

Docking-Designed Green Synthesis and *In-vitro* Anticancer Studies of New Binuclear Se-N-Heterocyclic Carbene Adducts and their Azolium Salts

Muhammad Atif^{1*}, Mansoureh Nazari V², Mohamed B Khadeer Ahamed³, Aman Shah Abdul Majid⁴, Maryam Aslam⁵, Muhammad Adnan Iqbal¹

¹Department of Chemistry, University of Agriculture, Faisalabad, Pakistan; ²Department of Pharmacy, Universiti Sains Malaysia, Pulau Penang, Malaysia; ³Department of Pharmacology, Universiti Sains Malaysia, Pulau Penang, Malaysia; ⁴Department of Medicine, Quest International University Perak, Perak, Malaysia; ⁵Department of Chemistry, Government College Women University, Faisalabad, Pakistan

ABSTRACT

Two binuclear selenium adducts (5 and 6) were designed using molecular docking approach while finding their promising interaction to four angiogenic factor-proteins including COX-1 (Cyclooxygenase-1), VEGF-A (Vascular Endothelial Growth Factor A), HIF (Hypoxia-Inducible Factor) and EGF (Human Epidermal Growth Factor). They were consequently synthesized using *in-situ* coordination approach. The green synthetic approach was employed for coordination as it was carried out in water instead of organic solvents. The synthesized adducts as well as their respective bis-benzimidazolium salts (2 and 4) were confirmed by ¹H and ¹³C-NMR along with FT-IR spectroscopy. The both were, then, subjected to *in-vitro* anticancer activities against breast adenocarcinoma cell line (MCF-7), cervical cancer cell line (Hela), mouse melanoma cell line (B16F10) and Retinal Ganglion Cell line (RGC-5) using MTT assay while comparing their activities with a commercially established standard-drug 5-Fluorouracil. However, the exceptional activities of both adducts and bis-benzimidazolium salts were explored.

Key words: Selenium adducts; Binuclear benzimidazolium salts; Se-NHC; Molecular docking; Breast cancer (MCF-7); Cervical cancer (Hela); Retinal Ganglion Cancer (RGC-5)

INTRODUCTION

Selenium is a nutritional supplement and its analogues are biocompatible-therapeutics. Moreover, anti-infective potential of its adducts with organic compounds is widely reported in literature which is due to its probable bioavailability at the target site. Wherein its adducts get hydrolyzed to consequently produce ionized selenium inside cellular membrane. Its pattern of hydrolysis and ionization offer versatility of its functional role for different biological targets. For instance, m-tri-fluoromethyl-diphenyl diselenide is used to treat mechanical allodynia; Selenomethionine (SeMet) for human colorectal cancer and bis-selenide 3-(4-fluorophenylselenyl)-2,5-diphenylselenophene for depression-related behavior. Moreover, many selenium analogues are under clinical trials as ebselen, a selenium heterocyclic compound, is in phase II of clinical trials to treat the retrieval of

hearing loss and some types of cancers. So the modulated hydrolysis of selenium adducts with respect to biological target is often considered a subject of its activity [1].

A better modulation-approach implies its coordination with N-heterocyclic carbene analogues which can endow a potential permeability over a wide range of lipophilicity to better cross membrane barriers. In this way, it becomes capable to show chemo-preventive activities like antioxidant, anti-depressant, anti-neoplastic anti-inflammatory, anti-neuro-degenerative, antimicrobial and anti-viral activities. Initially, mononuclear selenium N-heterocyclic carbenes analogues were synthesized and tested in this pursuit. Recently, binuclear N-heterocyclic carbene analogue were synthesized to further advance its versatility in their stability and lipophilicity. But a little advancement was made in context. The present work was directed, not only, to

Correspondence to: Muhammad Atif, Department of Chemistry, University of Agriculture, Faisalabad, Pakistan; E-mail: ranaatif219@gmail.com

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synthesize some more binuclear Se-N-heterocyclic carbene adducts, but also to promote a green synthetic approach for their synthesis while making use water as reaction media instead of organic solvents [2].

MATERIALS AND METHODS

Designing of anticancer Se-NHC adduct

Molecular docking approach to design potential anticancer Se-NHC Adduct was employed wherein target protein preparation was made while downloading python language from www.python.com; BIOVIA from <http://accelrys.com>; Molecular Graphics Laboratory (MGL) from <http://mgltools.scripps.edu>; discovery studio visualizer 2017 from <http://accelrys.com>; chem3D from <https://acms.ucsd.edu>; and autodock 4.2 from <http://autodock.scripps.edu>. The selected software was employed to select three dimensional X-ray crystallographic structure of HIF, VEGF-A, EGF and COX1 proteins as target for designed chemical analogues [3]. The crystallographic structures were selected from Protein Data Bank (PDB) (www.rcsb.org/pdb) using PDB ID: 4KZN for VEGF-A; PDB ID: 1EQH for COX1; PDB ID: 1JL9 for EGF and PDB ID: 1YCI for HIF, their representations are shown in Figure 1. The study was resumed while deleting all non-essential entities like small molecules, water molecule, nonpolar hydrogens, heteroatoms, non-standard residues and lone pairs were deleted and adding hydrogens in target receptor molecule. Dock Prep was used to optimize geometric fitting and to evaluate minimum energy a built-in tool for structures preparation before docking employed.

Ligand preparation in docking was accomplished while selecting two designed compounds from crystallography with available with identified structure of salts were used by pubchem to create their sdf format which was changed to PDB format using Pymol. The starting structures were prepared using Autodock tools, water molecule was eliminated, polar hydrogen and Kollman charges were added to the protein starting structure. Grid box was set with the size of $126 \times 126 \times 126 \text{ \AA}$ with the grid spacing of 0.375 \AA at the binding site. The starting structure both salts namely V and VI were constructed using BIOVIA draw while Sunitinib and 5-flourouracil were selected as positive control getting their structures from Pubchem website Gasteiger. The optimized ligands were assigned with charges using Autodock tools. 100 docking runs were monitored with crossover rate of 0.8 and mutation rate of 0.02. Moreover, 250 randomly placed individual was the population size. Lamarckian genetic algorithm was used in searching algorithm with a translational step of 0.2 \AA , a quaternion step of 5 \AA and a torsion step of 5 \AA [4].

Synthesis of designed analogues

All reagents and chemicals including benzimidazole as precursor; Dimethyl Sulfoxide (DMSO) and 1,4-dioxane as reaction media; o-xylene dibromide, n-hexyl and n-octyl bromide as alkylating agents; selenium powder for coordination were supplied by sigma aldrich and they were used as received without any sort of purification unless otherwise stated. All

reactions were accomplished in ordinary laboratory conditions. Bisbenzimidazolium salts (2 and 4) synthesized by their respective N-alkylation of benzimidazole. Selenium N-heterocyclic carbene adducts (5 and 6) were synthesized by *in-situ* approach according to reported procedures as shown in scheme-1. Melting points of synthesized compounds by Stuart melting point method SMP11 [5]. Their spectroscopic confirmation was made by Fourier transform infrared spectroscopic analysis using Perkin-Elmer 2000 spectrophotometer; $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$ analysis by Bruker advance 500. For the assessment of *in-vitro* anticancer activities, reagents for cell culturing, HIFBS (Heat Inactivated Foetal Bovine Serum) and trypsin, dulbecco's modified were supplied by Gibco, USA; eagle and RPMI 1640 medium and were purchased from Gibco, UK. All cell-lines including Hela, MCF-7, RGC-5 and B16F10 were purchased from ATCC, USA and were used as per (Figure 1).

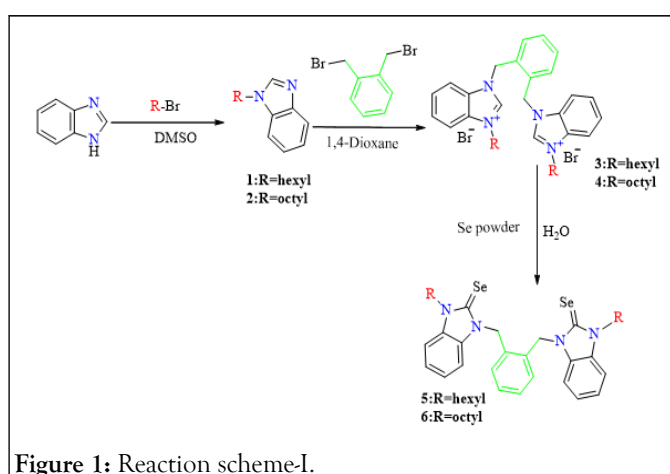


Figure 1: Reaction scheme-I.

