

Functionalized Nanoparticles for Drug Delivery, One- and Two-photon Photodynamic Therapy as a Promising Treatment of Retinoblastoma

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Abstract

Retinoblastoma is a rare cancer triggered by genetic mutation that forms in the eyes of children. In industrialized countries, 95% of patients are cured by chemotherapy and conservative treatments. However these treatments can increase the risk of secondary tumors in patients with a constitutional alteration of the retinoblastoma gene *Rb1*. Photodynamic therapy (PDT) represents a therapeutic approach and may reduce the incidence of secondary tumors. PDT is an established cancer treatment based on the light activation of a photosensitizing agent thus generating cytotoxic reactive oxygen species that cause cellular damage. We focused on mesoporous silica nanoparticles (MSN) for one-photon excited PDT combined with drug delivery and carbohydrate targeting applied on retinoblastoma. We demonstrated that bitherapy (camptothecin delivery and PDT) performed with MSN was efficient in inducing retinoblastoma cell death. Alternatively, MSN designed for two-photon excited PDT were also studied and irradiation in near-infrared at low fluency efficiently killed retinoblastoma cancer cells. These data provide new evidences of the potential of functionalized and targeted MSN for treatment of retinoblastoma and could lead to propose a non-invasive therapy with reduced side effects.

Keywords: Mesoporous silica nanoparticles; Two-photon; Photodynamic therapy; Drug delivery; Mannose; Galactose; Targeting; Retinoblastoma

Introduction

Retinoblastoma is the most common intraocular malignant tumor in childhood with an incidence of 1 for 20,000 births which occurs during the first five years of life. The retinoblastoma cell of origin has been debated for over a century; however, we now have a detailed understanding of the neuronal pathways that are deregulated in the primitive cells of the retina. The role of genomic instability of the retinoblastoma gene *Rb1* localized on chromosome 13q and the most relevant epidemiological issues concerning retinoblastoma have recently been pointed out to explain the variegated clinical expression of the disease [1-3]. The most common signs of retinoblastoma are a visible whiteness in the pupil called leukocoria and a strabismus. The unilateral form of the disease represents 70% of cases; however 30% of children develop the bilateral form in both eyes.

Retinoblastoma is often curable when it is diagnosed early, but if it is not treated promptly, this cancer can spread beyond the eye to other parts of the body. When tumor is exclusively localized into the eye, the majority of patients can be cured by therapeutic treatments including enucleation or conservative treatments using, intravenous and/or intra-arterial chemotherapy [4,5], cryotherapy [6], local thermotherapy or brachytherapy. External beam radiotherapy is increasingly restricted due to the risk of late effects. The potential risks of late effects associated with chemotherapy and carboplatin include increased risk of second cancer, as well as short-term important side effects. Besides, due to the lack of blood supply to vitreous seeding, chemotherapy drugs could not reach the vitreous body through blood-retinal barrier. This is one of the important limiting factors which triggered intravenous chemotherapy as a new injection technique for vitreous disease in retinoblastoma [7].

In this context, nanotechnologies seem to offer new perspectives for the treatment of retinoblastoma [8]. From day to day nanomedicine

appears as a non-invasive therapeutic solution. Encapsulated in nanocarriers, drugs could target only cancer cells by considering the tumor microenvironment, the biomarkers overexpressed on the cancer cells and different physical methods for non-invasive therapies [9]. The photodynamic therapy (PDT) is one of the biophysical methods of nanoparticle activation based on the combination of three partners which are a light-activated photosensitizer encapsulated in the nanoparticle, the surrounding oxygen able to produce a toxic compound and an energy laser beam to induce activation [10]. The activated photosensitizers transfer their energy to molecular oxygen which forms highly reactive singlet oxygen and other reactive oxygen species. This phenomenon induces the death of cells having internalized photosensitizers. It is important to note that the photosensitizers are photoactivable by a near-infrared wavelength that is non-mutagenic [11]. Since few years, the specific targeting of nanomedicine to carbohydrate receptors performs a useful strategy for improving the efficacy of treatments. Recently, carbohydrate-targeted systems have been developed to target cancer. Nanoparticles loaded with photosensitizers and covalently linked with mannose residues were synthesized and their improvement uptake in cancer cells was demonstrated *in vitro* and *in vivo* [12-15].

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In this study, we followed-up of the efficiently internalization of mannose functionalized mesoporous silica nanoparticles (MSN) in solid tumor. Then, we focused on MSN potential for one-photon PDT combined with drug delivery and carbohydrate targeting applied on a human retinoblastoma cell line. A comparison with MSN designed for two-photon PDT indicated the potential of this technique to efficiently kill retinoblastoma cancer cells using irradiation at low fluency.

