

## First Phytochemical Characterization and Essential Oil Analysis of the Traditional Catalan Wild Salad: “Coscoll” (*Molopospermum peloponnesiacum* (L.) Koch)

Andreu V<sup>1,2\*</sup>, Amiot A<sup>2</sup>, Safont M<sup>1</sup>, Levert A<sup>2</sup> and Bertrand C<sup>1,2</sup>

<sup>1</sup>Centre de Recherches Insulaires et Observatoire de l'Environnement, USR 3278, University of Perpignan Via Domitia, Perpignan, France

<sup>2</sup>AkiNaO, Perpignan, France

### Abstract

“Coscoll” (*Molopospermum peloponnesiacum* (L.) Koch) whose stems are traditionally consumed raw in salads in Catalonia is associated in oral tradition with many virtues as digestive, purifying, exciting, antioxidant and hemato-cathartic activities. However, stem composition and biological activity had never been studied. Nutritive values of plant material were determined by official methods and constituents from essential oil of *Molopospermum peloponnesiacum* stems were characterized for the first time in this study. The main constituents were dillapiol (60.1%) and 3-carene (15.2%). Moreover, total polyphenol content using Folin-Ciocalteu method, and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity were determined. *Molopospermum peloponnesiacum* stem total polyphenol content was 101.0 ± 10.0 mg/100 g fresh weight (Gallic Acid Equivalent) and DPPH radical scavenging activity, relatively high, was close from spinach one. Thus, these first results reveal the potential beneficial properties of this plant, in relation with its traditional use in Catalonia.

**Keywords:** *Molopospermum peloponnesiacum* stems; Nutritive values; Essential oil analysis; Total polyphenol content; DPPH radical scavenging activity

### Introduction

*Molopospermum peloponnesiacum* (L.) Koch is a perennial plant endemic to mountainous area of Southern Alps and Pyrenees. *Molopospermum* is a monotypic genus belonging to the Apiaceae family and Apioideae subfamily. Its phylogenetic classification is not clearly defined [1-3] but most recent phylogenetic study based on chloroplast genome trnQ-trnK region analysis seems to integrate *Molopospermum* into the clade of Annesorhizeae [4].

In Pyrenees, where the plant is also name “Coscoll” in Catalan language, *M. peloponnesiacum* young shoots are traditionally eaten raw in salads. Oral tradition extols many virtues of coscoll, as digestive, purifying, and exciting activities. It has been collected throughout generations in this country and cures are traditionally made from coscoll for its hemato-cathartic properties [5].

*M. peloponnesiacum* contains a large amount of volatile compounds and gives off a strong odor. Root and fruit essential oil have been identified in literature, containing mainly 3-carene, trimethylbenzoic acids, and dillapiol as major compounds [6-7]. However, to our knowledge, stem composition or biological activity had never been characterized. This paper thus deals with the chemical characterization of stems. We described nutritive characterization and essential oil analysis. In addition, we determined stem polyphenol content and 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity in order to identify potential beneficial properties in this plant material traditionally consumed.

### Materials and Methods

#### Plant material

Plant material was collected in May 2011 in Mantet Col (Pyrénées-Orientales, France) during inflorescence emergence stage, as stems are traditionally consumed. All plants specimen were 3-4 years old and were identified by Cédric Bertrand based on morphological description.

A voucher specimen was deposited at the Herbarium of the University Claude Bernard Lyon1, city of Villeurbanne, France, under the name “Collection Piola” and collector number 4.

An aliquot was dried and conserved for phytochemical analysis. Dry mass stems was 15% w:w.

#### Global characterization of plant material

Protein dosage was determined according to the AOAC Official Method 984.13 [8]. Lipid amount was determined using the ISO 1443 method [9] and lipid profile was analyzed according to the AOCS Ce 1h-05 method [10]. Soluble sugar content was determined after oximation and silylation then analyzed by gas chromatography with a 30 m SIL5CB column coupled with FID detection, using xylitol as internal standard. Mineral amount was evaluated according to the AOAC Official Method 923.03 [11]. Fiber dosage was determined according to the AOAC Official Method 985.29 [12]. Energetic values were calculated according European regulation 1169/2011 [13]. All essays were done on a mix of stems from six different plants.

