

Effect of Systemic Estradiol Administration on Circadian Body Temperature and Activity Rhythms in Female Rats

Uchida Y¹, Marui S¹, Tokizawa K¹ and Nagashima K^{1,2}

¹Laboratory of Integrative Physiology, Body Temperature and Fluid Laboratory, Waseda University, Japan

²Institute of Applied Brain Sciences, Waseda University, Saitama, Japan

*Corresponding author: Uchida Y, Women's Environmental Science Laboratory, Department of Health Sciences, Faculty of Human Life and Environment, Nara Women's University, Nara, Japan, Tel: +81-0742-20-3336; E-mail: yukioto@cc.nara-wu.ac.jp

Received Date: August 21, 2017; Accepted Date: August 28, 2017; Published Date: September 04, 2017

Copyright: © 2017 Uchida Y, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Abstract

The estrus cycle affects the circadian body temperature (T_b) and activity rhythms, and progesterone is related to these alterations. However, it is not clear whether estrogen (E_2) influences it. The present study examined whether E_2 affects the circadian T_b and activity rhythm. Ovariectomized rats were implanted with a silastic plate with or without E_2 underneath the dorsal skin ($E_2(-)$ and $E_2(+)$), and these along with sham operated rats (SH) were measured for T_b and activity for 2 weeks. The mean T_b was lower, and mean activity was higher in $E_2(+)$ than that in $E_2(-)$ in the day. In the dark phase, the slope of the relationship between the mean T_b and activity in $E_2(-)$ was the greatest. The slope in $E_2(+)$ and SH was greater in the light phase than that in the dark phase. The daily peak of T_b and activity was lower in $E_2(+)$ than that in $E_2(-)$. The appearance of the nadir in T_b was later in $E_2(+)$ and SH than that in $E_2(-)$. The appearance of the peak in T_b and activity was earlier in $E_2(+)$ and SH than that in $E_2(-)$. Thus, E_2 may modulate the circadian T_b and activity rhythm in female rats.

Keywords: Estradiol; Body temperature; Activity; Circadian

Introduction

It is reported that in an environment of thermo-neutral range [1], where autonomic thermoregulatory responses are minimum, the estrus cycle affects circadian body temperature (T_b) rhythm in women [2-9] and in female rats [10-12]. In women, the change of the rhythm is characterized as the increased mean T_b and decreased circadian amplitude in the luteal phase [7,12]. The estrus cycle in female rats generally lasts 4-5 days, consisting of four phases; two days of diestrus, followed by the proestrus and estrus phases. In the proestrus phase, the mean T_b in the light phase decreases [12]. Moreover, in the proestrus phase, the tail surface temperature (T_{tail}) decreases in the dark phase [13], and spontaneous activity increases [14,15], compared with that in the other phases. The peak in the activity appears earlier in the proestrus rather than on the first day of diestrus [11]. An increase in the mean, peak, and nadir of T_b and a decrease in the amplitude of T_b were observed in the luteal phase, which shows a higher progesterone level compared with that in the follicular phase in women [5]. However, the mechanism involved in the changes of the rhythms remains unclear.

It has been speculated that progesterone, the level of which changes with the estrus cycle, affects the circadian T_b rhythm [2,5,16] in a thermoneutral environment. Progesterone administration suppressed activity in female rats [17]. In contrast to the influence of progesterone on the circadian T_b rhythm, the role of estradiol (E_2) still remains controversial. E_2 did not alter the circadian T_b rhythm in rats [13] and in women [2,3,5]; however, some reports showed that E_2 decreased the T_{tail} in the dark phase in rats [13]. E_2 influences the circadian rhythm of activity in mice [18-21] and hamsters [22,23] but not in rats [14,24]. Thus, we hypothesized that E_2 might modulate the circadian rhythm of T_b and activity.

In the present study, we compared T_b and activity between sham operated rats, ovariectomized rats, and rats administered E_2 externally, to determine the effect of E_2 on the circadian rhythm of T_b and activity.

Methods

Animals

Female Wistar rats (n=24; 224 ± 2 g; age, 8 weeks; Takasugi, Saitama, Japan) were used in the present study. They were individually housed in cages (45 cm \times 25 cm \times 20 cm) at an ambient temperature (T_a) of $27 \pm 0.5^\circ\text{C}$ with a lighting schedule of 12-h light and 12-h complete darkness (lights on at 0700 h, 300 lux at their eye level). All experimental protocols were approved by the Institutional Animal Care and Use Committee of Waseda University.

