Global Toxicology 2020: If atrazine has a potential to impair a function of bovine cervix during periovulation period of oestrous cycle, *in vitro*? Michał H Wróbel, Institute of Animal Reproduction and Food Research, Poland

Abstract

Atrazine, which belongs to triazine class of herbicides, remains a staple of the American and Indian agriculture, while is subjected to strict regulation that effectively prevents its use in Europe. It was previously shown that atrazine increased the secretion of ovarian oxytocin (OT), which is a potent uterotonic agent. However, atrazine directly inhibit the myometrial contractions in cows. Hence, the aim of this study was to determine its effect on the motoric function of bovine cervix.

Cervical strips or cells from cows at 18-20 days of oestrous cycle were incubated (24-72h) with atrazine (10 ng/ml). The used dose of atrazine, which was chosen according to the our previous studies, has exerted any cytotoxic effect. However, it increased the force of cervical contractions and it has not affected the level of myosin light chain kinase (MLCK). While atrazine decreased the secretion of PGE2, amount of gap junction proteins (GAPs) and the second messengers IP3), which are involved (DAG, in transmission of signal to contraction.

Atrazine showed direct stimulation of the force of cervical contractions and inhibition of cellular signalling. Hence, it has potential to impair the function of cervix, which can be followed by failures in beginning of gestation.

INTRODUCTION

From the beginning of dairy animals taming a large number of years prior, man has had the option to perceive certain conduct and physical changes identified with the conceptive condition of steers (1,2). Among ruminants, for example, bovines, just as in

different species, during the oestrus stage a to some degree translucent and generally clingy fluid substance can be seen to bounteously come out from the female conceptive tract, one of the signs that has been perceived, from antiquated occasions, as the start of sexual receptivity (heat) (1,2). Such emission, known as cervical bodily fluid, applies a few physiological capacities that are basic for the improvement of the regenerative procedure in the cow, equivalent to in different creatures in which this liquid is delivered.

At the point when a drop of cervical bodily fluid gathered at oestrus is saved on a straightforward surface and permitted to dry at room temperature, the bodily fluid will in general solidify in profoundly masterminded geometric examples, portrayed basically by arborescent morphologies, among different courses of action (1,3-7). Evaluation of the properties of cervical bodily fluid all through the oestrous cycle shows that the degree of the crystallization wonder arrives at a most extreme at oestrus in contrast with some other phase of the cycle (4). This is basically because of brought levels of oestrogens up in this phase which are applying their consequences for the cervical mucosa, and, consequently, changing the bodily fluid highlights. This clarifies why the evaluation of bodily fluid crystallization has been recommended by certain specialists as a valuable apparatus to decide the beginning of sexual receptivity in dairy animals (1,8,9) and different creatures (10,11).

The target of this audit article is to talk about the primary parts of the wonder of bodily fluid crystallization, with exceptional enthusiasm for the attributes of the crystalline examples saw in cow-like cervical discharge.

BOVINE CERVICAL MUCUS

Cervical bodily fluid is created by bodily fluid discharging cells that line the sections and overlap looking like 'dazzle finished tombs,' present in the cervical epithelium (12). Emitted all the more bounteously at oestrus, bodily fluid volume can reach up to 100 mL. Concerning concoction structure, cervical bodily fluid is a hydrogel with 92 to 95% of water content (13) and includes solvent and non-dissolvable substances (14,15). Among the solvent substances are proteins, for example, lactoferrin (16), immunoglobulins, a few chemicals, e.g., glucosidases and lattice metalloproteases (17,18), various low-subatomic mass mixes, for example, starches (e.g., fructose and glucose) (13,19,20), amino acids, lipids, for example, cholesterol (13,20), and inorganic particles (electrolytes), the most significant being Na+, K+, Ca2+ and Cl-(21). Then again, the non-dissolvable portion comprises of high-atomic mass glycoproteins known as mucins (15,22-24), primarily those named emitted gel-shaping mucins (25). These are profoundly glycosylated proteins and likely establish the principle factor answerable for the rheological properties of bodily fluid, for example, its variable versatility, consistency and spinnbarkeit, among others (26).