#### Essential oil distillation

100 g of fresh stem parts from six *Molopospermum peloponnesiacum* specimen from the same mountain area were directly subjected to hydrodistillation for 2 h. Volatile oil was collected by extraction using

**\*Corresponding author:** Vanessa Andreu, Centre de Recherches Insulaires et Observatoire de l'Environnement, USR 3278, University of Perpignan Via Domitia, 66860 Perpignan, France, Tel: +33-468-662-073; E-mail: [vanessa.andreu@univ-perp.fr](mailto:vanessa.andreu@univ-perp.fr)

**Received** September 14, 2015; **Accepted** September 25, 2015; **Published** September 28, 2015

**Citation:** Andreu V, Amiot A, Safont M, Levert A, Bertrand C (2015) First Phytochemical Characterization and Essential Oil Analysis of the Traditional Catalan Wild Salad: “Coscoll” (*Molopospermum peloponnesiacum* (L.) Koch). Med Aromat Plants 4: 211. doi:10.4172/2167-0412.1000211

**Copyright:** © 2015 Andreu V., et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

heptane and aqueous phase was separated with anhydrous sodium sulfate. Extracted oils were stored in glass vessels at -20°C and protected from light. Essential oil extraction yield was 0.047% w:w fresh stems.

### GC-FID and GC-MS analysis of the essential oil

Essential oil components were identified using gas chromatography with Thermo Scientific Focus GC system coupled with DSQ II system mass detector. Analysis was realized with a HP-5MS column (30m × 0.25 mm i.d. × 0.25 μm, 5%-phenyl-arylene-95%-dimethylpolysiloxan, Phenomenex, Torrance, California, US). Column head flow was set at 1 ml/min, using helium as carrier gas. Column temperature was programmed as follows: 80°C for 3 min, 80°C to 110°C (2°C/min), 110°C to 240°C (5°C/min), 240°C to 290°C (10°C/min) and 290°C for 3 minutes. Total run time was 52 min.

Injector temperature was maintained at 280°C, and injection volume was 1.0 μL in split mode with a 10 ml/min split flow. Transfer line temperature was 300°C. Electron multiplier voltage was set to 1330 V by automatic tuning. Mass spectra were recorded at 70 eV with the mass range at 50-650. Linear retention indices were calculated for all constituents using homologous series of n-alkanes (Alkane solution C<sub>8</sub>-C<sub>20</sub>, Fluka, Buchs, Switzerland). Essential oil constituents were identified through mass spectra studies and were confirmed by comparing retention indices with those reported in literature [14-18].

Major compounds were quantified using gas chromatography with a Thermo Scientific Focus GC and flame ionization detector with the same conditions described previously. The relative quantification of the components was performed by comparison of their peak area. Mean and standard deviation were calculated on the basis of replicate from six different plants.

### Preparation of alcoholic extract

50 g of dry stems were extracted three times with 500 ml of ethanol for 15 minutes in an ultrasonic bath, then filtered and dried under vacuum. Extraction yield was 11.4% (w:w dry stems).

### Determination of total polyphenol contents

Total polyphenol content was determined by Folin-Ciocalteu phenol method as described by Singleton et al. [19] with some modifications. 175 μl of appropriately diluted sample or gallic acid standard were added to 25 μl of Folin-Ciocalteu phenol reagent and 50 μl of 20% (w/v) Na<sub>2</sub>CO<sub>3</sub> solution and mixed. After incubation for 30 minutes at room temperature, absorbance was measured at 725 nm versus a prepared blank containing solvent instead of gallic acid or sample. The calibration equation for gallic acid was  $y=0,0395 \times +0.0008$  (R<sup>2</sup>=0.9993).

Total phenolic content was expressed as mg gallic acid equivalent (GAE)/100 g of fresh plant. Mean and standard deviation (n=3) were calculated.

Folin reagent and gallic acid were purchased from Sigma-Aldrich (Saint-Louis, Missouri, USA).

### Determination of DPPH radical scavenging activity

*M. peloponnesiacum* stems ethanolic extract free radical scavenging activity was determined in 96-well microplates using the DPPH method [20] with some modifications. 20 μl ethanol solution containing a different concentration of *M. peloponnesiacum* stems extract was added to 200 μl of freshly prepared 2,2-diphenyl-1-picrylhydrazyl methanol solution (0,2 mM) (Sigma-Aldrich, Saint-Louis, Missouri, United

States). Ethanol was used as the control. After 60 minutes of incubation at room temperature in the dark, absorbance was measured at 515 nm using a microplate reader Aviso, Sirius HT (Ebersberg, Germany). Vitamin-C was used as standard (Supelco, Bellefonte, Pennsylvania, US).