Surgery

Under inhalation anesthesia with 2% sevoflurane, a radio transmitter (26 mm \times 8 mm, 2.2 g; PDT-4000 HR E-Mitter[®], Starr Life Sciences Corp., Oakmont, PA, USA) for measuring T_b and spontaneous activity was placed in the peritoneal cavity. Bilateral ovariectomy or sham (SH group; n=8) surgery was conducted through a dorsal skin incision. Silastic plates (5 mm \times 25 mm \times 15 mm; Silpot134, 0.2 ml; Silpot catalyst 184, 20 μl ; Dow Corning Toray Co., Ltd, Tokyo, Japan) with and without 17β -estradiol (Sigma, St. Louis, MO, USA) were prepared. One plate was placed under the right dorsal skin. Eight rats were implanted with an E_2 plate ($E_2(+)$ group) and eight other animals had a control plate without E_2 ($E_2(-)$ group). The placement of the estradiol-containing plate results in a constant E_2 level in the plasma at least for 7 days [25]. We set a pharmacological level of E_2 to determine the definitive effect of E_2 . Plasma E_2 concentration was kept at a high level in the $E_2(+)$ group (1208 ± 85 pg/ml) for 26 days. After the surgery, the rats were injected sc with

penicillin G (1,000 U, Meiji Pharmaceutical, Tokyo, Japan) to prevent post-surgical infection, and were placed in the cage at 27°C.

Experimental protocols

Before the surgery, body weight, food intake, and water intake were estimated at 0830-0900 for 5 days. Then, the entry to animal room was restricted for 2 weeks to avoid a time cue effect by the touches and sounds of experimenters during the T_b and activity measurements. After a non-contact period, the same measurements were again performed for 5 days. The signals from the radio transmitter were obtained through a receiver board ER-4000 Energizer/Receiver, Starr Life Sciences Corp.) Every 5 min, and were stored in a personal computer with a data-logging program (VitalView; Starr Life Sciences Corp.). The accuracy of the value of T_b was $\pm 0.1^\circ\text{C}$.

Statistics

The T_b and activity on each day were averaged every 30 min, and also for the light and dark phases and the entire day. The mean, nadir, and peak of the circadian T_b and activity rhythms and the difference between the maximum and minimum (amplitude) were estimated for each day of the 2-week measurement period, and the values were averaged. Differences in these values were assessed by two-way ANOVA with R language (R version 3.1.2, The R Foundation for Statistical Computing). Tukey-Kramer's test was used to identify the significant differences at specific time points of T_b and activity. The null hypothesis was rejected at the level of $P < 0.05$.

Results

The circadian T_b and activity rhythms in a day are shown in Figure 1A and 1B. T_b in the $E_2(-)$ group was lower than that in the SH group at 0-2, 9-10, and 15-23 hours. T_b in the $E_2(+)$ group was lower than that in the SH group at 2-4 and 9-23 hours (Figure 1A). T_b in $E_2(+)$ was lower than that in $E_2(-)$ at 2-16 and 21-24 hours. Activity was lower in the $E_2(-)$ group than that in the SH group at 0-3 and 10-23 hours. Activity was lower in the $E_2(+)$ group than that in the SH group at 7-8, 11-12, and 21-23 hours. Activity was higher in the $E_2(+)$ group than that in the $E_2(-)$ group at 1-3, 10-11, and 12-23 hours (Figure 1B).

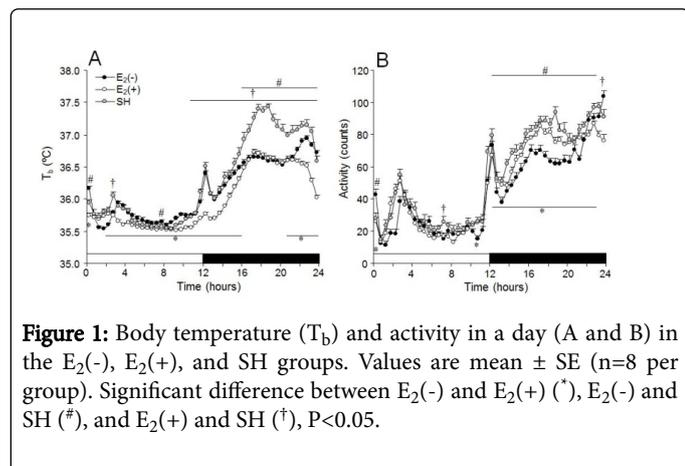


Figure 1: Body temperature (T_b) and activity in a day (A and B) in the $E_2(-)$, $E_2(+)$, and SH groups. Values are mean \pm SE (n=8 per group). Significant difference between $E_2(-)$ and $E_2(+)$ (*), $E_2(-)$ and SH (#), and $E_2(+)$ and SH (†), $P < 0.05$.