Materials and Methods

Synthesis of MSN

Different nanoparticles were designed linked or not with galactose (Gal) or mannose (Man) residues and loaded with or without fluorescein (FITC), camptothecin (CPT), one-photon photosensitizer (1 γ PS) or two-photon photosensitizer (2 γ PS). Synthesizes of MSN-FITC, MSN-FITC-Man, MSN-(CPT)-Gal or -Man, MSN-(CPT+1 γ PS)-Gal or -Man, MSN-(1 γ PS)-Man and MSN-(2 γ PS)-Man were already described in our previous work [8,12,13].

Cell culture conditions

Human retinoblastoma cancer cells (Y-79) were purchased from ATCC (American Type Culture Collection, Manassas, VA). Cells were cultured in Roswell Park Memorial Institute 1640 (RPMI 1640) culture medium supplemented with 20% fetal bovine serum, 100 U.mL⁻¹ streptomycin and allowed to grow in humidified atmosphere at 37°C under 5% CO₂.

Distribution of MSN in xenografted nude mice

Eight nude mice were subcutaneously injected in the flank with 10⁶ cells of pancreatic cancer cell line (capan-1). One month after cancer cell injection, 6 mice had developed subcutaneous tumors with a mean tumor volume of 500 mm³. Mice (n=2) were injected intravenously with 200 μ L of physiological serum alone (control) or supplemented with MSN-FITC (n=2) or MSN-FITC-Man (n=2) (16 mg.kg⁻¹). Three hours later, mice were sacrificed and tumor, kidneys and liver were removed. Then organs were crushed and distribution of MSN was quantified by measurement of FITC intensity, excitation 485 nm and emission 538 nm, through a fluorescence microplate reader (Gemini, Molecular Devices).

PDT one and two-photon excitation and phototoxicity measurement

For monophotonic irradiation, Y-79 cells were seeded into 96-well plates at 10,000 cells per well in 100 μ L culture medium and allowed to grow for 24 h. Then cells were incubated for 24 h with or without 20 μ g.mL⁻¹ MSN. After incubation, cells were submitted or not to laser irradiation for 30 min (650 nm, 90 J.cm⁻²). For two-photon excitation irradiation, Y-79 cells were seeded into 384 multi-well glass bottom (thickness 0.17 mm), with a black polystyrene frame, 2,000 cells per well in 50 μ L culture medium and allowed to grow for 24 h. Cells were then incubated for 24 h with or without 20 μ g.mL⁻¹ MSN. After incubation with MSN, cells were submitted or not to laser irradiation. Scanned areas were 1.5 \times 1.5 mm². The entire area of the well was irradiated at 760 nm by 3 scans of 1.25 s each. The average power delivered to the sample was measured with a thermoelectric optical energy meter and was 47 J.cm⁻². The laser beam is focused by a microscope objective lens (10X, NA 0.4). Two day after irradiations (one- or two-photon), a MTS assay was performed to evaluate the phototoxicity of MSN by adding 333 μ g.mL⁻¹ of tetrazolium salt (CellTiter 96 aqueous One Solution Reagent, Promega) solution. Cells were incubated 3 h at 37°C

and the absorbance was read at 490 nm in a plate reader to determine the formazan concentration, which is proportional to the number of living cells.

Cytotoxicity assay for drug delivery

For these experiments, Y-79 cells were seeded into 96-well plates at 10,000 cells per well in 100 μ L culture medium and allowed to grow for 24 h. Then cells were incubated at different times with or without 20 μ g.mL⁻¹ MSN. To evaluate the cytotoxicity of MSN, a MTS assay was performed as described above.

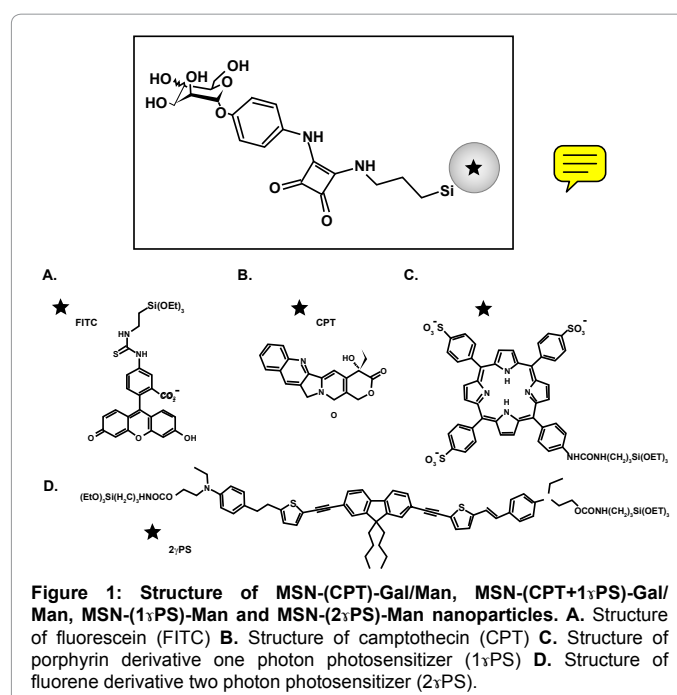
Statistical analysis

Statistical analysis was performed using the Student's *t* test to compare paired groups of data. A *p* value of <0.05 was considered to be statistically significant.

