

IL4I1: An Emerging Target to Reinvigorate Antitumor Immune Responses

Flavia Castellano^{1,2,3*} and Valérie Molinier-Frenkel^{1,2,4*}

¹INSERM, U955, Equipe 09, Créteil, France

²Université Paris Est, Faculté de Médecine, Créteil, France

³AP-HP, Hôpital H. Mondor - A. Chenevier, Plateforme de Ressources Biologiques, Créteil, France

⁴AP-HP, Hôpital H. Mondor - A. Chenevier, Service d'Immunologie Biologique, Créteil, France

*Corresponding authors: Flavia Castellano and Valérie Molinier-Frenkel, INSERM U955, IMRB eq 09, Hôpital Henri Mondor, 51 av du Maréchal de Lattre de Tassigny, F-94010 Créteil cedex, France; Tel: +33 1 49 81 37 65; Fax +33 1 49 81 35 66; flavia.castellano@inserm.fr, valerie.frenkel@inserm.fr

Received date: December 6, 2016; Accepted date: January 2, 2017; Published date: January 23, 2017

Copyright: © 2017 Castellano F, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Keywords: B-cell; Tumour; Lymphoma; Immunotherapy

Short Communication

One of the known cancer-promoting mechanisms dependent on the microenvironment consists of the expression of amino-acid catabolizing enzymes. The enzyme Interleukin-4 Induced gene 1 (IL4I1) is a secreted L-phenylalanine oxidase physiologically produced by antigen presenting cells of the myeloid and B cell lineage and in T helper type 17 cells. IL411 inhibits T cell proliferation and cytokine production, and facilitates naïve CD4⁺ T cells differentiation into regulatory T cells in vitro, in part via its capacity to produce H2O2 and deplete phenylalanine from the T cell microenvironment. In humans, IL411 is expressed in tumor-associated macrophages of most cancers and in tumor B cells of lymphoma originating from the germinal center. As shown in a mouse model of melanoma, its local production may facilitate human tumor growth by inhibiting the anti-tumor CD8+ T cell response. All these data, and others that are developed in this short review, point to IL4I1 as a new drugable target in the field of immunomodulation in cancer.

The research of the last decades has highlighted the dual role of the immune system in cancer development. Whereas the induction of a specific antitumor immune response can play an active role in cancer cell elimination [1], the selective pressure it exerts leads to the development of immunosuppressive mechanisms that facilitate subsequent tumor cell escape from the immune control. This last immunosubversive ability, which has been included as a cancer hallmark in 2011, offers new potential targets for therapeutic intervention in patients [2,3].

Different types of leukocytes infiltrate tumors and can affect local immune responses, but tumour-associated macrophages (TAMs) usually predominate and their density or transcriptional signature has been associated with poor prognosis and reduced survival in a large number of cancers [4,5]. One of the major TAM cancer promoting mechanism consists of the expression of amino-acid catabolizing enzymes [6]. The best-characterized enzymes belonging to this group, i.e. indoleamine 2,3 dioxygenase (IDO), arginase 1 (Arg 1) and inducible nitric oxid synthase (iNOS) participate in the inhibition of the adaptive immune response by decreasing essential or semiessential amino acids and producing different toxic metabolites in the lymphocyte environment, thus affecting their proliferation and function. Expression of these enzymes in the tumor bed and/or in lymphoid organs has a negative impact on survival. This can be reversed by the use of specific inhibitors in pre-clinical models, justifying the current phase I/II clinical trials of IDO inhibitors.

Interleukin-4 induced gene I (*IL411*) was initially described as an early-induced gene by IL4 treatment of B splenocytes [7] with high homology to L-amino acid oxidases. This protein corresponds to isoform 1, whose expression is restricted to lymphoid tissues [8]. In 2005, a second isoform, was described in rare cells of nervous tissue (e.g. Purkinje cells) and of the testis (Sertoli cells) [9]. Since then, three other isoforms have been referenced in databases. All these isoforms differ in the first exons encoding their respective signal peptides that should be cleaved at homologous positions in the proteins. Consequently, the five proteins should have identical sequences once processed.

Phe + H₂O + O₂ \longrightarrow phenylpyruvate + NH₃ + H₂O₂

Figure 1: L-aminoacid oxidation catalyzed by *IL411. IL411* degrades L-phenylalanine (Phe) to produce phenylpyruvate, hydrogen peroxide (H_2O_2) and ammonia (NH₃). An alternative substrate of *IL411* is arginin (not depicted).