Figure 2A and 2B show the relationship between T_b and activity during the light and dark phases. Table 1 shows the slope of the regression line between T_b and activity in the light and dark phases. The slope in the light phase was not different among groups; however,

in the dark phase, the slope in the $E_2(-)$ group was the greatest among the three groups. There was no difference in the slope in the $E_2(-)$ group between the light and dark phases; however, in the $E_2(+)$ and SH groups, the slope was greater in the light phase than that in the dark phase.

Slope of the regression line between T_b and activity in light and dark phases in the $E_2(-)$, $E_2(+)$, and SH groups.			
	$E_2(-)$	$E_2(+)$	SH
Light Phase	55.2 \pm 2.8	64.4 \pm 6.4 §	52.0 \pm 4.3 §
Dark Phase	53.8 \pm 2.9*.#	28.4 \pm 2.3	24.9 \pm 0.9

Table 1: Slope of the regression line between body temperature (T_b) and activity in the light and dark phases in the $E_2(-)$, $E_2(+)$, and SH groups. Values are mean \pm SE (n=8 per group). Significant difference between $E_2(-)$ and $E_2(+)$ (*), $E_2(-)$ and SH (#), and light and dark phases (§), $P < 0.05$.

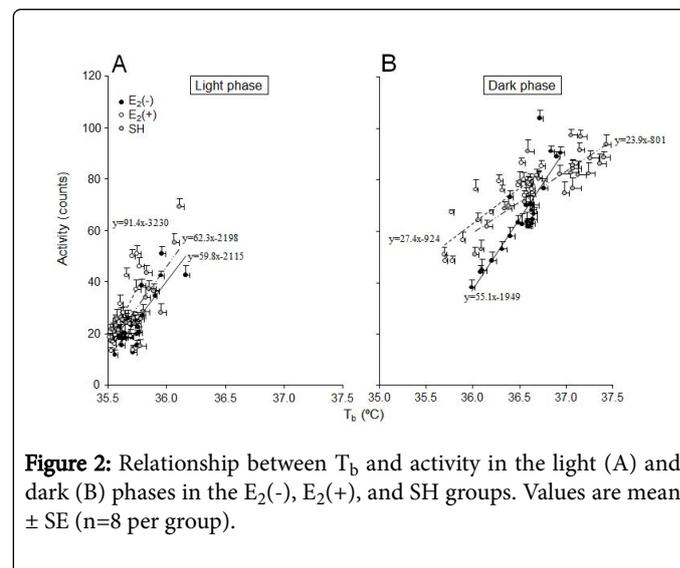


Figure 2: Relationship between T_b and activity in the light (A) and dark (B) phases in the $E_2(-)$, $E_2(+)$, and SH groups. Values are mean \pm SE (n=8 per group).

Table 2 shows the mean in the day, light, and dark phase, the amplitude, the nadir, and the peak of T_b ($^\circ\text{C}$) and activity (counts). The mean in a day of T_b in SH was the greatest among all groups. The value was not different between the $E_2(+)$ and $E_2(-)$ groups. The mean in the light phase of T_b was not different among the groups. On the other hand, the mean in the dark phase and the amplitude of T_b in the SH group were greater than those in the $E_2(-)$ group, and did not differ from those in the $E_2(+)$ group. The values in the $E_2(+)$ group were lower than those in the $E_2(-)$ group. The nadir of T_b was not different among groups; however, the peak of T_b in the SH group was greatest among the groups; the value in the $E_2(+)$ group was greater than that in the $E_2(-)$ group. The mean of activity in the SH group was greater than that in the $E_2(-)$ group, and was not different from that in the $E_2(+)$ group. The mean in the light phase of activity in the SH group was the greatest among groups; however, the value was not different between the $E_2(-)$ and $E_2(+)$ groups. The mean in the dark phase of activity in the SH group was greater than that in the $E_2(-)$ group, and was not different from that in the $E_2(+)$ group. The value in the $E_2(+)$ group was greater than that in the $E_2(-)$ group. The peak and amplitude of activity in the SH group was not different from that in the $E_2(-)$ group, and was greater than that in the $E_2(+)$ group. The value in the

E₂(+) group was greater than that in the E₂(-) group. The nadir of activity was not different among groups.