Synthesis of binuclear benzimidazolium salts

Synthesis of 2 (1-hexyl-3-(2-((3-hexyl-1H-benzo[d]imidazol-3-ium-1-yl)methyl) benzyl)-1H-benzo[d]imidazol-3-ium bromide): Firstly, 1-hexyl-1H-benzo[d]imidazole (compound 1) was prepared while treating benzimidazole (1 g, 8.46 mmol), KOH (0.71 g, 12.40 mmol) and hexyl bromide (1.19 mL, 8.46 mmol) in the presence of (20 mL) Dimethyl Sulfoxide (DMSO) as a solvent stirred for three hours at room temperature which was extracted by water-chloroform system to consequently dried with a yield 94.4% in form of thick yellowish fluid. Compound 1 was again subjected to further N-alkylation by o-xylene dibromide to produce 1-hexyl-3-(2-((3-hexyl-1H-benzo[d]imidazol-3-ium-1-yl)methyl) benzyl)-1H-benzo[d]imidazol-3-ium bromide (compound 2) wherein it was treated with (2.2 g, 7.9 mmol) of ortho-xylene dibromide in 25 mL of 1,4-dioxane under reflux for 24 hours. Slight creamy precipitates were emerged which were isolated by decantation and were washed with fresh 1,4-dioxane and dried getting a yield of 75% with melting point 246°C . The product was consequently confirmed as: FT-IR (KBr, cm^{-1}): 3410, 3390 ($\text{C}_{\text{aliph}}\text{-N}_{\text{benzimi}}$), 3080, 3030 (C-H_{arom}), 2947, 2825, 2900, 2890 ($\text{C-H}_{\text{aliph}}$), 1605, 1573, 1517 ($\text{C}_{\text{arom}}\text{-C}_{\text{arom}}$), 1214, 1407, 1417, 1426 ($\text{C}_{\text{arom}}\text{-N}_{\text{benzimi}}$). $^1\text{HNMR}$ (500 MHz, $\text{d}_6\text{-DMSO}$) δ ppm: 0.85 (t, $J=7.0 \text{ Hz}$, 6H, $2 \times \text{CH}_3$), 1.3-1.4 (br.m, 10H, $5 \times \text{CH}_2$), 3.5 (s, 4H, $2 \times \text{N-CH}_2\text{-Ar}$), 4.51 (t, $J=7.0 \text{ Hz}$, 4H, $2 \times \text{NCH}_2\text{-R}$), 7.20 (q, 2H, Ar $2 \times \text{CH}$), 7.37 (q, 2H, Ar $2 \times \text{CH}$), 7.64 (t, $J=7.5 \text{ Hz}$, 2H, Ar $2 \times \text{CH}$), 7.66 (t, $J=8.0 \text{ Hz}$, 2H, Ar $2 \times$

CH), 8.1 (d, J=8.0 Hz, 2H, Ar 2 × CH), 9.89 (s, 2H, 2 × NCHN); ¹³C (1H) NMR (125.5 MHz, DMSO-d₆) δ ppm: 14.6 (CH₃), 22.4, 25.9, 29.01, 39.9 (5 × CH₂), 47.3 (R-CH₂-N), 48.05 (Ar-CH₂-N), 114.6-132.4 (Ar-C) and 139.8 (NCN). Anal. Cal. For: C₃₄H₄₅Br₂N₄O: C, 80.27; H, 8.72; N, 11.1%. Found: C, 77.2; H, 7.5; N, 10.17%.

Synthesis of 4 (1-octyl-3-(2-((3-octyl-1H-benzo[d]imidazol-3-ium-1-yl)methyl)benzyl)-1H-benzo[d]imidazol-3-ium bromide):

The synthesis of compound 4 was followed by its retrosynthesis in same way as that of compound-2 only the difference was the reaction of n-octyl bromide with benzimidazole to get compound 3. Then 3 was reacted with equimolar ortho-xylene dibromide in same to get 4 which was prepared with 90 percent yield and was appeared in form of white crystalline powder with melting point 204°C. FT-IR (KBr, ν_{max}, cm⁻¹): 3403, 3378 (C_{aliph}-N_{benzimi}); 3085, 3055 (C-H_{arom}); 2983, 2894, 2813 (C-H_{aliph}); 1600, 1542 (C-H_{aliph}); 1173, 1406, 1423, 1488 (C_{arom}-N_{benzimi}). ¹H NMR (500 MHz, d₆-DMSO) δ ppm: 0.83 (t, J=7.0 Hz, 6H, 2 × CH₃), 1.2-1.35 (br.m, 20H, 10 × CH₂), 1.88 (qnt, J=7.5 Hz, 4H, 2 × CH₂), 4.56 (t, J=7.5 Hz, 4H, 2 × CH₂-N), 6.23 (s, 4H, 2 × Ar-CH₂-N), 7.1 (q, 2H, Ar 2 × CH), 7.43 (q, 2H, Ar 2 × CH), 7.64 (t, J=7.5 Hz, 2H, Ar 2 × CH), 7.7 (t, J=7.7 Hz, 2H, Ar 2 × CH), 8.03 (d, J=8.3 Hz, 2H, Ar 2 × CH), 8.2 (d, J=8.5 Hz, 2H, Ar 2 × CH), 10.12 (s, 2H, 2 × NCHN); ¹³C (1H) NMR (125.5 MHz, DMSO-d₆) δ ppm: 14.11 (CH₃), 21.2, 26.1, 25.8 (3 × CH₂), 29.06 (t, JC_{4/5}=40 Hz, JC_{6/7}=25 Hz, 3 × CH₂), 47.17 (R-CH₂-N), 47.84 (Ar-CH₂-N), 113-132.47 (Ar-C) and 143.12 (NCN). Anal. Cal. For: C₃₈H₅₄Br₂N₄O: C, 61.54; H, 7.20; N, 7.55%. Found: C, 61.43; H, 7.17; N, 7.46%.

Synthesis of binuclear Se-N-Heterocyclic Carbene (Se-NHC) compounds

Se-NHCs were synthesized using *in-situ* synthetic approach wherein 2 and 4 were coordinated to elemental selenium almost simultaneously after the production of N-heterocyclic carbene ligand from bis-benzimidazolium salts in alkaline aqueous conditions as per reported procedure wherein both of 5 and 6 adducts were synthesized by 3-5 hour stirring in reflux apparatus. When the products in form of insoluble mass along with unreacted dispersed-selenium particles started sticking to the walls of reaction flask and when there was no more aggregation of insoluble mass was observed the reaction was stopped. The hot reaction mixture was passed through the filtration apparatus, the insoluble mass (crude-product) retained on filter paper which was washed with fresh hot water several times so as to remove all of unreacted ligand and alkali molecules. When all of water was evaporated from product-contain filtration apparatus, fresh DMSO was poured so as to solubilize and pass the pure selenium adduct from filter paper to flask.

Synthesis of 5 (3,3'-(1,2-Phenylenebis(methylene))bis(1-hexyl-1H-benzo[d]imidazole-2(3H)-selenone): Adduct-5 was obtained with 78% yield in form of colorless to yellowish thick fluid and it was confirmed as FT-IR (KBr, ν_{max}, cm⁻¹): 3050, 3026 (C-H_{arom} stretch), 2918, 2910 (C-H_{aliph} stretch) 1700, 1612, 1588 (C=C_{arom} stretch), 1478, 1444, 1408, 1340 (CH₂-bending). ¹H NMR (500 MHz, d₆-DMSO) δ ppm: 0.83, 0.84 (t,

J=7.5 Hz, 6H, 2 × CH₃), 1.08-1.69 (20H, br.m, 10 × CH₂), 1.91 (qnt, 4H, 2 × CH₂), 3.34 (t, J=7.5 Hz, 4H, 2 × N-CH₂-R), 5.41, 5.26 (s, 4H, 2 × N-CH₂-Ar), 7.08 (s, 4H, Ar-H), 7.05 (sext, 4H, Ar-H), 7.03 (d, J=8.0 Hz, 2H, Ar-H), 8.06 (d, J=8.0 Hz, 4H, Ar-H), 8.16 (s, 2H, 2 × NCHN); ¹³C (1H) NMR (125.75 MHz, d₆-DMSO) δ ppm: 16.5 (CH₃), 22.50, 27.89 (CH₂), 28.15, 28.23, 28.27 (JC_{4/5}=13.9 Hz, JC_{5/6}=2.5 Hz), 31.11 (CH₂), 46.58 (N-CH₂-R), 48.43 (N-CH₂-Ar), 114.37 (d, J=11.2 Hz, Ar-C), 121.33 (J=2.5 Hz), 127.65, 129.40, 131.77, 134.76 (Ar-C), 153 (C-Se). Anal. Calcd: C, 61.44; H, 6.37; N, 8.43%. Found: C, 59.21; H, 5.87; N, 6.89%.

Synthesis of 6 (3,3'-(1,2-Phenylenebis(methylene))bis(1-octyl-1H-benzo[d]imidazole-2(3H)-selenone):

Compound 6 was also synthesized as per general given methodology, it was appeared with 78% yield in form of light yellowish sticky fluid. FT-IR (KBr, ν_{max}, cm⁻¹): 3053, 3023 (C-H_{arom} stretch), 2921, 2907 (C-H_{aliph} stretch) 1703, 1609, 1591 (C=C_{arom} stretch), 1481, 1441, 1411, 1337 (CH₂-bending). ¹H NMR (500 MHz, DMSO-d₆) δ ppm: 0.811, 0.825 (t, J=7.5 Hz, 6H, 2 × CH₃), 1.081-1.669 (28H, br.m, 10 × CH₂), 1.922 (qnt, 4H, 2 × CH₂), 3.46 (t, J=7.5 Hz, 4H, 2 × N-CH₂-R), 5.22, 5.28 (s, 4H, 2 × N-CH₂-Ar), 7.008 (s, 4H, Ar-H), 7.045 (sext, 4H, Ar-H), 7.063 (d, J=8.0 Hz, 2H, Ar-H), 8.046 (d, J=8.0 Hz, 4H, Ar-H), 8.16 (s, 2H, 2 × NCHN); ¹³C NMR (125.75 MHz, DMSO-d₆) δ ppm: 14.31 (CH₃), 22.53, 26.33 (CH₂), 27.56, 27.93, 28.21 (JC_{4/5}=13.9 Hz, JC_{5/6}=2.5 Hz), 31.66 (CH₂), 42.49 (N-CH₂-R), 48.18 (N-CH₂-Ar), 114.29 (d, J=11.2 Hz, Ar-C), 127.09 (J=2.5 Hz), 128.6, 129.2, 131.6, 135.6 (Ar-C), 163.63 (C-Se). Anal. Calcd: C, 63.32; H, 6.99; N, 7.77%. Found: C, 60.21; H, 6.17; N, 6.9%.