Following the discovery of IL4I1 expression in primary mediastinal B cell lymphoma [10], we have shown that the isoform 1 of IL4I1 is a glycosylated and secreted protein endowed with an L-amino acid oxidase activity [11]. Both mouse and human IL4I1 degrade phenylalanine to produce the corresponding α -keto acid (phenylpyruvate), H₂O₂ and NH₃ (Figure 1). We also recently showed that the human form could catabolize arginine and identified specific SNPs and mutations in the IL4I1 coding sequence, which determine hyperactive or hypomorphic IL4I1 molecules [12]. These SNP may impact the immune response of affected individuals, since the IL4I1 enzyme exerts immunosuppressive functions. Indeed, our first work indicated that the *IL4I1* enzymatic activity, including the H_2O_2 produced, inhibits the proliferation of T cells, particularly effector/ memory (CD45RO⁺) T cells, and decreases the production of inflammatory chemokines and Th1 cytokines (IFNy and IL-2) (Figure 2) [11,13]. In humans, IL4I1 is expressed by mononuclear phagocytes, i.e. dendritic cells, monocytes and macrophages, after stimulation of the STAT1 and/or NF-κB pathways by IFNγ and Toll-like receptor ligands (Figure 2). In vitro, a much lower level of IL4I1 activity is also measured in peripheral blood B cells stimulated via the STAT6 and/or NK-KB pathways (e. g. with a cocktail of IL-4 and CD40 ligand which mimics stimuli received by germinal center B cells), but not in unstimulated peripheral blood T cells [13]. In vivo, IL4I1 expression is detected in macrophages and/or dendritic cells from human lymphoid tissues and inflammatory lesions including tumors.

Page 2 of 4



Figure 2: IL4I1 effects on effector T cell activation: The production of IL411 is induced in myeloid antigen-presenting cells (APC, i.e. dendritic cells, monocytes and macrophages) by proinflammatory mediators, such as pathogen-associated molecular patterns (PAMP), interleukin 1 (IL-1), type I interferons (IFN α/β) and IFN γ , which activate the STAT1 and/or NFkB pathways after binding to toll-like receptors, the IL-1 receptor, the IFN α/β receptor and the IFNy receptor, respectively. IL411 enzymatic activity depletes the local milieu in Phe. Phe depletion and increased concentration of H₂O₂ combined with medium basification by NH₃ can be toxic for bacteria, thus participating to an anti-infectious response. Phe depletion and H2O2 also affect effector CD4+ and CD8+ T lymphocyte activation, leading to diminished proliferation and effector functions. In the context of a tumor, the Th1 response may lead to IL411 induction in TAM, which can then inhibit the antitumor T cell response.

Through an analysis of a series of 315 tumors, representing 30 different histological types, we have shown that IL4I1 is strongly expressed by TAM and, in some cases, by neoplastic cells (mesothelioma and several B-cell lymphoma subtypes including follicular lymphoma, Hodgkin lymphoma, proliferation centers of chronic lymphoid leukemia) [14]. In the TAM, IL4I1 expression may be induced by the local production of IFNy by antitumor T cells. Indeed, we did not observe the expression of IL4I1 in macrophages stimulated in vitro by T helper type2 (Th2) cytokines such as IL-4 and IL-13 and in macrophages present in Th2 inflammatory lesions, such as schistosomiasis-associated granulomas [13]. This is in contrast to what was observed in mouse macrophages, where IL4I1 has been detected in M2 macrophages in vitro [15] and at the onset of inflammation resolution in mouse central nervous system lesions [16]. Differences between mice and humans which have also been observed in the expression of the arginine catabolizing enzyme, Arg1 [17], may explain these discrepancies. Interestingly, data obtained from a transcriptomic comparison, in mouse and human, of the three different skin migratory DC subpopulations with lymphoid organ resident DC (i. e. mouse lymph node $CD8\alpha^+$ DC and their proposed human homologues, blood BDCA3+ DC) indicated that IL4I1 was one of the genes most differentially expressed. In this work, migratory DC (which highly expressed IL4II) were suggested to dampen the immune response induced by resident DC (which poorly expressed IL4I1) [18].