Mean in the day, light, and dark phase, amplitude, nadir, and peak of T _b (°C) and activity (counts) in the E ₂ (-), E ₂ (+), and SH groups				
		E ₂ (-)	E ₂ (+)	SH
T _b (°C)	Mean in a day	36.1 ± 0.0 [*]	36.0 ± 0.0	36.3 ± 0.0 ^{†, #}
	Mean in the light phase	35.7 ± 0.0 [*]	35.6 ± 0.0	35.7 ± 0.0 [†]
	Mean in the dark phase	36.5 ± 0.0 ^{*, §}	36.3 ± 0.0 [§]	36.9 ± 0.0 [§]
	Amplitude	1.5 ± 0.0 [*]	1.4 ± 0.0	2.2 ± 0.1
	Nadir	35.5 ± 0.0 [*]	35.5 ± 0.0	35.4 ± 0.0
	Peak	37.0 ± 0.0 [*]	36.8 ± 0.0	37.6 ± 0.0 ^{†, #}
Activity (counts)	Mean in a day	45 ± 1	49 ± 1 [*]	55 ± 1 [#]
	Mean in the light phase	24 ± 1	26 ± 1	29 ± 1 ^{†, #}
	Mean in the dark phase	67 ± 1 [§]	73 ± 3 ^{*, §}	80 ± 2 ^{#, §}
	Amplitude	99 ± 3 [*]	85 ± 2	97 ± 3 [†]
	Nadir	7 ± 1	8 ± 1	10 ± 1
	Peak	106 ± 3 [*]	93 ± 2	106 ± 2 [†]

Table 2: Mean in the day, light, and dark phase, amplitude, nadir, and peak of T_b (°C) and activity (counts) in the E₂(-), E₂(+), and SH groups. Values are mean ± SE (n=8 per group). Significant difference between E₂(-) and E₂(+) (*), E₂(-) and SH (#), E₂(+) and SH (†), and light and dark phases (§), P<0.05.

Table 3 shows the appearance time of the nadir and the peak of T_b and activity. The appearance time of the amplitude in T_b for the SH group was earlier than that in the E₂(-) group, and was not different from that in the E₂(+) group. The value in the E₂(+) group was earlier than that in the E₂(-) group. The appearance time of the nadir in T_b in the SH group was later than that in the E₂(-) group, and was not different from that in the E₂(+) group. The value in the E₂(+) group was later than that in the E₂(-) group. The appearance time of the peak in T_b in the SH group was earlier than that in the E₂(-) group, and was not different from that in the E₂(+) group. The value in the E₂(+) group

was earlier than that in the E₂(-) group. The appearance time of the amplitude in activity in the SH group was not different from that in the E₂(-) and E₂(+) groups. The value in the E₂(+) group was earlier than that in the E₂(-) group. The appearance time of the nadir in activity was not different among groups. The appearance time of the peak in activity in the SH group was earlier than that in the E₂(-) group, and was not different from that in the E₂(+) group. The appearance time of the peak in activity in the E₂(+) group was earlier than that in the E₂(-) group.

Appearance time of the nadir, peak of T _b and activity, and the amplitude in the E ₂ (-), E ₂ (+), and SH groups				
		E ₂ (-)	E ₂ (+)	SH
Appearance time of T _b (hours)	Amplitude	17.3 ± 0.8 ^{*, #}	10.6 ± 0.6	11.7 ± 0.7
	Nadir	4.3 ± 0.6	8.0 ± 0.4 [*]	6.9 ± 0.5 [#]
Appearance time of activity (hours)	Peak	21.6 ± 0.4 ^{*, #}	18.6 ± 0.4	18.7 ± 0.4
	Amplitude	19.3 ± 1.0 [*]	15.4 ± 0.9	16.4 ± 1.4
	Nadir	3.8 ± 0.9	4.8 ± 0.8	4.1 ± 0.8
	Peak	23.1 ± 0.2 ^{*, #}	20.02 ± 0.7	20.5 ± 0.9

Table 3: The amplitude, nadir, and peak of the appearance time of T_b and activity in the E₂(-), E₂(+), and SH groups. Values are mean ± SE (n=8 per group). Significant difference between E₂(-) and E₂(+) (*), E₂(-) and SH (#), E₂(+) and SH (†), P<0.05.