Anticancer studies

Preparation of cell culture: Initially, B16F10, RGC-5, MCF-7 and Hela cells were harbored and grown under incubated environment. The cell achieving 75%-80% confluency were selected for cell plating. After discarding the old medium, the cells were washed thrice using phosphate-buffer saline of pH 7.4 then the buffer was also washed completely. Then trypsin was added in way to evenly immerse the cell thoroughly and they were incubated in 5% CO₂ for 1 minute at 310 K. Then the cells were cautiously tapped so as to promote cell segregation which was monitored under inverted microscope. Then freshly prepared 10% Fetal Bovine Serum (FBS) was inoculated upto 5 mL as as to explore trypsin activity. Finally, 100 mL of cell media having concentration 2.5 × 10⁵ cells per one mL per well was added and incubated with 5% CO₂ as internal environment at 37°C.

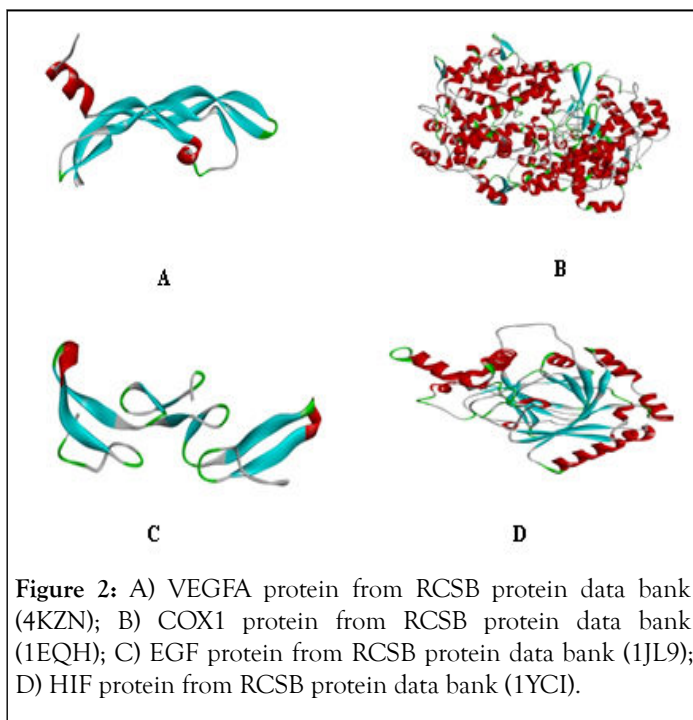
MTT assay: MTT assay was accomplished as per our reported methodology.

RESULTS AND DISCUSSION

Molecular docking

The activities of synthesized analogues were presumed while assessing their anti-angiogenetic potential wherein the potential of each analogue to inhibit four angiogenic factors including

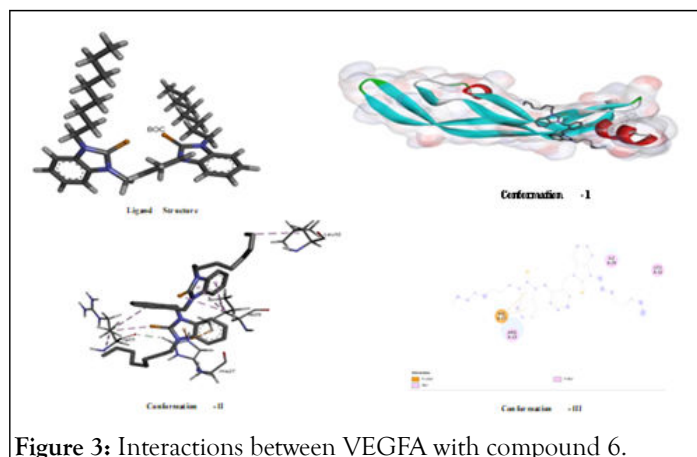
COX-1 (Cyclooxygenase-1), EGF (Human Epidermal Growth Factor), VEGF-A (Vascular Endothelial Growth Factor A) and HIF (Hypoxia-Inducible Factor) was studied (Figure 2). All of four factor-proteins are actively involved in the supply of oxygen and nutrients to cancer cells. The binding affinities and ligand efficiency of 5 and 6 was assessed, taking 5-fluorouracil and sunitinib as standard reference [6].



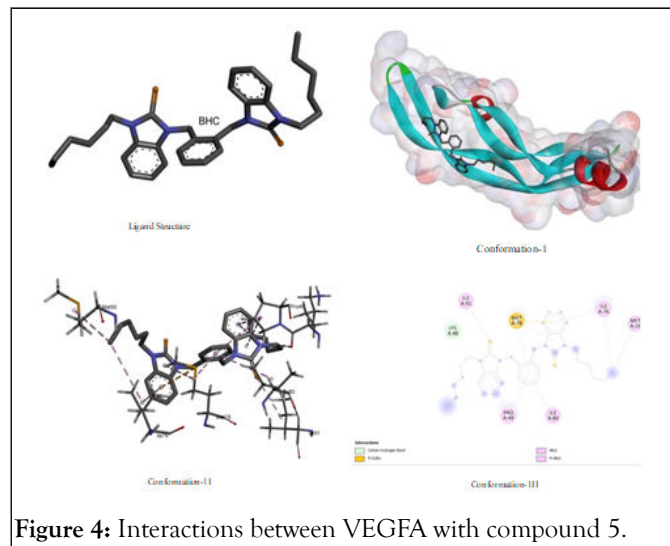
The docked conformation of synthesized Se-NHCs binding to block tested oncogenic proteins were studied focusing side-chain interactions wherein short chain electrostatic interactions specially are considered.

Docking interactions of test compounds with VEGFA

The interactions between VEGFA with compound 6: The docking study of 6 were also fascinating interactions to VEGFA, offering different types of binding sites. It has potential binding interaction wherein alkyl residues of arginine (Arg A:23), isoleucine (Ile A:29) and leucine (Leu A:32) interact with octyl chain and xylyl core; hydrogen-atom of histidine (His A:27) residues interact with benzimidazolium portion as per pictorial diagram given in Figure 3. In view of its promising interaction, it shows very low binding energy value (-6) demonstrating its higher stability of its supposed complexation to inhibition VEGFA [7].



The interactions between VEGFA with compound 5: Compound 5 also showed significant interaction for VEGFA with alkyl residues of (Met A:55), (Ile A:76), (Ile A:91) and (Pro A:49) of VEGFA. Moreover, its potential to interact with carbon-hydrogen bond of (Lys A:48) and with pi-sulfur bond of (Met A:78) is also significant. It showed good to better interaction for having binding energy value -4.66 compared to -6 value of sunitinib (positive control) which provokes for its synthesis to test their *in-vitro* and *in-vivo* trials. The anticancer trials while inhibiting VEGFA through multilateral geometric fitting as shown in Figure 4. As isoleucine has interaction with selenium portion; with benzilium ring of xyllyl; methionine has interaction also with xyllyl of benzimidazolium as well as benzimidazole portion through its sulfur residues.



The interactions between VEGFA with Fluorouracil (FU): Fluorouracil was taken as positive control which is well recognized anticancer standard drugs for comparative anticancer studies. Due to presence of a number of polar groups in its structural skeleton, it was found an interactive compound for its hot-spot binding with various functionalities especially with oxygen-hydrogen bond of (Phe A:38), (Ser A:50) and (Asp A:34) and carbon hydrogen bond of (Ile A:46) of VEGFA as shown in Figure 5.

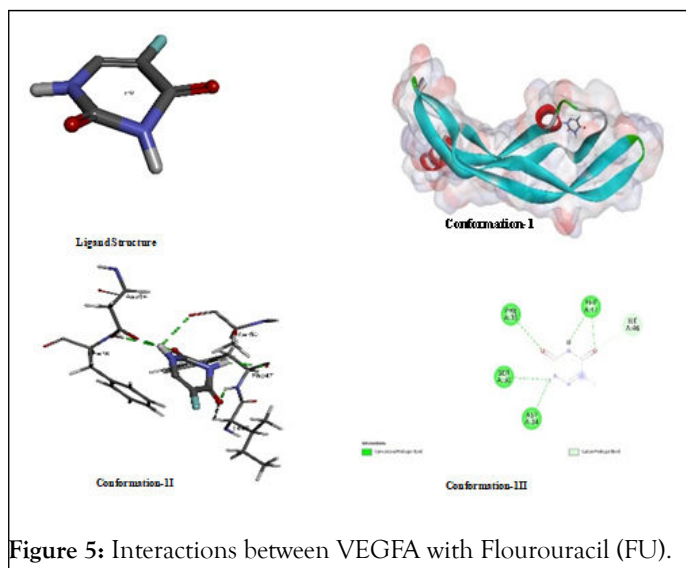


Figure 5: Interactions between VEGFA with Flourouracil (FU).

The interactions between VEGFA with sunitinib: Sunitinib offer its potential binding interaction with VEGFA which justifies its proven and reported potential as multi-kinase and angiogenesis inhibitor. The conformational view in the pictorial diagram shows that it has binding affinity with both of non-polar and polar amino acids in peptide backbone of VEGFA as it interacts with carbon hydrogen bond of glutamic acid (Glu A: 35) and aspartic acid (Asp A:11) which are polar amino acids and with the same of proline (Pro A:40) and tyrosine (Tyr A:39) which are often considered non-polar amino acids. Furthermore, it is also interactive with hydrogen of fluorine of Serine (Ser A:50) as shown in Figure 6.