The repetitive varieties in the degrees of oestrogens and progesterone markedly affect ox-like cervical bodily fluid. When all is said in done, these hormones apply their impact by straightforwardly following up on discharging endocervical cells, basically by means of systems interceded by traditional steroid receptors (27); as an outcome, the sythesis, the physicochemical and basic properties, and the rheological characteristics of the cervical emission are changed (28). With respect to, it has been demonstrated that bodily fluid water content shifts along the cycle, expanding at oestrus (29,30), due essentially to the ascent in oestradiol levels saw in this stage. Additionally, mucin types are differentially communicated during the phases of the oestrous cycle (12,24), a change likewise

identified with vacillations in sex steroid hormones.

FUNCTIONS OF BOVINE CERVICAL MUCUS

Bovine cervical bodily fluid has a few significant capacities in the regenerative procedure, among which are:

The cervical discharge secures the cow-like regenerative tract by keeping up the epithelial surfaces soggy and greased up. This is because of the significant level of hydration that portrays this gel (15), since mucins are fit for restricting huge volumes of water (22,31).

Cervical bodily fluid partakes in sperm determination and transport, being the main medium spermatozoa must experience when climbing to the site of treatment (15,32,33). During the periovulatory period, bodily fluid discharge increments, turning out to be not so much gooey but rather more hydrated, encouraging the rising of spermatozoa (15,32,33). Likewise, in this period, the bodily fluid structure would encourage the development of ordinary spermatozoa and rising of restrain the gametes with morphological modifications, going about as a specific channel (33). Then again, in the luteal stage, the sum and hydration of the emitted bodily fluid reductions while its increments. forestalling thickness spermatozoa relocation. As per Becher et al. (34), this capacity of ox-like cervical bodily 'channel' fluid to spermatozoa has demonstrated to be helpful in the regions of human andrology and gynecology with respect to sperm infiltration tests, during which cow-like bodily fluid was here and there used to make up for the little volume of cervical bodily fluid that can be acquired from a lady.

The cervical discharge establishes a safe obstruction that hinders the climb and colonization of microorganisms, since a portion of the mixes present in bodily fluid can restrain the infiltration and expansion of organisms (35).

During pregnancy, cervical bodily fluid shields the uterus from natural poisonous specialists, since during this period bodily fluid structures an exceptionally gooey obstruction known as cervical bodily fluid attachment (31,36).

Various substances have been recognized in the liquids of a dairy animals' conceptive tract, among which there are sex steroid hormones (37). These are additionally present in cervical bodily fluid and most likely tweak acrosome response (acrosomal the exocytosis), as it has been proposed for human cervical bodily fluid (38-41). In bovines, this thought is additionally bolstered by proof acquired by utilizing examining electron microscopy when contemplating cervical areas in follicular stage, in which spermatozoa with flawless acrosomal films in some bodily fluid filled luminal locales were watched (12). Be that as it may, further investigations are expected to clarify the job applied by sex steroid hormones present in ox-like cervical bodily fluid on acrosomal exocytosis.

CERVICAL CRYSTALLIZATION

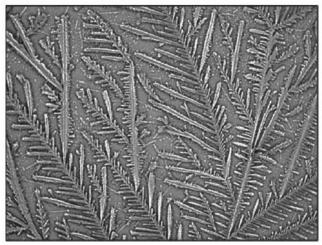
MUCUS

All in all terms, crystallization can be characterized as the procedure through which a part of a fluid arrangement changes to its strong stage, tending to isolate from the arrangement and to encourage as gems. Crystallization establishes a way to arrive at an increasingly steady, lower vitality state from a metastable arrangement by lessening the solute fixation (42). Crystallization is delivered by atomic accumulation prompting arrangement crystalline the of cores (nucleation), with the ensuing development of those cores. Accordingly, nucleation is the antecedent of crystalline development.

The procedure of crystallization isn't selective to cervical emission; it is available in various other organic discharges, for instance, human and cow-like salivation (43). Discharges ready to take shape are portrayed by containing mucoproteins (e.g., mucins) or natural mixes and electrolytes. other particularly salts, for example, NaCl, KCl and CaCl2 (13,44). Actually, NaCl is the principle salt found in cervical bodily fluid, furnishing the bodily fluid with ionic quality (21). When all is said in done, cervical bodily fluid crystallization has been concentrated by