The body weight, food intake, and water intake are shown in Figure 3A-3C, respectively. Pre-surgery body weight was not different among

the groups; however, post-surgery body weight in the SH group was greater than that in the E₂(-) group. The value in the E₂(+) group was

lower than that in the E₂(-) group (Figure 3A). Pre- and post-surgery food intake and water intake were not different among groups (Figure 3B and 3C). Post-surgery food intake decreased from the pre-surgery food intake in all groups (Figure 3B). Water intake showed no difference between the pre- and post-surgery (Figure 3C). Plasma estradiol concentration in the E₂(+) group was higher than that in the E₂(-) and SH groups (62 ± 6 and 201 ± 60 pg/ml, respectively).

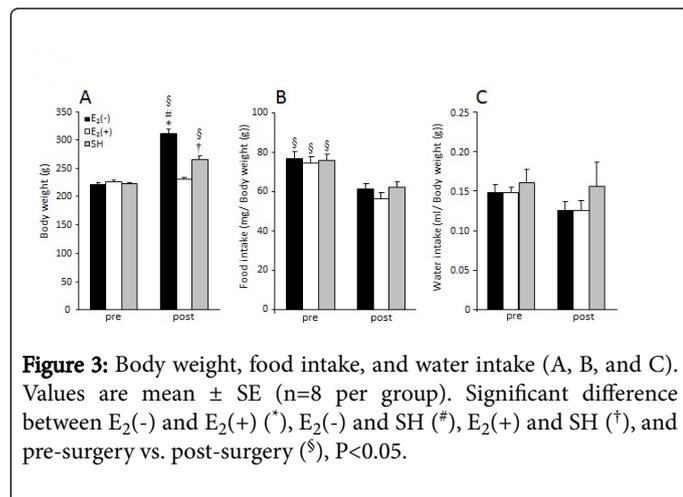


Figure 3: Body weight, food intake, and water intake (A, B, and C). Values are mean ± SE (n=8 per group). Significant difference between E₂(-) and E₂(+) (*), E₂(-) and SH (#), E₂(+) and SH (†), and pre-surgery vs. post-surgery (§), P<0.05.

Discussion

The present study showed that E₂ decreased the mean of T_b, and increased the mean of activity in the dark phase. E₂ affected the time-dependent relationship between T_b and activity. E₂ decreased the peak of T_b and activity, and delayed its appearance; however, E₂ did not influence the nadir of T_b and activity, but delayed their appearance. Thus, E₂ may modulate circadian rhythm of T_b and activity in female rats.

E₂ did not affect T_b in the light phase. This result coincides with those of a previous study showing that E₂ did not affect T_b in the light and dark phases [13]. Activity in the dark phase was decreased in ovariectomized mice [26] and rats [27]. The effect of E₂ on activity in the dark phase is controversial; E₂ increased it in the dark phase in mice [18,19], but did not influence it in the light and dark phases in rats [14,28]. The increased activity in the dark phase by E₂ coincided partly with the results of the previous studies.

In the E₂(+) group, the dependence of T_b on activity in the dark phase was lower than that in the light phase; however, it was not observed in the E₂(-) group (Table 1). Thus, E₂ may decrease the dependence of T_b on activity specifically in the dark phase. In literature, T_b is mainly determined by skin vasomotion and activity in rats at thermoneutral environments. E₂ decreased the T_{tail} in the dark phase at the thermoneutral range [14,28]. Vasoconstrictors like plasma adrenaline and arginine vasopressin have a circadian rhythm; higher in the light [29] and dark [30] phases, respectively. E₂ did not affect these [31,32]. Plasma renin activity and angiotensin related to the synthesis of vasoconstrictor angiotensin II were higher in the light phase in a day [33]. The influence of E₂ on angiotensin II is controversial as E₂ increased [34] or did not affect [35] it. Thus, it is difficult to assume that the vasoconstrictors induced the decreased T_{tail} by E₂ in the dark phase.

The circadian rhythm of peripheral vasodilators like endothelium-derived hyperpolarizing factor (EDHF) and endogenous hydrogen

sulfide (H₂S) is unknown yet. In the light phase, E₂ affected EDHF in the mesenteric artery in female rats [36,37] and H₂S production in the mesenteric artery in ewes [38]. Thus, E₂ may contribute to peripheral vasodilation rather than vasoconstriction through EDHF and H₂S in the light phase; however, the effect in the dark phase is unclear. E₂ is considered to affect sympathetic nerves because plasma norepinephrine fluctuated along with the estrus cycle in females [39]. E₂ may modulate the skin vasomotor circadian rhythm through vasodilators and sympathetic nerves. In summary, it was speculated that the dependence of T_b on activity was decreased due to strong skin vasomotion in the dark phase in the E₂(+) group.