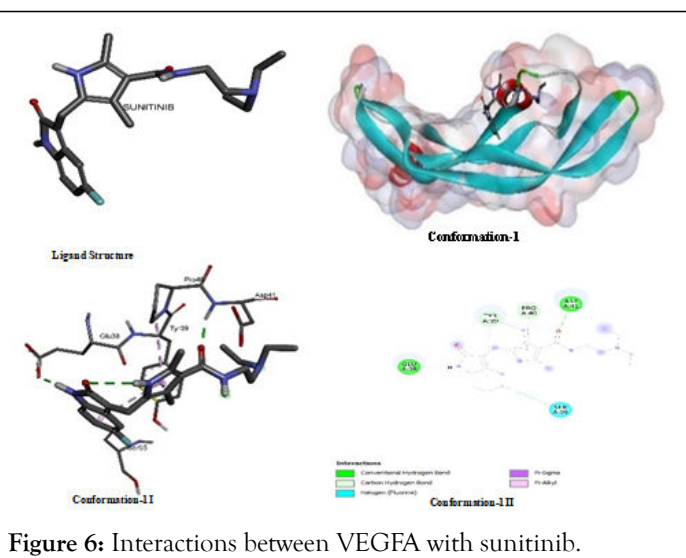


Figure 6: Interactions between VEGFA with sunitinib.

Docking interactions of test compounds with COX1

The interactions between COX1 and compound 5: The results after the assessment of interaction potential for 5 shows recurrence of previous trend as it showed with VEGFA. It was found a better and effective analogue among Se-NHC having alkyl groups as showed binding energy value (-7.82) as compare to (-7.32) of compound 6 which is more negative and consequently found more active anticancer as shown in Figure 7. And it has also been inferred that hexyl is among the most suitable ligands containing alkyl substituents. If we see it

conformational diagram in the given pictorial diagram it will be justified that it has pi-sigma, alkyl-alkyl and carbon-hydrogen interactions as major key ligating agents to inhibit COX1 while interacting with (Pro A:542); (Glu A:543); (Tyr A:64); and of (Ile A:43).

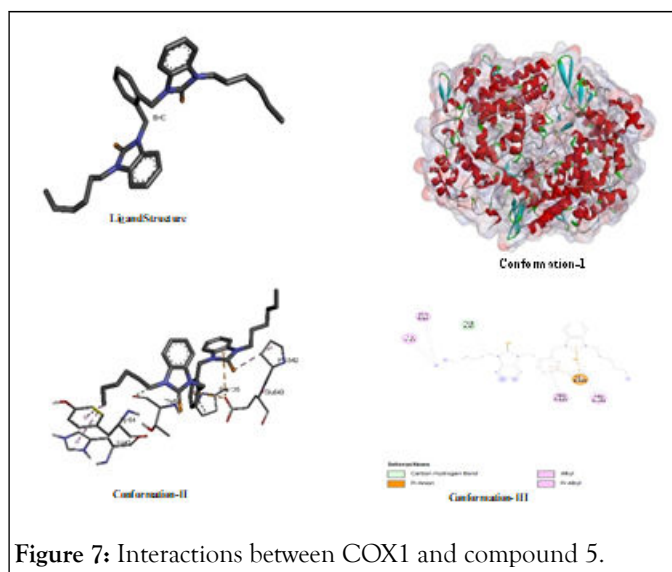


Figure 7: Interactions between COX1 and compound 5.

The interactions between COX1 and compound 6: In the assessment of COX1 interaction for 6, it was found that it showed its highest interaction potential to COX1 as compare to its interactions with other growth factor proteins like VEGF, EGF and HIF as shown in Figure 8 it shows higher stability of its probable complex with COX1 on basis of its lower binding energy is lower (-7.32) than that of positive control FU (-4.92) [8]. It shows interaction while interacting with alkyl groups, pi-bond and others of (His A: 226); (Phe A:142); (Val A:145) and (Ser A:143).

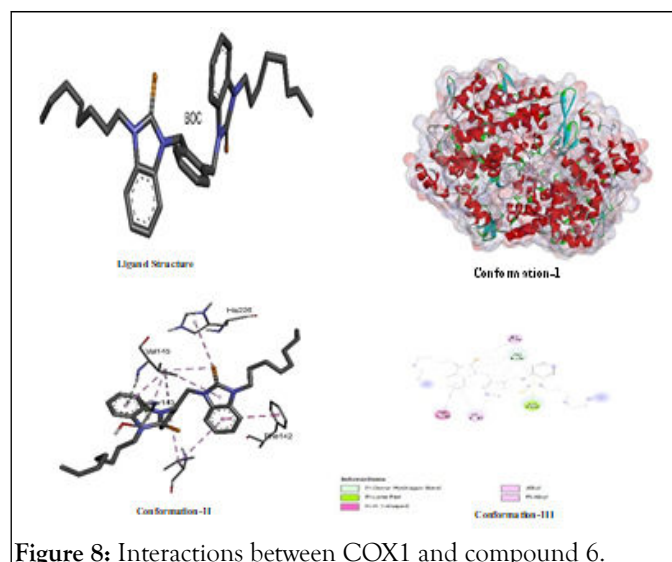


Figure 8: Interactions between COX1 and compound 6.

The interactions between COX1 and FU: The motif of versatile activity in blocking oncogenic proteins for flourourcil is admissible in docking analysis while presenting Figure 9 to demonstrate its binding to inhibit cyclooxygenase enzyme-1 (COX-1) wherein promising degree of binding due to its polar groups like oxygen atom and acidic hydrogen atom was noted. Polar atoms showed their electrostatic interaction especially by

hydrogen bonding or carbon hydrogen bond or others with (Thr A:331); (Thr A:549); (Ser A:545); and (Ile A:137).

Docking interactions of test compounds with HIF

The interactions between HIF and compound 5: HIF inhibition studies, 5 was considered a one of the most effective inhibitor showing its binding energy value (-6.34) which was very close that of positive control (sunitinib). It showed promising activity on the basis of its probable interactions like: pi-sigma, pi-alkyl, alkyl-alkyl and others. The major interacting amino acids were histidine, isoleucine, Phenylalanine, glutamic acid residues which were detected spatially as (Phe A:111); (His A:234); (Pro A:21); (Val A:336); (Ile A:222); (Ile A:344); (Llu A:340); and (Glu A:325) as shown in Figure 11. Its higher interaction paves a way for various types of structural changes to develop new derivative-analogues of this compound in view of its geometric fitting.

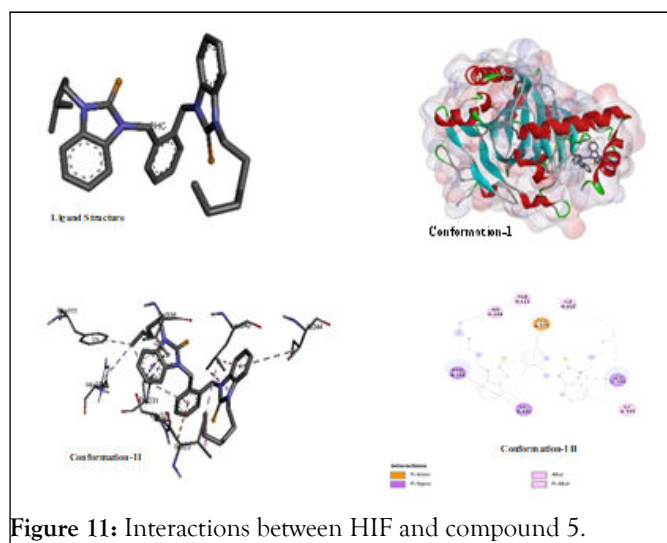


Figure 11: Interactions between HIF and compound 5.

The interactions between HIF and compound 6: Compound 6 demonstrated a binding energy value (-5.56) which was also very close to the highly active anticancer positive control (sunitinib). But the value was the lowest among all other test compounds but it was higher than fluorouracil. In the given pictorial diagram its interactions like: pi-alkyl, pi-sigma, alkyl-alkyl and others interactions targeting residues of different amino acids like threonine, glutamic acid, phenylalanine and leucine which were spatially as (Phe A:111); (Ile 322); Glu A:323); (Pro A:231); (Val A:338); and (Leu A:340) as shown in Figure 12.

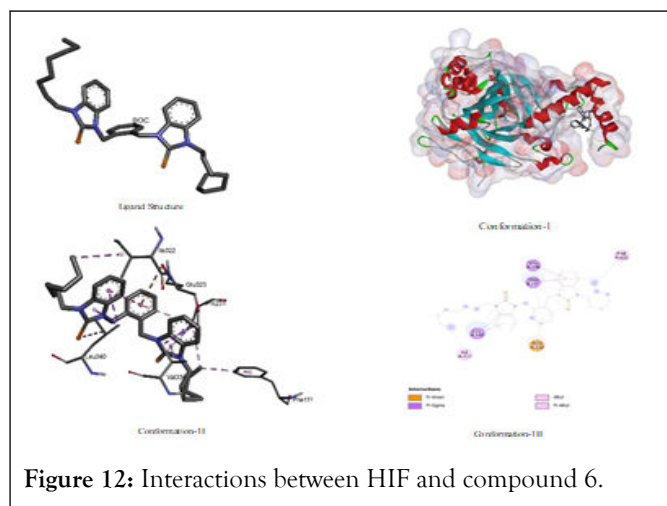


Figure 12: Interactions between HIF and compound 6.

