strychnos nuxvomica*. Journal of Biology and nature 4: 217-223.
- Gupta A, Khamkar PR, Chaphalkar SR (2014) Inhibition of nitric oxide production and pro-inflammatory cytokines by aqueous extract of *Terminalia arjuna* in human peripheral blood mononuclear cells. International Journal of pharmaceutical and biological science archive 2: 29-33.
- Gupta A, Chaphalkar SR (2016) Inhibition of antigen specific T cell population using *Calotropis gigantea* and *Terminalia arjuna*. Journal of Biology and nature.
- Gupta A, Chaphalkar SR (2016) Immunopharmacological screening of aqueous root extract of *Santalum album*. Journal of Herb Med Pharmacology 5: 07-11.
- Khajuria A, Gupta A, Singh S, Malik F, Singh J (2007) RLJ-NE-299A: A new plant based vaccine adjuvant. Vaccine 25: 2706-2715.
- Bhat BA, Dhar KL, Puri SC, Qurishi MA, Khajuria A (2005) Isolation, characterization and biological evaluation of *Datura lactones* as potential immunomodulators. Bioorg Med Chem 13: 6672-6677.
- Puri SC, Amna T, Khajuria A, Gupta A, Spitteller M (2007) Immunomodulatory activity of an extract of the novel fungal endophyte *Entrophospora infrequens* isolated from *Nothapodytes foetida* (Wight) sleumer. Acta Microbiol Immunol Hung 54: 237-260.