spreading bodily fluid example drops onto a glass slide so that, in the wake of drying at room temperature, a smear or film shapes on the slide. This can be seen without recoloring by utilizing a standard light magnifying instrument (1,4,5,45). The first to write about crystallization cervical emission was Papanicolaou (1946), who concentrated on the plant like courses of action discernible on ladies' bodily fluid, proposing that this marvel could be utilized as an indicator of ovulation (10). Because of the crystalline shapes that were watched, from that point on, cervical bodily fluid crystallization has additionally been called an 'arborization' or 'ferning wonder' (Figure 1). Garm and Skjerven (46) examined crystallization in the cervical bodily fluid of cows, finding bounteous fernlike precious stones during the follicular stage, which vanished during the luteal stage and were non-noticeable in the beginning periods of pregnancy. Afterward, a few scientists confirmed that the most elevated arborization happened at the beginning of, or during, oestrus (1,47,48). In such manner, Abusineina (1) expressed that the investigation of cervical bodily fluid according to the nearness or nonappearance of crystallization, and the kind of crystallization watched, is a pointer of the phase of oestrous cycle and the day of ovulation; thusly, it is helpful to affirm discoveries iust for clinical as test applications. MacDonald (29) revealed that in the periovulatory period, when spermatozoa can move through bodily fluid, the extent of water content is over 98%, and the salt substance in the dry buildup is over half. As the extent of salts in the dry buildup begins to diminish, so does the water content. This change prompts а reduction in the arborizations saw in the dried bodily fluid example. In the bodily fluid acquired from pregnant bovines, the water content is 90% arborization can be and no watched. Comparable to this, Noonan et al. (4) saw a reverse connection between the degree of arborization and the substance of drv issue in bodily fluid examples. cervical The convergence of dry issue in cervical bodily fluid arrived at least at oestrus and a top at mid-cycle, while bodily fluid ferning showed up at oestrus to a more prominent degree than at some other phase of the oestrous cycle.

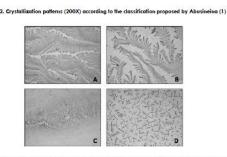
Figure 1. Typical morphology of bovine cervical mucus at oestrus observed under light microscopy (200X). An arborescent crystalline pattern resembling a fern frond can be observed



Likewise, concerning different properties of cervical bodily fluid, varieties in the event of crystallization during the oestrous cycle are similarly because of the changing degrees of sex steroid hormones. As a rule, oestrogens are considered to advance crystallization, while progesterone diminishes it (13). In such manner, it has been suggested that the higher event of arborizations at oestrus relies upon estrogen strength during the follicular stage (49). Oestrogens would cause an expansion in the ferning marvel through instruments that invigorate the electrolyte digestion in the cervical epithelium (50,51). Then again, in the luteal stage. expanded degrees of progesterone would neutralize the impacts of oestrogens on the cervix, clarifying the abatement in arborizations (50,52), a reality that is in concurrence with the inhibitor impact on precious stones recently revealed for this hormone (53). The impact applied by these sex steroids corresponds with the perception made by Elstein (54), who expresses that, among the numerous qualities of cervical bodily fluid, arborization is, no ifs, ands or buts, one of the most delicate to varieties in the degrees of sex steroids. These days, it is notable that crystallization comprises a valuable property for examining both cervical bodily fluid and the conceptive cycle (5,55). Then again, it is likewise worth referencing that the crystallization of steers salivation experiences changes during the oestrous cycle. and consequently the examination of such crystallization can help in the finding of early pregnancy (43).

CRYSTALLINE PATTERNS FOUND IN BOVINE CERVICAL MUCUS

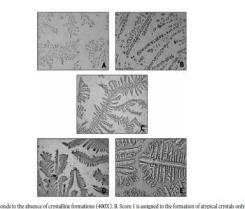
As far as anyone is concerned, the main model to group crystallization examples of cow-like cervical bodily fluid was proposed by Abusineina (1), who partitioned the watched courses of action into three kinds. Type A relates to the crystallization saw when the bodily fluid is translucent, acellular, flexible and effortlessly acquired from the cervix. Under a light magnifying lens, Type A crystallization is portrayed by the nearness of a long, dainty stem (principle hub), which might be straight, waving or bended. From such a stem, very much characterized venations of variable length project with little subvenations (Figure 2A). This kind of crystallization would be related with ovulation and created as an outcome of high estrogen levels (1). Type B crystallization relates to that saw when bodily fluid is semitranslucent, flexible and handily got from the cervix. At the point when seen under a light magnifying lens, this sort is closest fit as a fiddle to a plant frond (1). Venations and subvenations are all around characterized and simple to watch (Figure 2B). Type C compares to the crystallization of dark bodily fluid, confirming cellularity and being hard to get from the cervix. At the point when watched utilizing light microscopy, Type C crystallization is unpredictable and its plant like morphology is atypical. The focal hub is short, with or without venations and subvenations, which are unpredictable (1) (Figure 2C). Some scatter direct crystalline examples can be found, either cruciform (Figure 2D) or stellate.