Results

In previous work, we demonstrated the lectin-mediated cellular uptake of functionalized nanoparticles and the specific endocytosis involving carbohydrate receptors expressed by cancer cells [8]. Here, we designed MSN encapsulated with fluorescein (FITC) or camptothecin (CPT) or a porphyrin derivative one-photon photosensitizer (1 γ PS) [11] or a fluorene derivative two-photon photosensitizer (2 γ PS) [16] grafted with mannose or galactose through a squarate linker (Figure 1). In this work, we first verified the potential accumulation of MSN linked with mannose residue into mice tumors. For this, nude mice with subcutaneous tumors were intravenously injected with FITC-loaded MSN covered or not with mannose (Figure 1A). Three hours after injection we observed a higher accumulation in tumor of mice treated with MSN-FITC-Man, while MSN-FITC was mainly retained by liver (Figure 2). This demonstrates the high tumor-targeting efficiency of mannose residues grafted at the surface of MSN.

Then, we focused on Y-79 retinoblastoma cells in order to provide new evidences of the potential of targeted MSN for the treatment of retinoblastoma. To evaluate the efficiency of drug delivery combining



to PDT, cells were treated with MSN-(CPT)-Gal or -Man, MSN-(CPT+1 γ PS)-Gal or -Man (Figure 1B and 1C). CPT-drug delivery on Y-79 cells treated with MSN-(CPT)-Gal or -Man induced 24% and 26% of cell death, respectively. In contrast, when irradiated MSN-(CPT+1 γ PS)-Gal or -Man induced 68% and 58% of cell death, respectively (Figure 3). We have verified that MSN-(CPT+1 γ PS)-Gal/Man were no more toxic in the dark than MSN-(CPT)-Gal/Man and that irradiation alone was not toxic for cells (data not shown). These results suggest that a high therapeutic potential of synergistic use of drug delivery and PDT can be provided using the same MSN.

We next compared the PDT efficiency between 2 γ PS encapsulated mannose-grafted MSN (Figure 1D) and 1 γ PS-loaded one (Figure 1C). Cells were incubated for 24 h with 20 $\mu\text{g}\cdot\text{mL}^{-1}$ of MSN-(1 γ PS)-Man or MSN-(2 γ PS)-Man and one-photon (90 $\text{J}\cdot\text{cm}^{-2}$) or two-photon (47 $\text{J}\cdot\text{cm}^{-2}$) irradiations were performed. As shown in Figure 4, the photodynamic therapeutic potential of MSN-(2 γ PS)-Man is clearly higher even if power is less intense, 67% of cell death, than MSN-(1 γ PS)-Man, 28% of cell death. The nanoparticles were found to be non-toxic without irradiation and irradiation alone did not damage the cells (data not shown). These results indicate that MSN designed for two-photon excitation PDT present interesting therapeutic properties to kill retinoblastoma cancer cells.

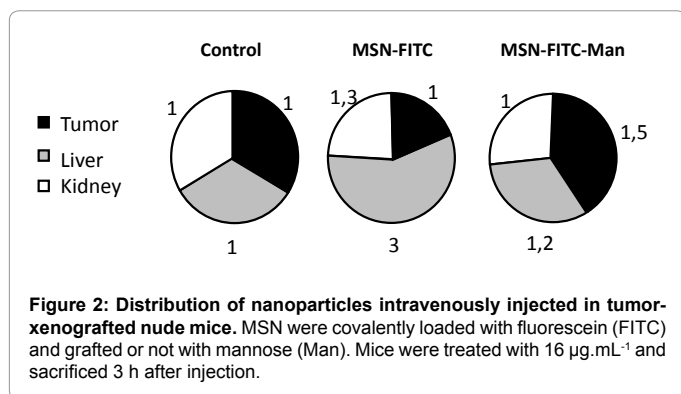


Figure 2: Distribution of nanoparticles intravenously injected in tumor-xenografted nude mice. MSN were covalently loaded with fluorescein (FITC) and grafted or not with mannose (Man). Mice were treated with 16 $\mu\text{g}\cdot\text{mL}^{-1}$ and sacrificed 3 h after injection.