IL4I1 is expressed by CD4⁺ T cells in some circumstances. While it is not detected in Th1 and Th2 subsets [7], the Annunziato's group has shown that the *IL4I1* mRNA is induced in Th17 cells by the ROR γ T master gene [19]. In this cell subset, *IL4I1* participates in controlling IL-2 production and cell proliferation in part by maintaining high

levels of the transcription factor Tob1, which prevents TCR-dependent cell-cycle progression (Figure 3) [20]. IL4I1 is also expressed by FoxP3+ Aiolos+ regulatory T cells and may modulate the pathogenicity of this cell population during inflammation-induced Th17 transdifferentiation (Figure 3) [21]. We recently confirmed that Th17 cells express active IL4I1 by detecting its enzymatic activity in ex-vivo sorted human Th17 populations (unpublished data) and in in vitro generated Th17 cells where its production appeared to be induced in an autocrine manner (Figure 3) [22]. Expression of IL4I1 in Th17 cells infiltrating some human cancers may contribute to their immunosuppressive and tumor-promoting properties [23]. Moreover, we have shown that IL4I1 biases naive CD4+ T cell differentiation towards that of FoxP₃⁺ regulatory T cells *in vitro* (Figure 3). Induction of regulatory T cells was associated with the inhibition of signaling via the mammalian target of rapamycin complex 1 (mTORC1) pathway, but did not impact mTORC2. Thus, local IL4I1 production may have consequences on the balance of effector versus suppressive T cells in the tumor microenvironment [13,22]. All these findings indicate that IL4I1 represents a key immunoregulatory molecule of different T-cell functions [24]. In addition, IL4I1 may play a role in the maturation of the T-dependent humoral response, since it is expressed by light zone germinal center B cells [25]. In these cells, it has been shown that IL4I1 is a target of mutations produced by the activation-induced cytidine deaminase [26]. This might have consequences on IL4I1 enzymatic activity and immunoregulatory properties. The expression of IL411 in innate lymphocyte subtypes has not yet been explored.



Figure 3: *IL4I1* effects on the balance between effector and regulatory CD4+ T cells: *IL4I1* production by dendritic cells during the priming of naïve CD4+ T cells favors regulatory T cell (Treg) differentiation. In the presence of Th17 inducing cytokines, *IL4I1* biaises Th17 differentiation towards cells that secrete less IL-17 and aquire the capacity to produce *IL4I1*. *IL4I1* production by Th17 blocks cell cycle progression, thus autolimiting the proliferation of this cell subset. Finally, circulating induced Treg cells (Ailos+ iTreg) can express *IL4I1* when converting into Th17 cells under inflammatory conditions. Local Phe depletion, which inhibits mTORC1 signaling, and H2O2 production participate in amplifying Treg cell induction from naïve T cells, but other yet undefined mechanisms are suspected. *IL4I1*-mediated immune escape of tumors may thus rely both on effector T cell inhibition and Treg cell induction.

To study the effect of *IL4I1* production in the context of a tumor, we have developed a preclinical murine melanoma model based on B16 cells transfected to express *IL4I1*. Following vaccination against a tumor antigen, we observed that *IL4I1* expressing-tumors developed

more frequently when compared to control tumors. This tumor escape was associated with the inhibition of proliferation and IFNy production of antitumor cytotoxic CD8⁺ T lymphocytes [27]. The minimal IL411 activities leading to this phenomenon were close to those detected in primary human melanoma, a tumor where the IL4I1 activity is supported exclusively by TAM. In line with these results, we recently observed that tumor growth was more easily controlled in the absence of the enzyme in IL4I1 KO mice [28] and unpublished data. Moreover, in the absence of IL4I1, the growth of primary tumors and metastases was delayed in a model of spontaneous melanoma induced by the Ret oncogene. This delay was accompanied by modifications of the primary tumor infiltrate, with an increase in Th1 cells and cytotoxic T cells and a diminution of polymorphonuclear myeloidderived suppressor cells [28]. Finally, in a HER2/neu positive mouse mammary carcinoma model, expression of Il4i1 in the microenvironment was associated with tumor recurrence [29,30]. In accordance with these preclinical data, Finak et al. identified IL4I1 as one of the 103 genes associated with pejorative prognosis, in a study of the genes expressed by the micro-dissected tumor stroma of human breast cancers. No other immunosuppressive enzyme-coding transcript was predictive of clinical evolution [31]. IL411 gene expression is also significantly up-regulated in laser-microbeam dissected triple negative breast cancer cells compared to normal ductal cells [32] and in fibroblasts present in precancerous mammary lesions [33], suggesting that IL4I1 may be expressed by breast cancer cells and tumor-associated fibroblasts in addition to TAM.

