

FTIR has the Potential to Detect Stem Cells in the Bovine Corneal Stroma

Christina S. Kamma-Lorger^{1*}, Katia Wehbe², Craig Boote¹, Keith M. Meek¹ and Gianfelice Cinque²

¹Structural Biophysics Group, School of Optometry and Vision Sciences, Cardiff University CF24 4LU, UK

²Diamond Light Source, Didcot, Oxfordshire OX11 0DE

Abstract

Fourier Transform Infrared Spectroscopy (FTIR) was used for the first time to locate stem cells in the corneal stroma. An off-line microscope was used in Diamond light source and a bovine cryo-section was examined. Differences in spectra from two different regions suggest the existence of a cell population that has stem cell characteristics. Synchrotron FTIR would provide a greater scanning resolution than the off-line microscope used, therefore it could be an ideal technique to locate stromal stem cells in the cornea.

Keywords: Fourier Transform Infrared Spectroscopy ; Cornea; Stem cells; Stroma

Following corneal injury or refractive surgery, an intense wound healing response is initiated that seeks to restore tissue function and stability. Stem cell populations, thought to play a key role in corneal regeneration, are suspected to reside in both the epithelial and stromal layers. Corneal epithelial stem cells have been widely investigated in the past and are one of the most well understood stem cell systems in the body [1-3]. However little is known about the stem cell population in the stromal layer (adjacent to the epithelium) which, following corneal trauma, is suspected to replace the normally quiescent stromal keratocytes of mesenchymal origin. Du et al. [4] proposed the existence of a stromal stem cell population in the cornea, but the exact location of these cells was not specified. In 2006 German et al. [2] successfully used FTIR spectroscopy to locate epithelial stem cells in the normal bovine cornea. Here we present the first documented application of FTIR techniques to detect stem cells in the corneal stroma.

An off line FTIR microscope in DIAMOND light source (Oxfordshire, UK) was used to obtain spectra from a healthy bovine cryo-section (30 μm thickness, limbus-to-limbus) at two different locations at 20 μm spatial resolution. (Settings used: 4 cm^{-1} , 128 scans, 15 \times objective, single channel detector). Statistical comparison (t-test) was employed in order to identify differences in the features of the FTIR spectra between the two different corneal areas. Comparison of the spectra between the two locations examined (Figure 1) revealed cell cycle differences previously attributed with stem cell characteristics [2]. These differences were statistically significant for spectral wavenumbers associated with C=O stretching vibration (i.e. $\sim 1725 \text{ cm}^{-1}$), cell cycle changes (i.e. $\sim 1315 \text{ cm}^{-1}$) and RNA expression (i.e. $\sim 1110 \text{ cm}^{-1}$) [2,5]. These findings are related to cellular proliferative activity and according to German et al. [2] these properties describe stem cell characteristics and, in our case, point strongly to the possibility of

another cell population, different to quiescent keratocytes, existing in the corneal stroma.

We hypothesise that key spectral features present in the data presented herein derive from a stromal stem cell population. However given the limited photon flux density of blackbody IR sources, combined with the extremely low absorbances typically obtained using corneal sections, these spectra may only be attributed in part to quiescent keratocytes. It is likely that a proportion of the signal is also due to other extracellular matrix components. Therefore, these preliminary findings require further investigation, ideally exploiting the higher light intensity available from a synchrotron source. This will render greater scanning resolution, facilitating collection of spectra even from single cells around 10 to 15 μm in diameter [2], and thereby giving a more detailed profile of the exact stromal cell population.

Acknowledgments

The authors would like to thank Dr. Che Connon for his useful commenting on the manuscript. This work was funded by a Medical Research Council 5-year programme grant (G0600755). Prof. Keith M. Meek is a Royal Society-Wolfson Research Merit Award Holder.

References

1. Daniels JT, Harris AR, Mason C (2006) Corneal epithelial stem cells in health and disease. *Stem Cell Rev* 2: 247-254.
2. German MJ, Pollock HM, Zhao B, Tobin MJ, Hammiche A, et al. (2006) Characterization of putative stem cell populations in the cornea using synchrotron infrared microspectroscopy. *Invest Ophthalmol Vis Sci* 47: 2417-2422.
3. Bentley AJ, Nakamura T, Hammiche A, Pollock HM, Martin FL, et al. (2007) Characterization of human corneal stem cells by synchrotron infrared microspectroscopy. *Mol Vis* 13: 237-242.
4. Du Y, Funderburgh ML, Mann MM, SundarRaj N, Funderburgh JL (2005) Multipotent stem cells in human corneal stroma. *Stem Cells* 23: 1266-1275.
5. Jayasuriya AC, Scheinbeim JI, Lubkin V, Bennett G, Kramer P (2003) Piezoelectric and mechanical properties in bovine cornea. *J Biomed Mater Res A* 66: 260-265.

***Corresponding author:** Christina S. Kamma-Lorger, Structural Biophysics Group, School of Optometry and Vision Sciences, Cardiff University CF24 4LU, UK, E-mail: KammaCS@cardiff.ac.uk

Received November 07, 2011; **Accepted** December 26, 2011; **Published** December 29, 2011

Citation: Kamma-Lorger CS, Wehbe K, Boote C, Meek KM, Cinque G (2011) FTIR has the Potential to Detect Stem Cells in the Bovine Corneal Stroma. *J Physic Chem Biophysic* 1:103. doi:10.4172/2161-0398.1000103

Copyright: © 2011 Gatenby RA, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