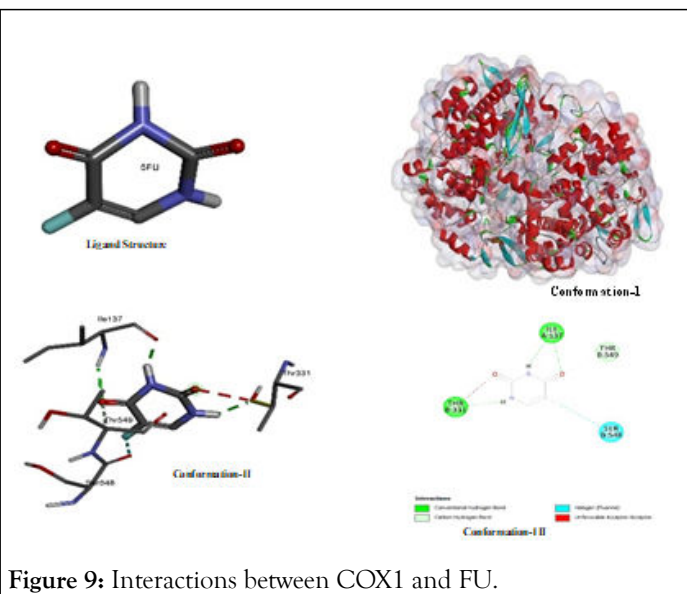


Figure 9: Interactions between COX1 and FU.

The interactions between COX1 and sunitinib: The reported interaction of sunitinib with COX1 again found promising as it was found in case of VEGFA. Its potential interaction with various amino acid residues especially with (Pro A:158); (Asp A:135); (Ile A:46); (Cys A:42); (Gln A:161); (Gln A:44); (Glu A:465); (Arg A:469); and (Leu A:152) shows its higher affinity to the both polar and non-polar amino acids as shown in Figure 10. In view of its interaction with amino acids of all types of polarities, it ligates through hydrogen bonding, pi-sigma interaction, pi-pi interaction, van der Waal interaction and others. It showed lowest binding energy value (-8.87) among all test ligands. Selective cyclooxygenase-1 inhibitors suppress cell proliferation and induces apoptosis in cancer cell lines. Inhibition of COX 1 leads to mitochondrial dysfunction and triggered caspase dependent mitochondrial apoptosis by inhibiting the NF κB pathway. There are established evidences that cyclooxygenase-1 expression appears commonly in breast, cervical and melanoma cancer cell lines.

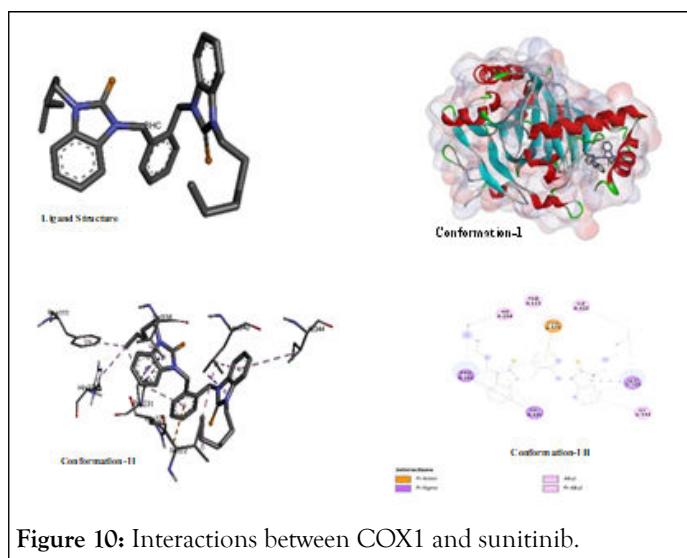


Figure 10: Interactions between COX1 and sunitinib.

The interactions between HIF and sunitinib: Higher binding interaction testified our selection of sunitinib as positive control for this study. In case of interactional study with hypoxia inducible factor protein, it showed a binding energy value (-7.4) which extremely negative to offer greater inhibition potential. It was found that HIF interacts with various types of amino acid residues of both polar and less polar nature through hydrogen bonding and van der Waal interaction as shown in Figure 13 wherein it showed consistent affinity to arginine, threonine, serine, glycine and alanine which are spatially as (Arg A:33); (Ser A:34); (Arg A:215); (Phe A:37); (Thr A: 39); (Ala A:10); (Ser A: 11); and (Gly A:14).

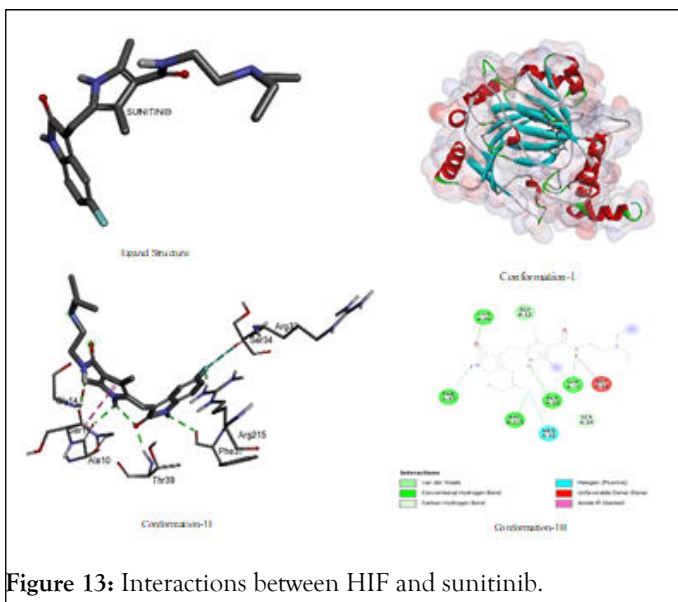


Figure 13: Interactions between HIF and sunitinib.

The interactions between HIF and FU: In spite of its proven and versatile efficiency of fluorouracil but in case of scavenging HIF protein, it was not found an effective analogue because of its higher binding energy value (-4.68) among all other test ligands. It might be due to its higher polarity which might not afford geometric fitting in non-polar environment of amino acid residues but it has, binding interaction with (Leu A:101); (Ser A: 118); (Gln A:147) through hydrogen bonding, carbon hydrogen bond and its fluorine linkages shown in Figure 14.

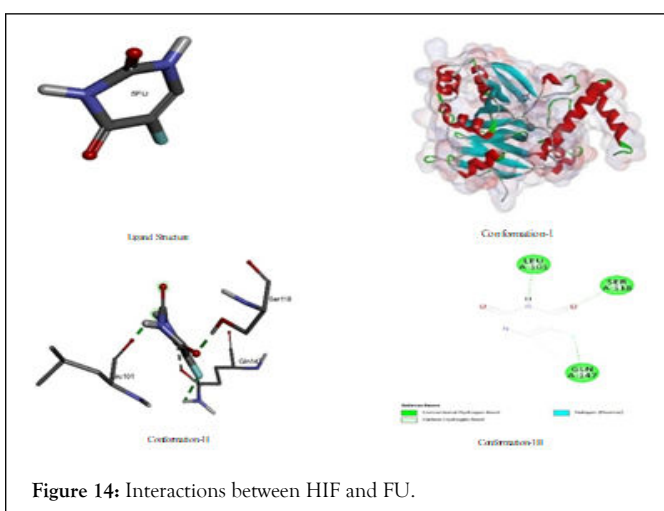


Figure 14: Interactions between HIF and FU.

Docking interactions of test compounds with EGF

The interactions between EGF with compound 5: Interaction of EGF with 5 was also fascinating and was better than that of 6 but it was not much closer to the that of positive control as 5 has a value (-5.08) compared to (-7.09) of sunitinib. As the both 5 and 6 have almost similar binding energy value which might be ascribed to the presence of non-polar alkyl chains attached on nitrogen atoms which show a lack of binding potential in elaborated in Figure 15.

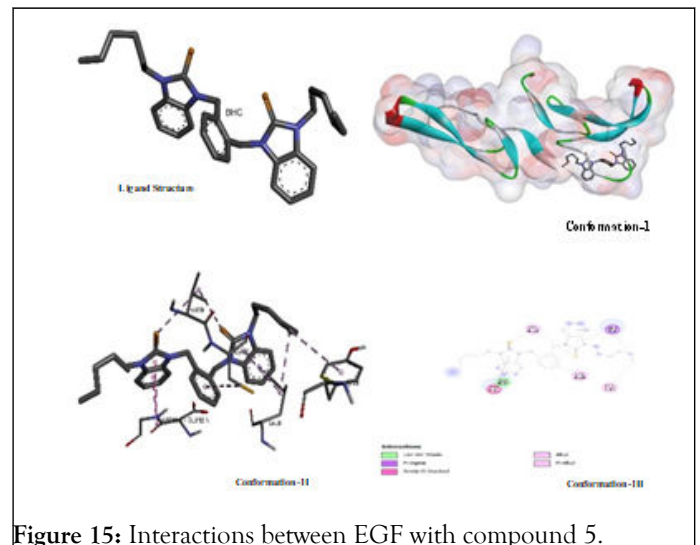


Figure 15: Interactions between EGF with compound 5.

The interactions between EGF with compound 6: In case of 6, all docked conformations were interactive to block Epidermal Growth Factor-A (EGF) showing a binding energy value (-4.36) which higher than that of 5 as well as than that of two studied positive controls that reveals its lower stability after binding with EGF [9]. However, it has shown multiple type of binding sites originating from interacts with benzilium of xyllyl portion and from various structural appendages while targeting various amino acid residues of (Cys A:14); (Cys A:20); (Asp A: 11); (Cys A:8); (Asp A:17); and (Val A:26) as shown in Figure 16.