We provided evidence that the *in vitro* immunosuppressive effect of *IL4I1* is mediated through the production of H_2O_2 but, as for other amino-acid degrading enzymes, deprivation of an essential amino acid can also be involved [11,22]. *IL4I1* presents some ancestral antibacterial function, as other enzymes of the L-amino acid oxidase family, which are involved in anti-infectious defense in primitive species such as mollusks or reptiles. We have observed that this antibacterial effect is principally mediated by the toxicity of hydrogen peroxide, which can be amplified by medium basification due to ammonia production [34]. However, we did not observe such an effect of ammonia on T lymphocyte activation (unpublished data). We currently favor the hypothesis that different mechanisms cooperate to limit effector T lymphocyte functions and facilitate the expansion and differentiation of Tregs in tumors.

Stimulation of the antitumor immune response is gaining huge interest in clinical settings as spectacular successes have been obtained with the use of immune checkpoint inhibitors. However, not all patients experience long-lasting remissions and some cancer types respond poorly to this class of therapeutics. New immune targets are currently under clinical evaluation and others, such as *IL4I1*, may deserve careful exploration. Indeed, the data presented above argue that *IL4I1* could play an important role in dampening the anti-cancer immune response and understanding and blocking its activity may therefore be a promising strategy to improve current chemotherapy and immunotherapy approaches in cancer treatment. Moreover, the absences of overt autoimmune manifestations in *IL4I1* knock-out mice (unpublished data) suggests that such a strategy would not lead to major immune adverse effects in patients.

References

1. Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pages, C, Tosolini M, et al. (2006) Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 313: 1960-1964.

- 2. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. Cell 144: 646-674.
- Larkin J, Hodi FS, Wolchok JD (2015) Combined Nivolumab and Ipilimumab or Monotherapy in Untreated Melanoma. N Engl J Med 373: 1270-1271.
- Bingle L, Brown NJ, Lewis CE (2002) The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. J Pathol. 196: 254-265.
- Ugel S, De Sanctis F, Mandruzzato S, Bronte V (2015) Tumor-induced myeloid deviation: when myeloid-derived suppressor cells meet tumorassociated macrophages. J Clin Invest. 125: 3365-3376.
- Murray PJ (2016) Amino acid auxotrophy as a system of immunological control nodes. Nat Immunol 17: 132-139.
- Chu CC, Paul WE (1997) Fig1, an interleukin 4-induced mouse B cell gene isolated by cDNA representational difference analysis. Proc Natl Acad Sci U S A 94: 2507-2512.
- Chavan SS, Tian W, Hsueh K, Jawaheer D, Gregersen PK, et al. (2002) Characterization of the human homolog of the IL-4 induced gene-1 (Fig1). Biochim Biophys Acta 1576: 70-80.
- Wiemann S, Kolb-Kokocinski A, Poustka A (2005) Alternative premRNA processing regulates cell-type specific expression of the IL411 and NUP62 genes. BMC Biol 3: 16.
- Copie-Bergman C, Boulland ML, Dehoulle C, Möller P, Farcet JP, et al. (2003) Interleukin 4-induced gene 1 is activated in primary mediastinal large B-cell lymphoma. Blood 101: 2756-2761.
- 11. Boulland ML, Marquet J, Molinier-Frenkel V, Moller P, Guiter C, et al. Human IL4I1 is a secreted L-phenylalanine oxidase expressed by mature dendritic cells that inhibits T-lymphocyte proliferation. Blood 2007. 110: 220-227.
- 12. Molinier-Frenkel V, Mestivier D, Castellano F (2016) Alterations of the immunosuppressive IL4I1 enzyme activity induced by naturally occurring SNP/mutations. Genes Immun 17: 148-152.
- Marquet J, Lasoudris F, Cousin C, Puiffe M, Martin-Garcia, et al. (2010) Dichotomy between factors inducing the immunosuppressive enzyme IL4I1 in B lymphocytes and mononuclear phagocytes. Eur J Immunol 40: 2557-2568.
- Carbonnelle-Puscian A, Copie-Bergman C, Baia M, Martin-Garcia N, Allory Y, et al. (2009)The novel immunosuppressive enzyme IL4I1 is expressed by neoplastic cells of several B-cell lymphomas and by tumorassociated macrophages. Leukemia. 23: 952-960.
- 15. YueY, Huang W, Liang J, Guo J, Ji J, et al. (2015) IL4I1 Is a Novel Regulator of M2 Macrophage Polarization That Can Inhibit T Cell Activation via L-Tryptophan and Arginine Depletion and IL-10 Production. PLoS One10: e0142979.
- Psachoulia K, Chamberlain KA, Heo D, Davis SE, Paskus JD, et al. (2016) IL411 augments CNS remyelination and axonal protection by modulating T cell driven inflammation. Brain 139: 3121-3136.
- 17. Rath M, Müller I, Kropf P, Closs EI, Munder M (2014) Metabolism via Arginase or Nitric Oxide Synthase: Two Competing Arginine Pathways in Macrophages. Front Immunol 5: 532.
- Anandasabapathy N, Feder R, Mollah S, Tse SW, Longhi MP, et al. (2014) Classical Flt3L-dependent dendritic cells control immunity to protein vaccine. J Exp Med 211: 1875-1891.
- Santarlasci V, Maggi L, Mazzoni A, Capone M, Querci V, et al. (2014) IL-4-induced gene 1 maintains high Tob1 expression that contributes to TCR unresponsiveness in human T helper 17 cells. Eur J Immunol 44: 654-661.
- Santarlasci V, Maggi L, Capone M, Querci V, Beltrame L, et al. (2012) Rarity of human T helper 17 cells is due to retinoic acid orphan receptordependent mechanisms that limit their expansion. Immunity 36: 201-214.
- 21. Scarlata CM, Celse C, Pignon P, Ayyoub M, Valmori D (2014) Differential expression of the immunosuppressive enzyme IL411 induce Aiolos, but not natural Helios, FOXP3 Treg cells. Eur J Immunol 45: 474-479.