A. Type A Crystallization is characterized by the presence of a long, thin 'stem' (main axis), which may be straight, waving or curved. From such a stem, well-defined venations of variable length protrude, some of them showing tiny subvenations. B. Type B Crystallization resembles the shape of a fern frond. Venations and subvenations are well-defined and easy to observe. C. Type C Crystallization is irregular and its fern-like morphology is atypical. The central axis is short, with or without venations and subvenations, which are irregular. Type C coasionally presents cruciform arrangements (D), among other shapes.

Another model for the order of cow-like cervical bodily fluid at oestrus was proposed by Bishnoi et al. (56) and actualized by

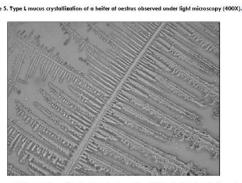
Tsiligianni et al. (8), in light of a subjective scale (extending from 0 to 4). Score 0 compares to the nonattendance of crystalline arrangements (Figure 3A). Score 1 is appointed to the development of atypical precious stones just (Figure 3B). Score 2 is allocated when numerous atypical and a couple of regular plant like precious stones are watched (Figure 3C). Score 3 speaks to the development of numerous average plant like precious stones and a couple of atypical gems (Figure 3D). At long last, score 4 is given to designs demonstrating the average greenery frond precious stone game plans (Figure 3E). ms according to the classification proposed by Bishnoi et al. (56)



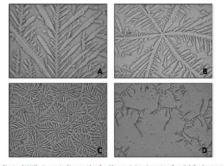
A. score v corresponds to me assence or crystalline tormations (400X). In Score 1 is assigned to the formation or atypical crystal only (400X). C. Score 2 corresponds to many atypical and some typical ferm-filte crystals (200X). D. Score 3 represents the formation of many pipcial ferm frond crystals (200X). D. Score 4 is given to patterns showing the typical ferm frond crystal arrangement (400X).

A notable order model for the crystallization of cervical bodily fluid of the periovulatory period was accounted for by Odeblad (57) and later approved by different examinations (58,59). At first proposed for ladies' cervical discharge (57), it is intriguing and worth referencing that, when considering ox-like cervical bodily fluid as indicated by the arrangement proposed by Odeblad, geometrical crystallizations fundamentally the same as those acquired for human cervical bodily fluid are watched (5,7). The sorts (and subtypes) proposed by Odeblad, and detectable in bovines, are: Type S Crystallization: its morphology takes after straight lines that tend towards an equal game plan (Figure 4). Type L Crystallization: it is portrayed by a palm leave or greenery frond morphology, with an all around characterized focal hub and 90° expanding (Figure 5), like the examples saw in human cervical bodily fluid (57,59,60). Type P Crystallization: gathering a few crystalline subtypes, this example has 60° edge branchings beginning from the fundamental hub. It is partitioned

into five subtypes, however those by and large saw in cow-like cervical bodily fluid are four: Subtype P2, comprising of a very much characterized principle stem (hub), from which branchings project to the two sides, framing 60° edges with the fundamental stem. This crystalline example is clearly plant like (Figure 6A). Subtype P6B has an alluring geometry, looking like a star, with a focal core from which six very much characterized tomahawks jut (7). Every hub frames a 60° edge with the following, and branchings of variable length start from every hub (Figure 6B). By and large, this subtype has been found to frame to some degree bigger crystalline units than different subtypes of P bodily fluid (58); likewise, subtype P6B in people would be connected to the fruitfulness top (61). Subtype Pa has a crystallization place from which numerous branchings illuminate every which way (Figure 6C). Ultimately, another subtype compares to bodily fluid Pt, which doesn't have such a precise course of action like the previously mentioned subtypes; the precious stones have all the earmarks of being more scatter and not generally joined (Figure 6D), as saw in ladies (58, 59).