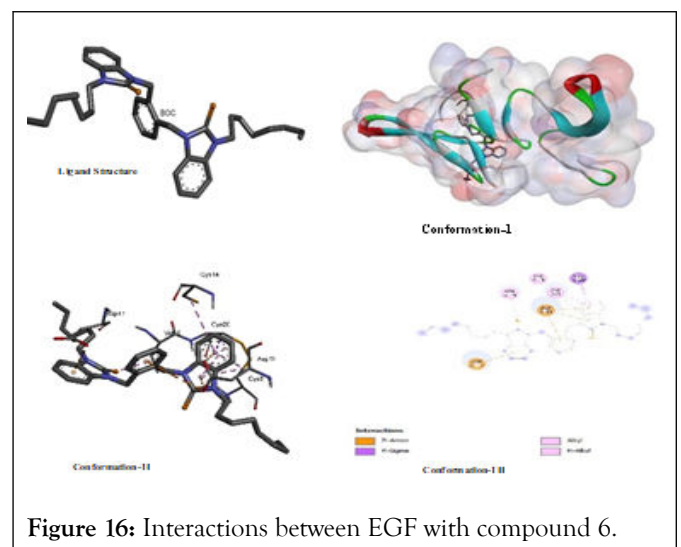


Figure 16: Interactions between EGF with compound 6.

The interactions between EGF with FU: Fluorouracil has long been known as an active cytotoxic agent for cancer cells and this fact has been testified again in our study. In given pictorial diagram, almost all polar groups are showing pronounced interaction with different amino acids residues of EGF which include: Cysteine, tyrosine, proline, aspartic acid, leucine and glycine which are spatially as (Pro A:7); (Cys A:12); (Tyr A:13); (Gly A:12); (Cys A:20); (Leu A:8); and (Asp A:1) as elaborated in Figure 17. All amino acids are interacting through hydrogen bonding or other strong electrostatic interaction using their polar groups to associate either with electronegative nitrogen/oxygen atom or electropositive hydrogen atom [10]. Due this reason, it has higher stability of its complex with EGF-protein.

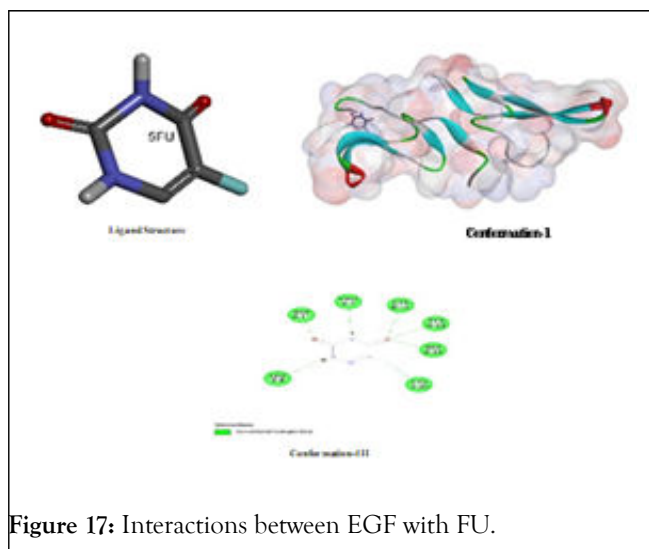


Figure 17: Interactions between EGF with FU.

The interactions between EGF with sunitinib: After fluorouracil, sunitinib showed lowest binding energy value to demonstrate highest stability of its probable complex with EGF that is likely due its higher degree of interactions with multiple

types of amino acid residues present in the target protein as shown in Figure 18. Its main target amino acids are cysteine, glycine, tyrosine, aspartic acid, leucine and histidine shown in Figure 18. Its polar and non-polar groups both showed multiple types of binding interactions including van der waal, hydrogen bonding, pi-alkyl, alkyl-alkyl, pi-ionic pi-anionic, pi-sigma interactions with (Cys A:14); (Cys A:20); (Cys A:6); (Tyr A:13); (Gly A:18); (Leu A:8); and (Asp A:11) might be due to the presence of its both polar and non-polar groups.

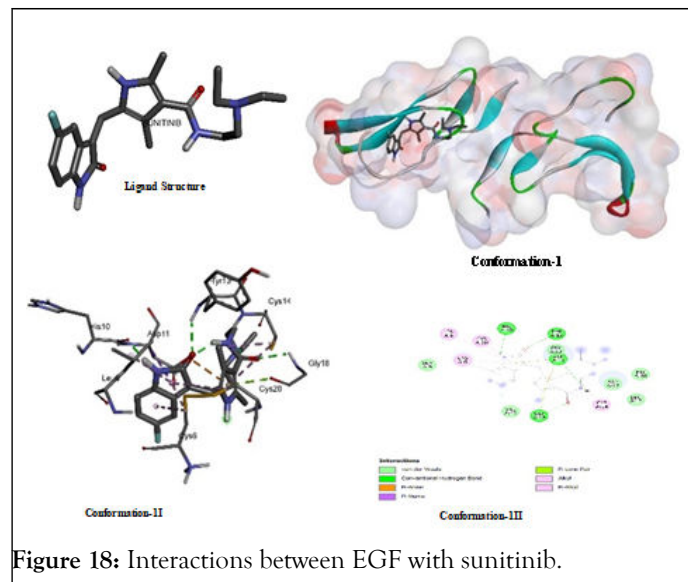


Figure 18: Interactions between EGF with sunitinib.

The overall summarized results of all docking outcomes are given in Table 1. Wherein comparative binding energies for all devised analogues in terms of Gibbs free energy.

Table 1: Comparative binding potential of test compounds 5 and 6 with angiogenic factors.

Ligands	Run											Ki
VEGF												
6	29	-4.4	-4.39	-4.28	-4.24	-4	-3.88	-3.76	-3.75	-3.75	593.30 uM	
5	97	-5.3	-5.08	-5.01	-4.97	-4.93	-4.87	-4.74	-4.73	-4.66	130.58 uM	
Sunitinib	76	-6.71	-6.56	-6.48	-6.45	-6.44	-6.05	-6.03	-6	-6	12.12 uM	
5FU	3	-4.97	-4.24	-4.05	-4	-3.93	-3.86				229.15 uM	
EGF												
6	66	-6.18	-5.42	-4.92	-4.88	-4.58	-4.48	-4.39	-4.37	-4.36	29.74 uM	
5	3	-5.87	-5.77	-5.64	-5.57	-5.43	-5.38	-5.09	-5.08	-4.96	49.83 uM	
5FU	36	-4.97	-4.93	-4.71							227.34 uM	
Sunitinib	43	-8.11	-7.98	-7.7	-7.44	-7.42	-7.31	-7.1	-7.09	-7.06	1.14 uM	
Cox1												
6	96	-9.65	-9.03	-8.36	-8.11	-8.1	-7.98	-7.61	-7.45	-7.32	84.46 nM	
5	80	-9.16	-8.6	-8.54	-8.37	-8.35	-8.2	-8.18	-7.99	-7.82	192.44 nM	

5FU	2	-5.09	-5.04	-5.01	-5	-4.97	-4.95	-4.95	-4.92	-4.92	187.11 uM
Sunitinib	85	-10.3	-9.84	-9.71	-9.28	-9.22	-9.03	-8.93	-8.88	-8.87	28.04 nM
Hif											
6	41	-6.74	-6.54	-6.47	-6.17	-6.05	-5.9	-5.81	-5.64	-5.56	11.40 uM
5	57	-7.64	-6.83	-6.63	-6.49	-6.49	-6.47	-6.45	-6.38	-6.34	2.49 uM
5FU	3	-5.26	-5.17	-5	-4.92	-4.84	-4.79	-4.76	-4.76	-4.68	138.96 uM
Sunitinib	85	-8.21	-8.08	-7.92	-7.84	-7.76	-7.74	-7.72	-7.59	-7.4	964.34 nM

Synthesis and preliminary cytotoxic confirmations

In this study, preliminary synthetic confirmation was made on the basis of solubility, melting points and their physical states. In this context, it was noted that selenium adducts appeared as light-brown sticky mass and their respective benzimidazolium salt 2 and 3 appeared in form of creamy white-flakes. Bis-benzimidazolium salts were soluble in hot distilled water and the adducts, on the other hand, were soluble in alkanols and DMSO. Moreover, the melting points of the adducts were lower than that of their respective salts. In this way, synthesized analogues were preliminary assumed successful in view of the concordance of their properties with reported literature [11].

Spectroscopic confirmations: The confirmation of successful synthesis as per their proposed structures, ^1H and ^{13}C -NMR along with FT-IR spectroscopic studies were accomplished as per the concordance of reported literature. The change in FT-IR spectra of benzimidazolium salts (2 and 4) and their corresponding Se-NHC adducts (5 and 6) in 1100-1600 cm^{-1} region was observed. The emergence of distinct peaks in case of salts and their suppression in case of selenium adducts in region 1750-1550 cm^{-1} were clearly observed. Furthermore, another distinct four-fingers like pattern of peaks were observed in region 1500-1300 cm^{-1} in case of selenium adducts as a sign to confirm their successful synthesis. In ^1H -NMR spectra, appearance of singlet peak in $\delta 7.5$ - $\delta 10$ range confirmed the synthesis 2 and 4. And these disappearance of acidic proton peak confirmed the synthesis of 5 and 6. Moreover, ^{13}C NMR spectra of salts (2 and 4) further verified the successful synthesis of Se-NHC adducts wherein chemical shift value of NCN carbon was changed from region (142-148 δ ppm) in azolium salts as compared to selenium adducts (5 and 6) which appeared in the range (165-168 δ ppm) as per reported literature. The successful synthesis of 1 and 3 was confirmed on ^{13}C -NMR on account of appearance new peaks emerged in $\delta 49$ - $\delta 56$ range.

In-vitro anticancer of synthesized azolium salts and their respective adducts: After molecular docking analysis four cancer cell lines namely: Murine melanoma cancer cell line (B16F10), rat Retinal Ganglion Cancer cell line (1-8RGC5), cervical cancer cell line (HeLa) and breast cancer cell line (MCF7-1) were selected for *in vitro* anticancer study.