The effect of E₂ on circadian T_b and activity rhythm is controversial; E₂ did not influence the circadian rhythm of T_b in female rats [28] and women [40], but increased the activity and amplitude, delayed their peaks, and advanced their onset [18]. E₂ decreased the peak of T_b and activity, advanced its appearance, and delayed the appearance of the nadir in T_b. The circadian rhythm of activity in mice administered with E₂ was similar to that in mice administered with E₂ α and β receptors agonists [18]. It was speculated that the E₂ α and β receptors are related to the alteration in T_b by E₂ in the present study, though a mechanism yet unknown.

The decreased body weight by E₂ coincided with that observed in previous studies [41-43]. E₂ administration did not affect the food intake per body weight. E₂ administration in ovariectomized rats decreased the food intake in a day [44] and in the light phase [41,42,45]. Food intake in previous studies was not calculated per body weight. This may influence a difference in the results between the present and previous studies.

The estrus cycle affects water intake and drinking behavior. Drinking behavior in the estrus phase was lesser than that in other phases [46]. Isoprenaline-induced water intake in the proestrus and estrus phases was lower than that in the other phases [47]. Ovariectomy in rats resulted in increased water intake [48]. E₂ administration decreased the water intake in female rats after water deprivation [49]. E₂ administration prolonged the onset of drinking behavior in female rats administered with NaCl solution [50]. Our result that E₂ decreased water intake coincided that of with previous studies. The result that E₂ did not influence water intake per body weight or food intake could not be compared with previous studies, because they were not calculated previously. The apparent water intake seemed to decrease due to the decreased body weight by E₂.

The present study showed that E₂ affected the time-dependent relationship between T_b and activity, and modulated the circadian rhythm of T_b and activity in female rats.

Acknowledgements

We are grateful to Prof. Kazuyoshi Tsutsui (Waseda University) and Prof. Keiko Morimoto (Nara Women's University) for their advice for this research. The present research was partially supported by the Ministry of Education, Science, Sports, and Culture; Grant-in-Aids for Scientific Research (B), No. 20390066; Grant-in-Aid for Research Activity, No. 24800047; Grant-in-Aid for challenging Exploratory Research, No. 16K13055; MEXT. KIBANKEISEI (2010); the Strategic Research Platforms for Private University; and Hayashi Memorial Foundation for Female Natural Scientists.

Author Contributions

KN supervised the entire project. YU and KN designed the study and wrote the manuscript. YU, SM and KT performed experiments.