This crystalline pattern, resembling a palm leave or fern frond, has a structure with a straight main axis and ramifications protruding at a 90 angle from which small indentations can originate, again at a 90° angle. Figure 6. Subtype P crystallizations identified in the cervical mucus of heiters at oeshus under light microscopy



A Subtype IP crystallization (400X): characterized by an evident tem-like morphology. It consists of a veil-detined main aix, from which branchings protude on both sides, froming 60v angles with the main aix is 1. Subtype F0_crystallization (400X): a dar-like geometry, with a central nucleus from which six well-defined axes protrude at a 60° angle with each other. From each of these axes, branchings of variable length originate. C. Subtype Pa_crystallization (200X): units of this crystalline pattern are commonly found close to one another. In each, a central point can be recognized from which branches originate in all directions. D. Subtype P1 crystallization (400X): this a main axis from which variable length branches originate. Both the main axis and the branches seem to be constituted by somewhat discontinuous crystal patterns.

Correspondingly, it merits referencing that, when seen under light microscopy, cow-like

cervical bodily fluid likewise shows some crystalline game plans that can't be acceptably sorted into any of the recently depicted models (Figure 7). At long last, certain arboriform crystallizations of yearling cervical bodily fluid have been accounted for to show a fractal-like association, i.e., crystalline structures are involved littler parts that take after the entire in a littler scope (6). These fractal designs have additionally been proposed for human bodily fluid (62), despite the fact that their organic criticalness is yet to be clarified.

Finishing up REMARKS

During the oestrous cycle, and particularly at oestrus, it is conceivable to distinguish a few geometric courses of action for the crystalline examples of ox-like cervical bodily fluid; this reality has made it conceivable to propose the portrayed arrangements. recently The explanation behind the presence of such kinds of crystallization has not been totally clarified, yet thinking about that ox-like cervical epithelium could be involved diverse secretory districts (12,63), those crystalline examples would be aftereffect of the differential impact applied by raised degrees of oestrogens at oestrus on such areas. Therefore, cervical bodily fluid is most likely a heterogeneous element shaped by the admixture of a few subtypes of emission (59,64,65), with extents changing in the periovulatory period and, less significantly, during different phases of the cycle. This reality would clarify the presence of various morphological sorts (and subtypes) of bodily fluid crystallization; other conceivable fundamental causes are varieties in salt of substance (because alterations in electrolyte digestion at the degree of the cervix) and water content, just as the plan and kind of mucin present in cervical bodily fluid because of changes in levels of sex steroid hormones.

The investigation of the crystallizations present in cow-like cervical bodily fluid at oestrus and different stages could prompt a more profound comprehension of ox-like conceptive physiology, both in physiological and pathophysiological conditions. Having the option to relate one explicit sort of cervical bodily fluid crystallization with a dairy animals' richness pinnacle could be particularly applicable conceptive in administration. Also, partner a specific example of crystallization with oestrogens and progesterone levels could be vital in the field of veterinary medication. In such manner, it is realized that changes in cervical bodily fluid ultrastructure are connected to fruitfulness issues (absence of pregnancy) in cows (66,67), as it has additionally been accounted for in ladies (60). Among ladies with fruitfulness issues because of endocrinemetabolic turmoil known as polycystic ovary disorder (PCOS), an adjustment in cervical bodily fluid ultrastructure has been watched along with alterations in the examples of bodily fluid crystallization (60); moreover, modified rheological properties (e.g., changes in flexibility) have likewise been accounted for in cervico-vaginal discharges of PCOS ladies (68). Thinking about the previous, it is conceivable that adjustments in bodily fluid ultrastructure and rheology are likewise joined by changes in crystallization designs in certain dairy animals with fruitfulness issues optional to endocrine aggravations.

At last, further exploration around there should concentrate on distinguishing the physiological significance of the distinctive crystalline examples of cow-like cervical bodily fluid at oestrus, just as on recognizing the biochemical instruments activating changes in electrolyte digestion, in bodily fluid hydration, and in mucin articulation at the cervix, all of which could clarify the watched varieties in crystallization designs. It is additionally of significance to clarify the component through which changes in sex levels impact the sorts steroid of crystallizations saw among solid dairy animals and in those experiencing conceptive scatters.

References

1. Abusineina ME. A study of the fern-like crystalline patterns of the cervical and vaginal mucus of cattle. Vet Rec. 1962;74:619-21.