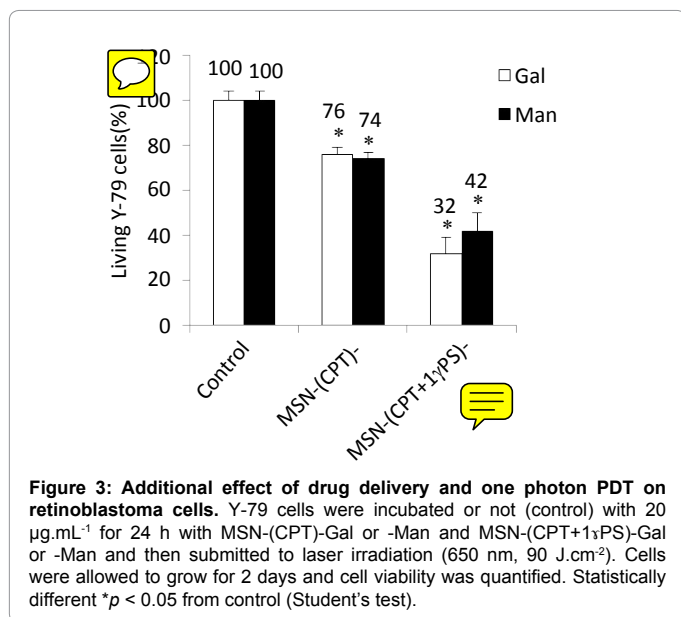


Figure 3: Additional effect of drug delivery and one photon PDT on retinoblastoma cells. Y-79 cells were incubated or not (control) with 20 $\mu\text{g}\cdot\text{mL}^{-1}$ for 24 h with MSN-(CPT)-Gal or -Man and MSN-(CPT+1 γ PS)-Gal or -Man and then submitted to laser irradiation (650 nm, 90 $\text{J}\cdot\text{cm}^{-2}$). Cells were allowed to grow for 2 days and cell viability was quantified. Statistically different * $p < 0.05$ from control (Student's test).

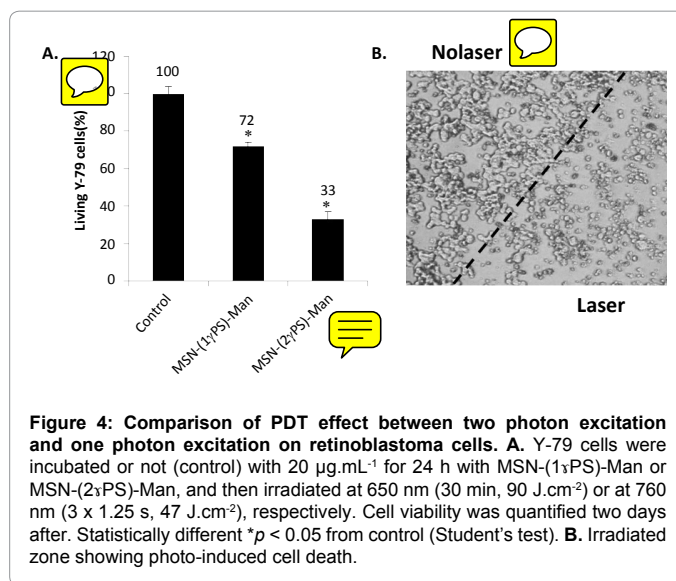


Figure 4: Comparison of PDT effect between two photon excitation and one photon excitation on retinoblastoma cells. A. Y-79 cells were incubated or not (control) with 20 $\mu\text{g}\cdot\text{mL}^{-1}$ for 24 h with MSN-(1 γ PS)-Man or MSN-(2 γ PS)-Man, and then irradiated at 650 nm (30 min, 90 $\text{J}\cdot\text{cm}^{-2}$) or at 760 nm (3 x 1.25 s, 47 $\text{J}\cdot\text{cm}^{-2}$), respectively. Cell viability was quantified two days after. Statistically different * $p < 0.05$ from control (Student's test). B. Irradiated zone showing photo-induced cell death.

Conclusion

In this study, a significant therapeutic synergy between one-photon PDT combined with CPT-drug delivery has been revealed. Furthermore, promising results of two-photon PDT were obtained at 760 nm irradiation and low fluency 47 $\text{J}\cdot\text{cm}^{-2}$. Using of two-photon photosensitizers the window for excitation is extended into the near infrared region, thereby making the process more viable to be used on the human body. The data provide new evidences of the potential of functionalized and targeted MSN for treatment of retinoblastoma and could lead to propose a non-invasive therapy. Considering their sizes and properties in release, specific targeting and minimizing or eliminate side effects resulting directly from current administration methods, MSN appear to be the most promising tool to meet the primary requirements of an ideal ocular delivery system [17]. Current investigations are carried out to identify which carbohydrate receptors are overexpressed in retinoblastoma to improve therapeutic efficiency.

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