References

1. Romanovsky AA, Ivanov AI, Shimansky YP (2002) Selected contribution: Ambient temperature for experiments in rats: A new method for determining the zone of thermal neutrality. *J Appl Physiol* 92: 2667-2679.
2. Shechter A, Varin F, Boivin DB (2010) Circadian variation of sleep during the follicular and luteal phases of the menstrual cycle. *Sleep* 33: 647-656.
3. Cagnacci A, Arangino S, Tuveri F, Paoletti AM, Volpe A (2002) Regulation of the 24h body temperature rhythm of women in luteal phase: Role of gonadal steroids and prostaglandins. *Chronobiol Int* 19: 721-730.
4. Baker FC, Waner JI, Vieira EF, Taylor SR, Driver HS, et al. (2001) Sleep and 24 hour body temperatures: A comparison in young men, naturally cycling women and women taking hormonal contraceptives. *J Physiol* 530: 565-574.
5. Shibui K, Uchiyama M, Okawa M, Kudo Y, Kim K, et al. (2000) Diurnal fluctuation of sleep propensity and hormonal secretion across the menstrual cycle. *Biol Psychiatry* 48: 1062-1068.
6. Shechter A, Lesperance P, Ng Ying Kin NM, Boivin DB (2012) Nocturnal polysomnographic sleep across the menstrual cycle in premenstrual dysphoric disorder. *Sleep Med* 13: 1071-1078.
7. Cagnacci A, Volpe A, Paoletti AM, Melis GB (1997) Regulation of the 24-hour rhythm of body temperature in menstrual cycles with spontaneous and gonadotropin-induced ovulation. *Fertil Steril* 68: 421-425.
8. Lee KA (1998) Circadian temperature rhythms in relation to menstrual cycle phase. *J Biol Rhythms* 3: 255-263.
9. Parry BL, LeVeau B, Mostofi N, Naham HC, Loving R, et al. (1997) Temperature circadian rhythms during the menstrual cycle and sleep deprivation in premenstrual dysphoric disorder and normal comparison subjects. *J Biol Rhythms* 12: 34-46.
10. Yochim JM, Spencer F (1976) Core temperature in the female rat: Effect of ovariectomy and induction of pseudopregnancy. *Am J Physiol* 231: 361-365.
11. Kent S, Hurd M, Satinoff E (1991) Interactions between body temperature and wheel running over the estrous cycle in rats. *Physiol Behav* 49: 1079-1084.
12. Rashotte ME, Ackert AM, Overton JM (2002) Ingestive behavior and body temperature during the ovarian cycle in normotensive and hypertensive rats. *Am J Physiol Regul Integr Comp Physiol* 282: R216-R225.
13. Williams H, Dacks PA, Rance NE (2010) An improved method for recording tail skin temperature in the rat reveals changes during the estrous cycle and effects of ovarian steroids. *Endocrinology* 151: 5389-5394.
14. Takezawa H, Hayashi H, Sano H, Saito H, Ebihara S (1994) Circadian and estrous cycle-dependent variations in blood pressure and heart rate in female rats. *Am J Physiol* 267: R1250-R1256.
15. Anantharaman-Barr HG, Decombaz J (1989) The effect of wheel running and the estrous cycle on energy expenditure in female rats. *Physiol Behav* 46: 259-263.
16. Baker FC, Selsick H, Driver HS, Taylor SR, Mitchell D (1998) Different nocturnal body temperatures and sleep with forced-air warming in men and in women taking hormonal contraceptives. *J Sleep Res* 7: 175-181.
17. Axelson JE, Zoller LC, Tomassone JE, Collins DC (1986) Effects of silastic progesterone implants on activity cycles and steroid levels in ovariectomized and intact female rats. *Physiol Behav* 38: 879-885.
18. Royston SE, Yasui N, Kondilis AG, Lord SV, Katzenellenbogen JA, et al. (2014) ESR1 and ESR2 differentially regulate daily and circadian activity rhythms in female mice. *Endocrinology* 155: 2613-2623.
19. Blattner MS, Mahoney MM (2014) Estrogen receptor 1 modulates circadian rhythms in adult female mice. *Chronobiol Int* 31: 637-644.
20. Blattner MS, Mahoney MM (2012) Circadian parameters are altered in two strains of mice with transgenic modifications of estrogen receptor subtype 1. *Genes Brain Behav* 11: 828-836.
21. Brockman R, Bunick D, Mahoney MM (2011) Estradiol deficiency during development modulates the expression of circadian and daily rhythms in male and female aromatase knockout mice. *Horm Behav* 60: 439-447.
22. Takahashi JS, Menaker M (1980) Interaction of estradiol and progesterone: Effects on circadian locomotor rhythm of female golden hamsters. *Am J Physiol* 239: R497-504.
23. Morin LP, Fitzgerald KM, Zucker I (1977) Estradiol shortens the period of hamster circadian rhythms. *Science* 196: 305-307.
24. Gerall AA, Napoli AM, Cooper UC (1973) Daily and hourly estrous running in intact, spayed and estrone implanted rats. *Physiol Behav* 10: 225-229.
25. Tsutsui K, Li D, Ukena K, Kikuchi M, Ishii S (1998) Developmental changes in galanin receptors in the quail oviduct and the effect of ovarian sex steroids on galanin receptor induction. *Endocrinology* 139: 4230-4236.
26. Sanchez-Alavez M, Alboni S, Conti B (2011) Sex- and age-specific differences in core body temperature of C57Bl/6 mice. *Age (Dordr)* 33: 89-99.
27. Izumo N, Ishibashi Y, Ohba M, Morikawa T, Manabe T (2012) Decreased voluntary activity and amygdala levels of serotonin and dopamine in ovariectomized rats. *Behav Brain Res* 227: 1-6.
28. Marui S, Nagashima K (2015) Reduction of plasma estradiol level affects daily rhythms of body core and tail skin temperature in female rats. *J Physiol Sci* 65 Supplement 1: S238.
29. De Boer SF, Van der Gugten J (1987) Daily variations in plasma noradrenaline, adrenaline and corticosterone concentrations in rats. *Physiol Behav* 40: 323-328.
30. Leal AM, Forsling ML, Moreira AC (1995) Diurnal variation of the pituitary-adrenal and AVP responses to stress in rats under food restriction. *Life Sci* 56: 191-198.
31. Mehrotra S, Gupta S, Villalon CM, Boomsma F, Saxena PR, et al. (2007) Rat carotid artery responses to alpha-adrenergic receptor agonists and 5-HT after ovariectomy and hormone replacement. *Headache* 47: 236-246.
32. Barron WM, Schreiber J, Lindheimer MD (1986) Effect of ovarian sex steroids on osmoregulation and vasopressin secretion in the rat. *Am J Physiol* 250: E352-E361.
33. Hilfenhaus M (1976) Circadian rhythm of the renin-angiotensin-aldosterone system in the rat. *Arch Toxicol* 36: 305-316.
34. Bell C, Bakhle YS (1975) Effects of chronic oral contraceptive treatment on the conversion of angiotensin I to angiotensin II in the rat. *J Pharmacol Exp Ther* 193: 160-165.
35. Gallagher PE, Li P, Lenhart JR, Chappell MC, Brosnihan KB (1999) Estrogen regulation of angiotensin-converting enzyme mRNA. *Hypertension* 33: 323-328.
36. Liu MY, Hattori Y, Sato A, Ichikawa R, Zhang XH, Sakuma I (2002) Ovariectomy attenuates hyperpolarization and relaxation mediated by endothelium-derived hyperpolarizing factor in female rat mesenteric artery: A concomitant decrease in connexin-43 expression. *J Cardiovasc Pharmacol* 40: 938-948.
37. Nawate S, Fukao M, Sakuma I, Soma T, Nagai K, et al. (2005) Reciprocal changes in endothelium-derived hyperpolarizing factor- and nitric oxide-system in the mesenteric artery of adult female rats following ovariectomy. *Br J Pharmacol* 144: 178-189.
38. Lechuga TJ, Zhang HH, Sheibani L, Karim M, Jia J, et al. (2015) Estrogen replacement therapy in ovariectomized nonpregnant ewes stimulates uterine artery hydrogen sulfide biosynthesis by selectively up-regulating cystathionine beta-synthase expression. *Endocrinology* 156: 2288-2298.
39. Goldstein DS, Levinson P, Keiser HR (1983) Plasma and urinary catecholamines during the human ovulatory cycle. *Am J Obstet Gynecol* 146: 824-829.