Standard curves for assay was obtained by measuring the DPPH scavenging activity of 1, 5, 10, 25, 50 mg vitamin C/L. DPPH scavenging activity was expressed as mg vitamin C equivalent (VCE)/100 g of fresh plant. Mean and standard deviation (n=3) were calculated.

## Results and Discussions

### Global characterization of *Molopospermum peloponnesiacum* stems

Content of soluble sugars, fibers, protein, ashes and lipids in fresh and dried stems are presented in Table 1. Soluble sugars, fibers and protein values were particularly high in *Molopospermum peloponnesiacum* stems whereas lipid amount was very low.

### Essential oil analysis

The essential oil composition is listed in Table 2. Main compounds identified in stem essential oil were dillapiol (60% relative content) and 3-carene (15% relative content). Stem essential oil composition was very close to the root essential oil composition determined in literature for dillapiol chemotype [7]. Dillapiol is a well-known phenylpropanoid, extracted from essential oils of several plants as matico (*Piper aduncum*), parsley (*Petroselinum crispum*), pepper elder (*Peperomia pellucida*) or dill (*Anethum graveolens*) [21-24]. This compound presents *in vitro* antieishmanial, gastroprotective and anti-inflammatories activities [21-23]. Its structure, close to phenylisopropylamine structures could suggest a possible psychotropic activity as tonic or exciting [25].

### Determination of total polyphenol content and DPPH radical scavenging activity

Total polyphenol content and 2,2-diphenyl-1-picrylhydrazyl radical scavenging activity of *M. peloponnesiacum* stems are shown in Table 3. Total polyphenol content determined for the *peloponnesiacum* stems expressed in Gallic Acid Equivalent (GAE) was 101.0 ± 10.0 mg GAE/100 g fresh weight. DPPH radical scavenging capacity expressed in vitamin C equivalent (VCE) was 77.7 ± 5.59 mg VCE/100 FW. *M. peloponnesiacum* stem has relatively strong DPPH radical scavenging

Analyse	Unit	Value*
Water	g/100g FW	85
Soluble sugar	g/100g FW (g/100g DW)	5.56 (37.0)
including simple sugar	g/100g FW (g/100g DW)	3.46 (23.1)
Fiber	g/100g FW (g/100g DW)	4.66 (31.1)
Protein	g/100g FW (g/100g DW)	3.28 (21.9)
Ashe	g/100g FW (g/100g DW)	1.46 (9.7)
including sodium	mg/100g FW (mg/100g DW)	3.9 (26)
Lipid	g/100g FW (g/100g DW)	0.04 (0.3)
FA saturated	g/100g FW (g/100g DW)	0.02 (0.1)
FA monosaturated	g/100g FW (g/100g DW)	0.00 (0.00)
FA polysaturated	g/100g FW (g/100g DW)	0.03 (0.2)
Energetic value	Kcal/100g FW (Kcal/100g DW) KJ/100g FW (KJ/100g DW)	45 (300)188 (1255)

FA : Fatty acids ; FW : fresh weight ; DW : Dry weight.

\* Values were obtained according official dosage methods; essays were done on a mix of stem from six different plants

