



## Design of an $\alpha$ -L-transfucosidase for the synthesis of fucosylated HMOs

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### Abstract:

Human milk oligosaccharides (HMOs) are recognized as benefiting breast-fed infants in multiple ways. As a result, there is growing interest in the synthesis of HMOs mimicking their natural diversity. Most HMOs are fucosylated oligosaccharides.  $\alpha$ -L-Fucosidases catalyze the hydrolysis of  $\alpha$ -L-fucose from the non-reducing end of a glucan. They fall into the glycoside hydrolase GH29 and GH95 families. The GH29 family fucosidases display a classic retaining mechanism and are good candidates for transfucosidase activity. We recently demonstrated that the  $\alpha$ -L-fucosidase from *Thermotoga maritima* (Tm $\alpha$ Fuc) from the GH29 family can be evolved into an efficient transfucosidase by directed evolution (Osanjco et al. 2007). In this work, we developed semi-rational approaches to design an  $\alpha$ -L-transfucosidase starting with the  $\alpha$ -L-fucosidase from commensal bacteria *Bifidobacterium longum* subsp. *infantis* (BiAfcB, Blon\_2336). Efficient fucosylation was obtained with enzyme mutants (L321P-BiAfcB and F34I/L321P-BiAfcB) enabling in vitro synthesis of lactodifucotetraose, lacto-N-fucopentaose II, lacto-N-fucopentaose III and lacto-N-difucohexaose I. The enzymes also generated more complex HMOs like fucosylated para-lacto-N-neohexaose (F-p-LNnH) and mono- or difucosylated lacto-N-neohexaose (F-LNnH-I, F-LNnH-II and DF-LNnH). It is worth noting that mutation at these two positions did not result in a strong decrease in the overall activity of the enzyme, which makes these variants interesting candidates for large-scale transfucosylation reactions. For the first time, this work provides an efficient enzymatic method to synthesize the majority of fucosylated HMOs.

### Biography:

Dora Molnar-Gabor is a Head of DSP and Analytics at Glycom A/S Supporting and supervising the analytical activities of the R&D Analytical Group Analytical support of Human Milk Oligosaccharides production Development and implementation of new analytical methods and techniques for identification and quantitation of HMOs.



### Recent Publications:

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2. Asakuma S , Hatakeyama E, Urashima T, Yoshida E, Katayama T, Yamamoto K, Kumagai H, Ashida H, Hirose J, Kitaoka M. 2011. Physiology of consumption of human milk oligosaccharides by infant gut-associated bifidobacteria. *J Biol Chem*. 286:34583–34592.
3. Asakuma S , Urashima T, Akahori M, Obayashi H, Nakamura T, Kimura K, Watanabe Y, Arai I, Sanai Y. 2008. Variation of major neutral oligosaccharides levels in human colostrum. *Eur J Clin Nutr*. 62:488–494.
4. Ashida H , Miyake A, Kiyohara M, Wada J, Yoshida E, Kumagai H, Katayama T, Yamamoto K. 2009. Two distinct alpha-L-fucosidases from *Bifidobacterium bifidum* are essential for the utilization of fucosylated milk oligosaccharides and glycoconjugates. *Glycobiology*. 19:1010–1017.
5. Ashline DJ , Yu Y, Lasanajak Y, Song X, Hu L, Ramani S, Prasad V, Estes MK, Cummings RD, Smith DF et al. . 2014. Structural characterization by multistage mass spectrometry (MSn) of human milk glycans recognized by human rotaviruses. *Mol Cell Proteomics*. 13:2961–2974.

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