2. Pommerenke WT. Some biochemical aspects of the cervical secretions. Ann N Y Acad Sci. 1962;97:581-90.

3. Lamond DR, Shanahan AG. Chemical changes in cervical mucus from normal and ovariectomized cows treated with hormones. Biol Reprod. 1969;1(4):335-43.

4. Noonan JJ, Schultze AB, Ellington EF. Changes in bovine cervical and vaginal mucus during the estrous cycle and early pregnancy. J Anim Sci. 1975;41(4):1084-9.

5. Cortés ME. Morphological and ultrastructural characterization of different types of bovine cervical mucus using light and scanning electron microscopy [tesis doctoral]. [Santiago de Chile]: Pontificia Universidad Católica de Chile; 2012.

6. Cortés ME, Hauyón R, Vigil P, González F. Evidence of fractality in a pattern of crystallization of bovine cervical mucus obtained at oestrus. Int J Morphol. 2012;30(4):1461-5.

7. Cortés ME, González F, Hauyón R, Vigil P. Highly symmetrical crystallization in six rectilinear and well-defined axes found in bovine cervical mucus obtained at oestrus: a finding. Rev Fac Med Vet Zoot. 2014;61(2):164-70.

8. Tsiligianni T, Karagiannidis A, Brikas P, Saratsis P. Relationship between certain physical properties of cervical mucus and fertility in cows. Deutsche Tierárztliche Wochenschrift. 2000;107:28-31. [Links]

9. Tsiligianni T, Amiridis GS, Dovolou E, Menegatos I, Chadio S, Rizos D, Gutiérrez-Adán A. Association between physical properties of cervical mucus and ovulation rate in superovulated cows. Can J Vet Res. 2011;75(4):248-53.

10. Papanicolaou GN. A general survey of the vaginal smear and its use in research and diagnosis. Am J Obstet Gynecol. 1946;51:316-28.

11. McDonald MF, Raeside JI. Use of the cervical mucus smear in assessing ovarian activity in the ewe. Nature. 1956;178(4548):1472-3.

12. Mullins JK, Saacke RG. Study of the functional anatomy of bovine cervical mucosa with special reference to mucus secretion and sperm transport. Anat Rec. 1989;225(2):106-17.

13. Tsiligianni T, Karagiannidis A, Brikas P, Saratsis P. Chemical properties of bovine cervical mucus during normal estrus and estrus induced by progesterone and/or PGF2a. Theriogenology. 2001;56(1):41-50.

14. Schumacher GFB. Soluble proteins in cervical mucus. En: Blandau RJ, Moghissi KS (eds). The biology of the cervix. Chicago: University of Chicago Press; 1973. p. 201-33.

15. Rutllant J, López-Béjar M, López-Gatius F. Ultrastructural and rheological properties of bovine vaginal fluid and its relation to sperm motility and fertilization: a review. Reprod Domest Anim. 2005;40(2):79-86.

16. Rao KSPB, Roberts TK, Masson PL, Heremans JF. Lactoferrin, a major soluble protein of bovine oestrous cervical mucus. J Reprod Fertil. 1973;32(1):89-92.

17. Tsiligianni T, Karagiannidis A, Saratsis P, Brikas P. Enzyme activity in bovine cervical mucus during spontaneous and induced estrus. Can J Vet Res. 2003;67(3):189-93.

18. Kim SH, Baek JS, Lee HJ, Min KS, Lee DH, Yoon JT. Detection of matrix metalloprotease-9 and analysis of protein patterns in bovine vaginal mucus during estrus and pregnancy. J Embr Transfer. 2012;27:93-100.

19. el-Naggar MA, Baksai-Horváth E. The sugar content of the cervico-vaginal mucus of cattle during the sexual cycle, with special reference to fructose. Acta Vet Acad Sci Hung. 1971;21(1):15-20.

20. Zaaijer D, van der Horst CJG. Cyclic changes in hormones, carbohydrates and indole metabolism in cervical mucus of normal, fertilizing cows and the relationship with non-fertility. Cytobios. 1983;37(146):113-27.

21. Sato M, Nihei A, Ohta M, Masaki J. Changes in sodium, potassium and chloride concentrations of bovine cervical mucus during the time of estrus induced by prostaglandin F2a analogue. Tohoku J Agricult Res. 1981;32(4):40-9.