**Table1:** Nutritive composition of *Molopospermum peloponnesiacum* stems.

N°	RI	RI ref	Ref	Compound	Relative content (%) <sup>*</sup>	mg/100g FW <sup>*</sup>
1	937	939	[14]	α-pinene	2.77 +/- 1.16	1.29 +/- 0.54
2	975	975	[14]	Sabinene	0.14 +/- 0.03	0.06 +/- 0.01
3	984	979	[14]	β-pinene	0.92 +/- 0.35	0.43 +/- 0.16
4	990	991	[14]	Myrcene	0.56 +/- 0.17	0.26 +/- 0.08
5	1016	1011	[14]	3-carene	15.22 +/- 4.18	7.1 +/- 1.94
6	1026	1027	[15]	o-cymene	0.35 +/- 0.04	0.16 +/- 0.02
7	1034	1029	[14]	limonene	3.68 +/- 0.85	1.71 +/- 0.4
8	1058	1060	[14]	γ-terpinen	0.57 +/- 0.16	0.26 +/- 0.08
9	1089	1089	[14]	terpinolene	0.38 +/- 0.03	0.17 +/- 0.01
10	1101	1097	[14]	α-linalool	0.29 +/- 0	0.13 +/- 0
11	1131	1129	[14]	allo-ocimene	0.07 +/- 0	0.03 +/- 0
12	1230	1226	[14]	citronellol	0.39 +/- 0.17	0.18 +/- 0.08
13	1237	1236	[16]	2-Nonyl acetate	0.215 +/- 0.025	0.1 +/- 0.01
14	1337	1338	[14]	δ- elemene	1 +/- 0.45	0.46 +/- 0.21
15	1352	1353	[14]	citronellyl acetate	0.78 +/- 0.16	0.36 +/- 0.08
16	1422	1419	[14]	trans-caryophyllene	1.19 +/- 0.64	0.55 +/- 0.3
17	1459	1455	[14]	α-humulene	0.83 +/- 0.23	0.38 +/- 0.11
18	1484	1485	[14]	D-germacrene	1.86 +/- 0.85	0.86 +/- 0.4
19	1498	1500	[14]	bicyclogermacrene	0.9 +/- 0.1	0.42 +/- 0.05
20	1562	1563	[14]	trans-nerolidol	0.31 +/- 0	0.14 +/- 0
21	1623	1621	[14]	dillapiol	60.12 +/- 8.04	28.04 +/- 3.75
22	1667	1654	[14]	α-cadinol	0.29 +/- 0.09	0.13 +/- 0.04
23	1683	1676	[17]	tetradecanol-1	2.72 +/- 0.09	1.27 +/- 0.04
24	1769	ND	ND	m/z 252	1.62 + 0.33	0.75 +/- 0.15
25	1882	1883	[18]	1-hexadecenol	1.19 +/- 0.31	0.55 +/- 0.14

Total monoterpenes 27.33% 12.7 mg/100 g stems

Total sesquiterpenes 5.09% 2.3 mg/100 g stems

Total other compounds 64.32% 29.9 mg/100 g stems

Total identified 96.74% 44.9 mg/100 g stems

RI : Retention indices calculated with n-alkanes. Percentage are calculated from FID data.

RI ref : Retention indices from literature.

ND : Not determined.

FW : fresh weight.

\* Data are the mean values of six replicates

**Table 2:** Composition of *Molopospermum peloponnesiacum* stem essential oil.

Total Polyphenol Content*	DPPH radical scavenging capacity*
mg GAE.100g <sup>-1</sup> FW	mg VCE/100 g FW
101.0 ± 10.0	77.7 ± 5.59

GAE : Gallic Acid Equivalent ; FW: Fresh Weight ; VCE : Vitamin C Equivalent.

\*Data are the mean value of three replicates

**Table 3:** Total polyphenol content (TPC) and DPPH free-radical scavenging capacity of *Molopospermum peloponnesiacum* fresh stems.

capacity. For comparison, this result is close to that obtained in literature for spinach (71 mg VCE/100 FW) and nearly twice that of lettuce (43 mg VCE/100 FW) [26].

## Conclusion

In this study, *Molopospermum peloponnesiacum* stems composition was characterized for the first time. We looked at its nutritive values and antioxidant capacity. Our results indicate a large amount of soluble sugars, fibers and protein in the stems and an essential oil containing dillapiol and 3-carene as major compounds, similar to that described previously for root essential oil. This first study permitted to chemically characterize this part of the plant that is traditionally consumed and to get a better idea of its potential beneficial properties.

## Acknowledgements

The authors of this study are very thankful to Daniel Aspe for his help in

plant collection and for sharing his knowledge on "coscoll". The spectroscopic experiments have been performed using the "Biodiversité et Biotechnologies Marines" (Bio2Mar) facilities at the University of Perpignan.