40. Gudmundsson A, Goodman B, Lent S, Barczy S, Grace A, et al. (1999) Effects of estrogen replacement therapy on the circadian rhythms of serum cortisol and body temperature in postmenopausal women. *Exp Gerontol* 34: 809-818.
41. Takamata A, Torii K, Miyake K, Morimoto K (2011) Chronic oestrogen replacement in ovariectomised rats attenuates food intake and augments c-fos expression in the suprachiasmatic nucleus specifically during the light phase. *Br J Nutr* 106: 1283-1289.
42. Asarian L, Yousefzadeh E, Silverman AJ, Silver R (2002) Stimuli from conspecifics influence brain mast cell population in male rats. *Horm Behav* 42: 1-12.
43. Geary N, Asarian L (1999) Cyclic estradiol treatment normalizes body weight and test meal size in ovariectomized rats. *Physiol Behav* 67: 141-147.
44. Santollo J, Eckel LA (2008) Estradiol decreases the orexigenic effect of neuropeptide Y, but not agouti-related protein, in ovariectomized rats. *Behav Brain Res* 191: 173-177.
45. Varma M, Chai JK, Meguid MM, Laviano A, Gleason JR (1999) Effect of estradiol and progesterone on daily rhythm in food intake and feeding patterns in fischer rats. *Physiol Behav* 68: 99-107.
46. Eckel LA, Houpt TA, Geary N (2000) Spontaneous meal patterns in female rats with and without access to running wheels. *Physiol Behav* 70: 397-405.
47. Findlay AL, Fitzsimons JT, Kucharczyk J (1979) Dependence of spontaneous and angiotensin-induced drinking in the rat upon the oestrous cycle and ovarian hormones. *J Endocrinol* 82: 215-225.
48. Tarttelin MF, Gorski RA (1971) Variations in food and water intake in the normal and acyclic female rat. *Physiol Behav* 7: 847-852.
49. Krause EG, Curtis KS, Davis LM, Stowe JR, Contreras RJ (2003) Estrogen influences stimulated water intake by ovariectomized female rats. *Physiol Behav* 79: 267-274.
50. Jones AB, Curtis KS (2009) Differential effects of estradiol on drinking by ovariectomized rats in response to hypertonic NaCl or isoproterenol: Implications for hyper- vs. hypo-osmotic stimuli for water intake. *Physiol Behav* 98: 421-426.