22. Gibbons RA. Chemical properties of two mucoids from bovine cervical mucin. Biochem J. 1959;73(2):209-17.

23. Gibbons RA, Glover FA. The physicochemical properties of two mucoids from bovine cervical mucin. Biochem J. 1959;73(2):217-25.

24. Pluta K, Irwin JA, Dolphin C, Richardson L, Fitzpatrick E, Gallagher ME, et al. Glycoproteins and glycosidases of the cervix

during the periestrous period in cattle. J Anim Sci. 2011;89(12):4032-42.

25. Gipson IK. Human endocervical mucins. Ernst Schering Research Foundation Workshop. 2005; 52:219-44.

26. Wolf DP, Sokoloski J, Khan MA, Litt M. Human cervical mucus. III. Isolation and characterization of rheologically active mucin. Fertil Steril. 1977;28(1):53-8.

27. Nicosia SV. Physiology of cervical mucus production. Semin Reprod Med. 1986;4(4):313-21.

28. Sharma V, Prasad S, Gupta HP. Studies on physical and rheological properties of cervico-vaginal mucus during early pregnancy in buffaloes (*Bubalus bubalis*). Vet World. 2013;6(8):508-11.

29. MacDonald RR. Cyclic changes in cervical mucus. 2. The role of saline. J Obstet Gynaecol Br Commonw. 1969;76(12):1094-9.

30. Merilan CP. Evaluation of bovine cervical mucus during estrous cycle by nuclear magnetic resonance. J Dairy Sci. 1983;66(5):1184-8.

31. Becher N, Waldorf KA, Hein M, Uldbjerg N. The cervical mucus plug: structured review of the literature. Acta Obstet Gynecol Scand. 2009;88(5):502-13.

32. Barros C, Vigil P, Herrera E, Pérez A, Guadarrama A, Bustos-Obregón E. *In vitro* interaction between human spermatozoa and human cervical mucus. Microsc Electrón Biol Cel. 1983;7:13-8.

33. Barros C, Vigil P, Herrera E, Argüello B, Walker R. Selection of morphologically abnormal sperm by human cervical mucus. Arch Androl.1984;12 Suppl:95-107.

34. Becher AC, Failing K, Kauffold J, Wehrend A. Establishment of a practical sperm penetration test for bovine semen. Tierárztliche Praxis Grofitiere. 2013;41(5):297-303.

35. Brownlie J, Hibbitt KG. Antimicrobial proteins isolated from bovine cervical mucus. J Reprod Fertil. 1972;29(3):337-47.

36. Marshall FHA, Hammond J. Fertility and animal breeding. Bull Ministry of Agriculture and Fisheries. 1937;39:1-52.

37. Short RV. Steroid concentrations in normal follicular fluid and ovarian cyst fluid from cows. J Reprod Fertil. 1962;4:27-45.

38. Vigil P, Toro A, Godoy A. Physiological action of oestradiol on the acrosome reaction in human spermatozoa. Andrologia. 2008;40(3):146-51.

39. Vigil P, Orellana RF, Cortés ME. Modulation of spermatozoon acrosome reaction. Biolog Res. 2011;44(2):151-9.

40. Vigil P, Cortés ME. El misterio del inicio de la vida humana. Diálogos. 2011;1:22-5.

41. Vigil P. La fertilidad de la pareja humana. 4ta ed. Santiago de Chile: Ediciones Universidad Católica; 2013. p. 76-77.

42. Weber PC. Physical principles of protein crystallization. Adv Protein Chem. 1991;41:1-36.

43. Skalova I, Fedorova T, Brandlova K. Saliva crystallization in cattle: new possibility for early pregnancy diagnosis? Agric Tropica et Subtrop. 2013;46(3):102-4.

44. Rydberg E. Observation on the crystallization of the cervical mucus. Acta Obstet Gynecol Scand. 1948;28(2):172-87.

45. Odeblad E. The spread out technique. Advantages, pitfalls and biological interpretation. Actas IV Symposium Internacional de Métodos Naturales. Barcelona; 1995. p. 295-303.

46. Garm O, Skjerven O. Undersokelse av cervikalslim for diagnose av tidlig drektighet og endokrint betingede forstyrrelser av seksualcyklus hos husdyr. Nordisk Veterinærmedicin. 1952;4:1098-103.