## References

- Liu MR, Plunkett GM, Lowry PP, Van Wyk BE, Tilney PM (2006) The taxonomic value of fruit wing types in the order Apiales. Ame J Bot 93: 1357–1368.
- Calviño CI, Tilney PM, Van Wyk BE, Downie SR (2006) A molecular phylogenetic study of southern African Apiaceae. Am J Bot 93: 1828–1847.
- Magee AR, Van Wyk BE, Tilney PM, Downie SR (2008) Ezosciadium (Apiaceae): a taxonomic revision of yet another early diverging South African apioid genus. Plant Syst Evol 276: 167–175.
- Magee AR, Calviño CI, Liu M, Downie SR, Tilney PM, et al. (2010) New tribal delimitations for the early diverging lineages of Apiaceae subfamily Apioideae. Taxon 59: 567–580.
- Parada M, Carrio E, Vallès J (2011) Ethnobotany of food plants in the Alt emporda region (Catalonia, Iberian Peninsula). J Appl Bot Food Qual 84: 11–25.
- Kubeczka KH, Ullmann I (1981) Terpenoids of the essential oil from *Molopospermum peloponnesiacum* roots. Phytochem 20: 828–830.
- Kubeczka KH, Ullman I (1983) Chemotypes of *Molopospermum peloponnesiacum* (L.) Koch. Z Naturforsch 38c: 189–193.
- Association Of Analytic Chemistry (2005) AOAC Official Method 984.13 (A-D). (18<sup>th</sup> edn), Official Methods of Analysis of AOAC international, Gaithersburg, Maryland, US.
- International Organization For Standardization (1973) ISO 1443. Meat and meat products. Determination of total fat content. International organization for standardization, Geneva, Swiss.
- American Oil Chemists Society (2005) Official method Ce 1h-05. (6<sup>th</sup> edn), Official methods and recommended practices of the AOCS, Champaign, Illinois, US.
11. Association of Analytic Chemistry (2005) AOAC Official Method 923.03 (A-D). (18<sup>th</sup> edn), Official Methods of Analysis of AOAC international, Gaithersburg, Maryland, US .
- Association of Analytic Chemistry (2005) AOAC Official Method 985.29 (A-D) Official. (18<sup>th</sup> edn), Methods of Analysis of AOAC international, Gaithersburg, Maryland, US.
- Règlement Européen (2011) Règlement (UE) no 1169/2011 du parlement européen et du conseil du 25 octobre 2011 concernant l'information des consommateurs sur les denrées alimentaires. Journal Officiel de l'Union Européenne L304: 18 – 63.
- Adams RP (2001) Identification of essential oil components by gas chromatography/quadrupole mass spectroscopy. (3<sup>rd</sup> ed), Allured Publishing Corporation Carol Stream, Illinois, US.
- Maxia A, Falconieri D, Piras P, Porcedda S, Marongiu B, et al. (2012) Chemical composition and antifungal activity of essential oils and supercritical CO<sub>2</sub> extracts of *Apium nodiflorum* (L.) Lag. Mycopathologia 174: 61–67.
- National Institute Of Standars And Technology (2004) Retention Data, NIST Mass Spectrometry Data Center. Gaithersburg, Maryland, US.
- Saroglou V, Dorizas N, Kyriotakis Z, Skaltsa HD (2006) Analysis of the essential oil composition of eight *Anthemis* species from Greece. J Chrom A 1104: 313–322.
- Hazzit M, Baaliouamer A, Faleiro ML, Miguel MG (2006) Composition of the essential oils of *Thymus* and *Origanum* species from Algeria and their antioxidant and antimicrobial activities. J Agri Food Chem 54: 6314–6321.
- Singleton VL, Rossi JA (1965) Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. Ame J Enol Vitic 16: 144–158.
- Brand Williams W, Cuvelier ME, Berset C (1995) Use of a free radical method to evaluate antioxidant activity. LWT - Food Sci Technol 28: 25–30.
- Parise Filho R, Pastrello M, Pereira Camerlingo CE, Silva GJ, Agostinho LA, et al. (2011) The anti-inflammatory activity of dillapiole and some semisynthetic analogues. Pharm Biol 49: 1173–1179.
- Parise-Filho R, Pasqualoto KF, Magri FM, Ferreira AK, Da Silva BA, et al.

- 
- (2012) Dillapiole as antileishmanial agent: discovery, cytotoxic activity and preliminary SAR studies of dillapiole analogues. *Arch Pharm (Weinheim)* 345: 934-944.
23. Rojas Martínez R, Arrieta J, Cruz Antonio L, Arrieta Baez D, Velázquez-Méndez AM, et al. (2013) Dillapiole, isolated from *Peperomia pellucida*, shows gastroprotector activity against ethanol-induced gastric lesions in Wistar rats. *Molecules* 18: 11327-11337.
24. Lamarti A, Badoc A, Deffieux G, Carde JP (1993) Etude des arylpropènes extraits de l'huile essentielle des fruits de Persil [*Petroselinum crispum* (mill.) A. W. Hill]. *Bull Soc Pharm Bordeaux* 132: 90-98.
25. Shulgin AT (1967) Psychotropic phenylisopropylamines derived from apiol and dillapiol. *Nature* 215: 1494-1495.
26. Floegel A, Kim DO, Chung SJ, Koo SI, Chun OK (2011) Comparison of ABTS/DPPH assays to measure antioxidant capacity in popular antioxidant-rich US foods. *J Food Compos Anal* 24: 1043-1048.