Page 4 of 4

- 22. Cousin C, Aubatin A, Le Gouvello S, Apetoh L, Castellano F, et al. (2015) The immunosuppressive enzyme IL4I promotes FoxP3(+) regulatory T lymphocyte differentiation. Eur J Immunol 45: 1772-1782.
- Martin F, Apetoh L, Ghiringhelli F (2012) Controversies on the role of Th17 in cancer: a TGF-Î²-dependent immunosuppressive activity? Trends Mol Med 18: 742-749.
- Romagnani S (2016) IL4I1: Key immunoregulator at a crossroads of divergent T-cell functions. Eur J Immunol 46: 2302-2305.
- Victora GD, Schwickert TA, Fooksman DR, Kamphorst AO, Meyer-Hermann M, et al. (2010) Germinal center dynamics revealed by multiphoton microscopy with a photoactivatable fluorescent reporter. Cell 143: 592-605.
- 26. Yamane A, Resch W, Kuo N, Kuchen S, Li Z, et al. (2011) Deepsequencing identification of the genomic targets of the cytidine deaminase AID and its cofactor RPA in B lymphocytes. Nat Immunol 12: 62-69.
- 27. Lasoudris F, Cousin C, Prevost-Blondel A, Martin-Garcia N, Abd-Alsamad I, et al. (2011) IL411: an inhibitor of the CD8å° antitumor T-cell response in vivo. Eur J Immunol 41: 1629-1638.
- Bod L, Lengagne R, Wrobel L, Ramspott J P, Kato M, et al. (2016) IL4induced gene 1 promotes tumor growth by shaping the immune microenvironment in a spontaneous model of melanoma. Oncoimmunol In press.

- Worschech A, Kmieciak M, Knutson KL, Bear HD, Szalay A A, et al. (2008) Signatures associated with rejection or recurrence in HER-2/neupositive mammary tumors. Cancer Res 68: 2436-2446.
- 30. Pavlides S, Tsirigos A, Vera I, Flomenberg N, Frank PG, et al. (2010) Transcriptional evidence for the "Reverse Warburg Effect" in human breast cancer tumor stroma and metastasis: similarities with oxidative stress, inflammation, Alzheimer's disease, and "Neuron-Glia Metabolic Coupling". Aging 2: 185-199.
- Finak G, Bertos N, Pepin F, Sadekova S, Souleimanova M, et al. (2008) Stromal gene expression predicts clinical outcome in breast cancer. Nat Med 14: 518-527.
- 32. Komatsu M, Yoshimaru T, MatsuoT, Kiyotani K, Miyoshi Y, et al. (2013) Molecular features of triple negative breast cancer cells by genome-wide gene expression profiling analysis. Int J Oncol 42: 478-506.
- 33. Lisanti M P, Tsirigos A, Pavlides S, Reeves KJ, Peiris-Pages M, et al. (2014) JNK1 stress signaling is hyper-activated in high breast density and the tumor stroma: connecting fibrosis, inflammation, and stemness for cancer prevention. Cell Cycle 13: 580-599.
- Puiffe ML, Lachaise I, Molinier-Frenkel V, Castellano F (2013) Antibacterial properties of the mammalian L-amino acid oxidase IL4I1. PLoS One 8: e54589.