In-vitro anticancer activities of the test compounds against B16F10: B16F10 cell-line was selected for its reported use as a model in assessing anti-metastasis potential of developed anti-cancer agents (drugs). Because in assessing the potential of any

drug, B16F10 cells can be considered as a good determinant to assess the activity of anticancer agent especially in context to skin cancers. Cell growth was markedly inhibited after exposure to the test compounds in a dose-dependent manner. It was also showed that numerous morphological changes occurred in cells treated with test compounds. Apoptosis was confirmed by the presence of apoptotic bodies and nuclear condensation. The control, cultured cells without test compounds and cultured with fluorouracil has been shown in each elaboration. However, fluorouracil (positive control) exhibited significant apoptotic activity but some test compounds showed better activity than that. The activity of 2 and 5 demonstrate that tumor cells which have an intrinsic tumorigenic potential in the control samples has offered significant cytotoxic effect in their respective sample snips in the Figure 19.

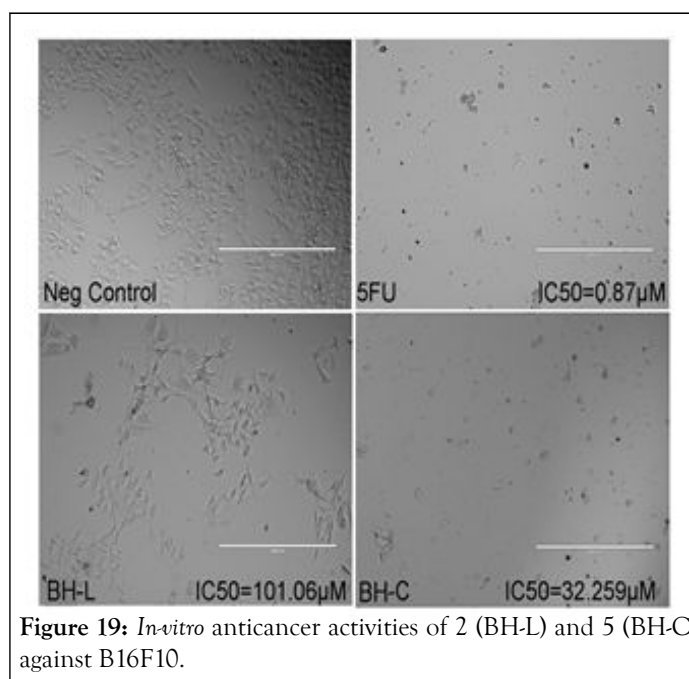
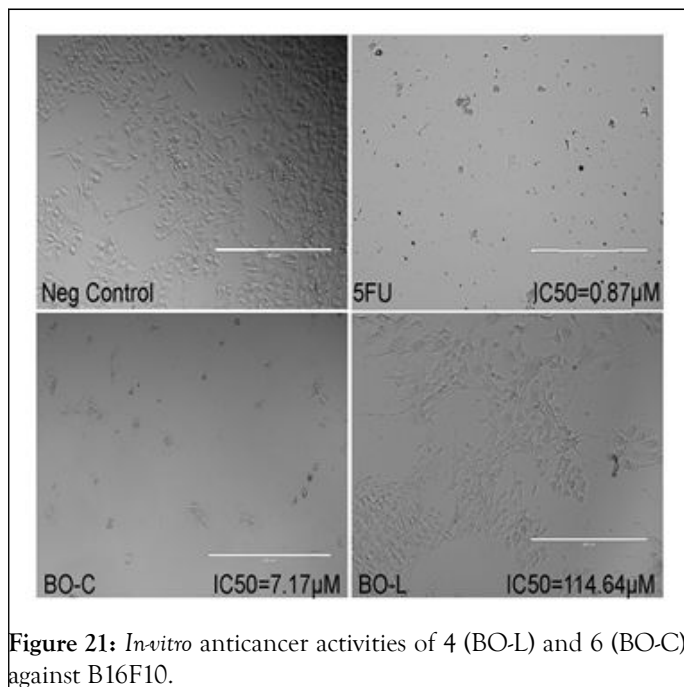
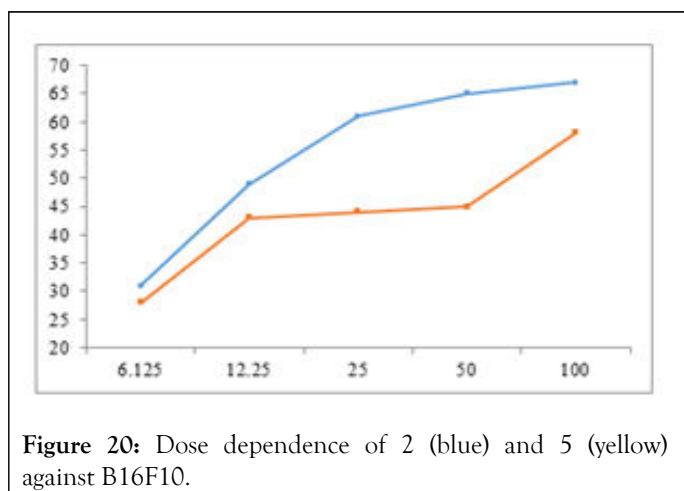


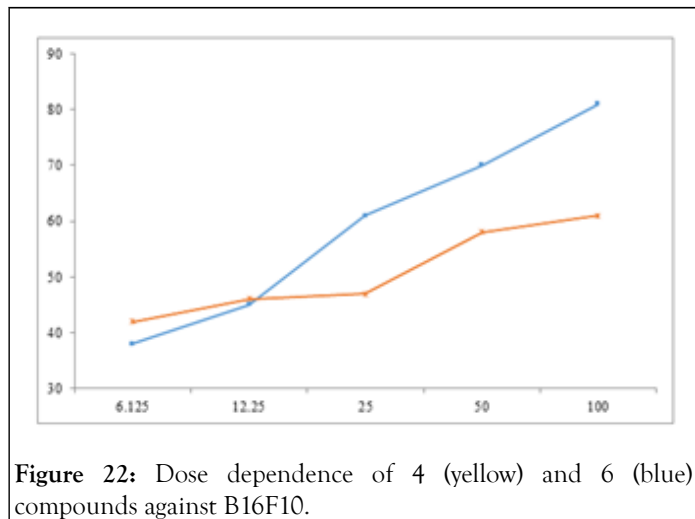
Figure 19: *In-vitro* anticancer activities of 2 (BH-L) and 5 (BH-C) against B16F10.

It can be considered an extension of study of Haque, et al., who synthesized 1,3-bis(3-ethylbenzimidazolium-1-ylmethyl) benzenebis(hexafluorophosphate) A-1 and 1,3-bis(3-butylbenzimidazolium-1-ylmethyl) benzenebis(hexafluorophosphate) A-2 as well as their respective silver complexes 1,3-bis(3-ethylbenzimidazolium-1-ylmethyl) benzenedisilver(I)bis(hexafluorophosphate) A-3 and 1,3-bis(3-butylbenzimidazolium-1-ylmethyl) benzenedisilver(I) bis(hexafluorophosphate) A-4 respectively for anticancer

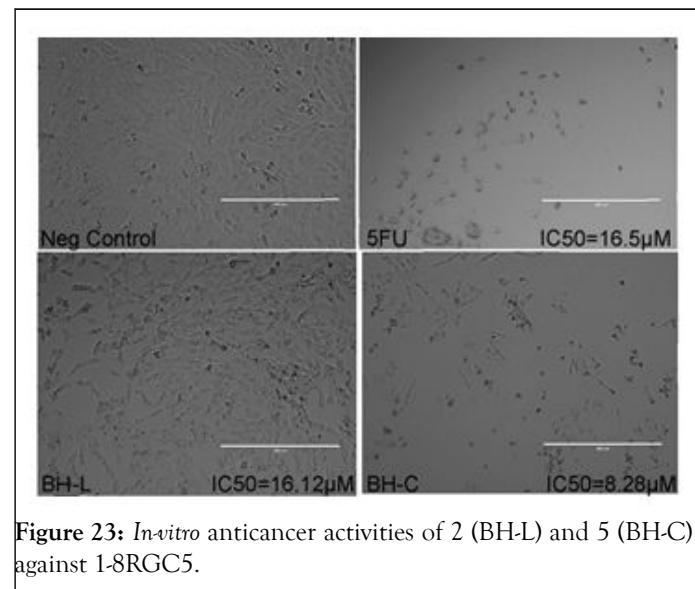
applications taking fluorouracil as positive control but showed IC₅₀ values of 8.7 and 19.4 mM for A-1 and A-2 and 0.2 mM and 1.3 mM for A-3 and A-4 respectively and 5mM for fluorouracil against HCT 116 cancer cell-line. The good to excellent activity was gained much importance because of inherent toxicity of silver on human body. The present study as aimed at designing and synthesizing bisbenzimidazolium salts and their corresponding selenium-adducts which are biocompatible for human beings. Among these synthetic analogues 2 and 5, however, showed higher IC₅₀ values (32 mM) and (101 mM) respectively and no noticeable increase with increase in dosage as shown in Figure 20 but their release likely to have biodegradability and biocompatibility (Figures 20 and 21).



In case of B16F10, compound 6 was found an active cytotoxic agent but not 4, it showed IC₅₀ value of 7.17 mM as compare to 32.25 mM that of 5, however, the trend demonstrated by molecular docking almost reciprocal. But the dosages dependence shoed promising increase in activity with increase in concentration especially in case of 6 as shown in Figure 22.