47. Bone JF. Crystallization patterns in vaginal and cervical mucus smears as related to bovine ovarian activity and pregnancy. Am J Vet Res. 1954;15(57):542-7.

48. Alliston CW, Patterson TB, Ulberg LC. Crystallization patterns of cervical mucus as related to estrus in beef cattle. J Animal Sci. 1958;17(2):322-5.

49. Ghannam SA, Sorensen AM. Early pregnancy diagnosis in the bovine. J Dairy Sci. 1967;50(4):562-7.

50. Roland M. A simple test for the determination of ovulation, estrogen activity, and early pregnancy using the cervical mucus secretion. Am J Obstet Gynecol. 1952;63(1):81-9.

51. Zondek B. Some problems related to ovarian function and to pregnancy. Rec Prog Hormone Res. 1954;10:395-423.

52. Forman I. Cervical mucus arborization; aid in ovulation timing. Obstet Gynecol. 1956;8(3):287-92.

53. Campos da Paz A. The crystallization test as a guide to the treatment of cervical hostility. Fertil Steril. 1953;4(2):137-48.

54. Elstein M. Cervical mucus: Its physiological role and clinical significance. Br Med Bull. 1978;34:83-8.

55. Silaban NL, Setiatin dan Sutopo ET. Tipologi ferning sapi jawa brebes betina berdasarkan periode berahi. Animal Agriculture J. 2012;1(1):777-88.

56. Bishnoi BL, Vyas KK, Dwaraknath PK. Note on spinnbarkeit and crystallization pattern of bovine cervical mucus during oestrus. Indian J Anim Sci. 1982;52(6):438-40.

57. Odeblad E. The discovery of different types of cervical mucus and the Billings ovulation method. Bulletin of the Natural Family Planning Council of Victoria. 1994;21:1-34.

58. Menárguez M. Caracterización morfológica de diversos tipos de moco cervical humano mediante microscopía de luz y microscopía electrónica de barrido [tesis doctoral]. [Murcia]: Universidad de Murcia; 2012.

59. Menárguez M, Pastor LM, Odeblad E. Morphological characterization of different human cervical mucus types using light and scanning electron microscopy. Hum Reprod. 2003;18(9):1782-9.

60. Vigil P, Cortés ME, Zúñiga A, Riquelme J, Ceric F. Scanning electron and light microscopy study of the cervical mucus in women with polycystic ovary syndrome. J Electron Microsc. 2009;58(1):21-7.

61. Odeblad E, Ingelman-Sundberg A, Menárguez M, Temprano H, Pouyanmehr S, Vigil P, et al. Types of cervical secretion. Actas VIII Symposium Internacional sobre Regulación Natural de la Fertilidad: Aplicaciones a la Salud Reproductiva. Leioa, España; 2006;1-15.

62. Yang G, Yang Z, Zhou H, Huang T. Fractal analysis of the fernleaf crystallization of cervical mucus. J Yunnan Univ National. 2011;20(1):75-8.

63. Heydon RA, Adams NR. Comparative morphology and mucus histochemistry of the ruminant cervix: differences between crypt and surface epithelium. Biol Reprod. 1979;21(3):557-62.

64. Odeblad E. The functional structure of human cervical mucus. Acta Obstet Gynecol Scand. 1968;47(S1):57-79.

65. Richardson L, Hanrahan JP, O'Hara L, Donovan A, Fair S, O'Sullivan M, et al. Ewe breed differences in fertility after cervical AI with frozen-thawed semen and associated differences in sperm penetration and physicochemical properties of cervical mucus. Anim Reprod Sci. 2011;129(1-2):37-43.

66. López-Gatius F, Labérnia J, Santolaria P, Rutllant J, López-Béjar M. The relationship of the rheological behavior of the vaginal fluid at the time of insemination to the pregnancy rate in dairy cows. Theriogenology. 1997;48(5):865-71.

67. Rutllant J, López-Béjar M, Santolaria P, Yániz J, López-Gatius F. Rheological and ultrastructural properties of bovine vaginal fluid obtained at oestrus. J Anat. 2002;201(1):53-60.

68. Shamim N, Usala SJ, Biggs WC, McKenna GB. The elasticity of cervicalvaginal secretions is abnormal in polycystic ovary syndrome: Case report of five PCOS women. Indian J Endocrinol Metab. 2012;16(6):1019-21.

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