Anticancer activity against 1-8RGC5: To better assess the anticancer activity of the test compounds, multiple cancer cell lines were selected wherein second-one was rat ganglion cancer cell-line (1-8RGS5) which is often considered as a representative cell-line to assess optic nerve carcinomas. In case of compound 2 and 5 both showed excellent activity in comparison to standard 5-flourouracil wherein bisbenzimidazolium salt 2 showed IC₅₀ value (16.1 mM) almost equal to that of standard and selenium adduct two times higher than that of the same (8.2 mM) compared to (16.5 mM) of standard as shown in Figure 23. This study recurrently justifies the higher cytotoxicity of selenium adducts to cancer-cells than that of their respective azolium salts [12].



Moreover, it was explored in our docking analysis that 5 has consistent affinity to interact with proline and methionine which is not attributed to 2 because of its ionic nature which might be ascribed to higher bioavailability of 5. But in case of 4 and 6 the both were found cytotoxic to 1-8RGC5 cell-line but the previous trend didn't survive because of the higher activity of bisbenzimidazolium salt 4 instead of selenium adduct 6 was explored as shown in Figure 24 which might attributed to stronger ionic interaction of 4 with polar functionalities of oncogenic proteins.

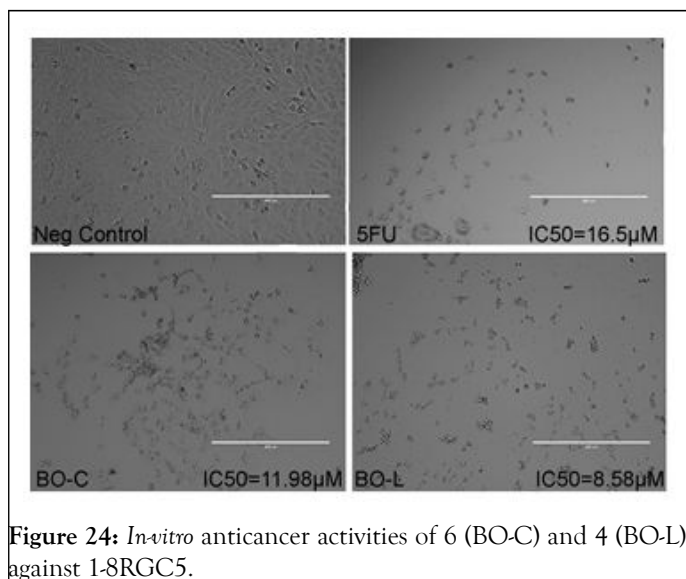


Figure 24: *In-vitro* anticancer activities of 6 (BO-C) and 4 (BO-L) against 1-8RGC5.

***In-vitro* anticancer activities against HeLa:** The photomicrograph of the HeLa cells in Figure 25 shows much clear membrane blebbing and nuclear condensation as compare to the cancer cells treated with 5-fluorouracil. But 5 shows its activity at higher IC_{50} value ($17.48 \mu M$) compared to that of ($4.9 \mu M$) for 5-fluorouracil. It infers that 5 is not so active against HeLa cell-line that might be due to some sort of hindrance in its bioavailability to target oncogenic functions.

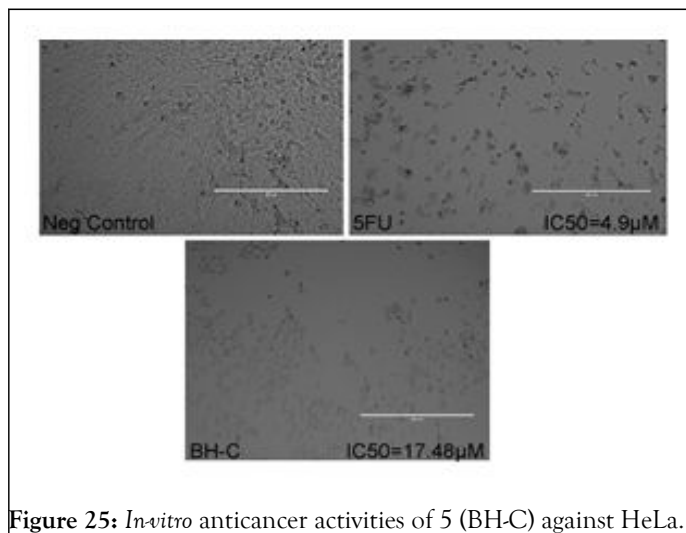


Figure 25: *In-vitro* anticancer activities of 5 (BH-C) against HeLa.

Cytotoxicity of test compounds against MCF7-1: In view of emerging trend of human breast carcinoma (MCF7-1) incidence, the study was also aimed to explore cytotoxicities of test compounds against MCF7-1 cell-line also. In case of 5, promising cytotoxicity with IC_{50} value ($7.22 \mu M$) approximately two times the standard 5-fluorouracil with IC_{50} value ($11 \mu M$) was explored. On the other hand, its respective bisbenzimidazolium salt 2 showed three times lower cytotoxic potential with IC_{50} value ($38.37 \mu M$) as found as compare to the same as shown in Figure 26.

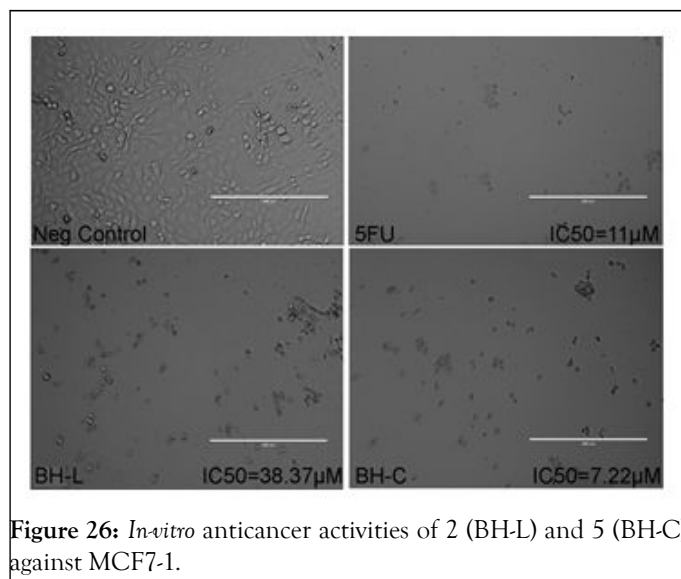


Figure 26: *In-vitro* anticancer activities of 2 (BH-L) and 5 (BH-C) against MCF7-1.

In the whole of study, the activity of compound 4 and 6 against MCF7-1 cell-line was the most important highlight in view of exceptionally higher cytotoxicity than that of the standard (5-fluorouracil). Compound 4 showed more than three times with IC_{50} value ($3.58 \mu M$) and 6 demonstrated IC_{50} value ($6.14 \mu M$) which is two times than that of 5-fluorouracil having IC_{50} value ($11 \mu M$) with, seemingly, apoptotic pattern of cell-death as shown in Figure 27. In this highlight, it was surprisingly found that both benzimidazolium salt 4 being its ionic nature and its corresponding selenium adduct 6 showed higher cytotoxic potential [13].

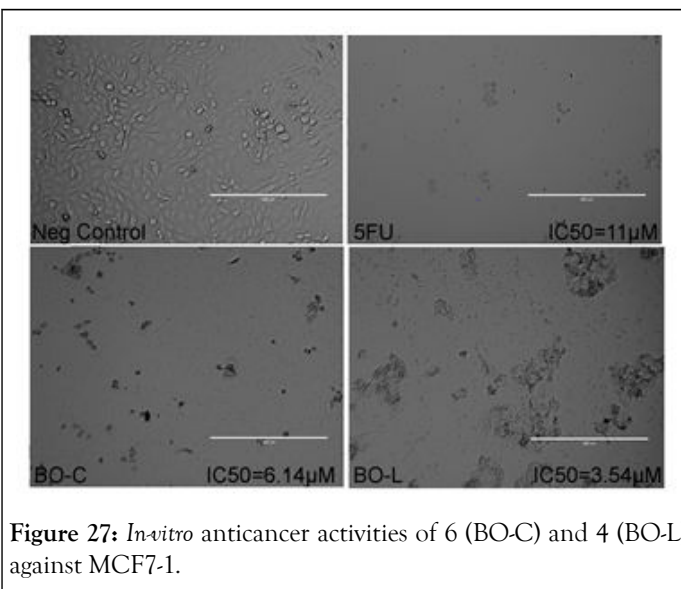


Figure 27: *In-vitro* anticancer activities of 6 (BO-C) and 4 (BO-L) against MCF7-1.

CONCLUSION

After designing of selenium-N-heterocyclic carbene adducts for anticancer activities by molecular docking approach, it was found that 5 and 6 both offer potential binding to oncogenic COX1 and 5 shows binding to HIF. So both of selenium adducts were successfully synthesized by *in-situ* approach and consequently their successful confirmation along with their benzimidazolium salts by FT-IR and NMR spectroscopy. When they subjected to *in-vitro* anticancer activities using four cancer

cell line including: Murine melanoma cancer cell line (B16F10), rat Retinal Ganglion Cancer cell line (1-8RGC5), cervical cancer cell line (HeLa) and breast cancer cell line (MCF7-1) in comparison to standard 5-fluorouracil. It was found that azolium salts 2 showed equivalent to that of the standard; and it adduct 5 showed two times to that of the same. On the other hand, Se-adduct 6 showed almost equivalent the standard but its azolium salt showed two times the activity of standard in case of 1-8RGC5. But in case of MCF7-1 both Se-adduct 6 and its azolium salt 4 showed activity higher than the that of standard so it is to conclude that the studied Se-adducts and their respective azolium salts both are biologically active compounds and they are needed to explored biologically by futher studies.

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CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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