

Clinical Impact of Radiation-Resistant Mesenchymal Stem Cells in Bone Marrow Deduced from Preclinical Studies

Yuichi Michikawa, Masaharu Hazawa, Ai Saotome-Nakamura, Takeshi Yasuda, Takaya Gotoh and Katsushi Tajima*

Research Program for Radiation Emergency Medicine, Research Center for Radiation Emergency Medicine, National Institute of Radiological Sciences, Anagawa 4-9-1, Inage, Chiba, 263-8555, Japan

Abstract

Mesenchymal stem cells in the bone marrow have attracted great interest over the past decades, not only as a basic scientific subject but also as a novel and advanced clinical tool. More than 100 mesenchymal stem cell-related clinical trials are currently registered in the world. Hematopoietic stem cells in bone marrow are extremely radiation-sensitive, whereas mesenchymal stem cells show considerable radiation-resistance. Intrinsic cellular mechanisms, including highly efficient reactive oxygen species-scavenging ability and active DNA damage response pathways, have been reported to explain the radiation-resistance of mesenchymal stem cells in the bone marrow. The precise interactions between residual host mesenchymal stem cells and donor mesenchymal stem cells at the time of bone marrow transplantation following whole-body irradiation, however, remain unknown. This short review summarizes our current understanding of the clinical impact of the radiation-resistance of endogenous mesenchymal stem cells in the bone marrow.

Keywords: Mesenchymal stem cells; Bone marrow; Radiation

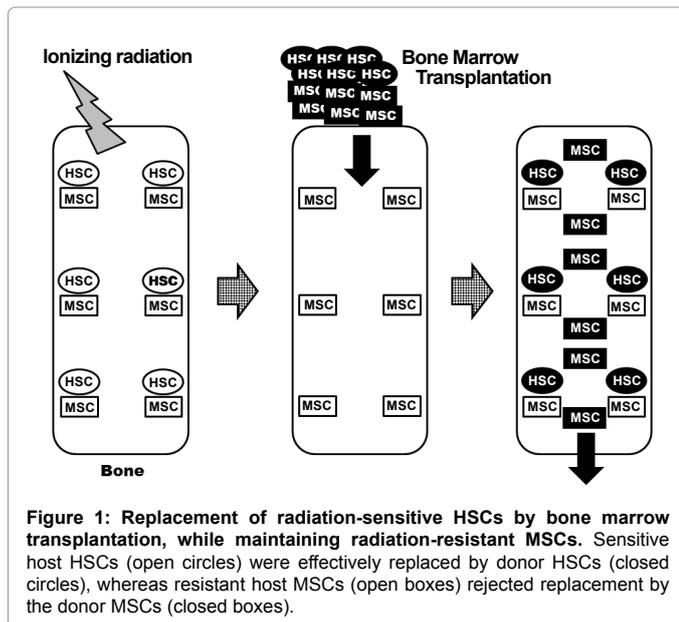
Introduction

Human bone marrow contains at least two distinct types of stem cells [1] in terms of sensitivity to radiation. Hematopoietic Stem Cells (HSCs) are extremely sensitive to radiation [2,3], whereas Mesenchymal Stem Cells (MSCs) are highly resistant to radiation-induced damage [4,5]. Exposure of bone marrow to radiation leads to the rapid depletion of radio-sensitive HSCs and their progenitors, and hematopoietic failure presenting with pancytopenia [3]. Bone marrow transplantation is a useful clinical treatment for this hematopoietic failure. Engrafted donor HSCs may home to the most appropriate sites, created by depletion of the host HSCs and their progenitors, and then reconstruct hematopoiesis. Host MSCs that survive radiation exposure support the regeneration of the donor hematopoietic system [6], although the underlying mechanism is unclear. MSCs are considered key components of the HSC niche, a specialized microenvironment

that regulates the maintenance of HSCs and the production and maturation of hematopoietic progenitors [6]. Because MSCs are also present as minor components in transplanted bone marrow cells, donor MSCs are likely to meet and interact with the host MSCs in the patient's bone marrow, as illustrated in figure 1. This short review focuses on our current understanding of the biologic relevance of radiation resistance of host MSCs in bone marrow based on the clinical outcome, while pointing out unresolved questions to facilitate further research in this field.

MSC Basics

The term "mesenchymal stem cell" has been traditionally used for heterogeneous cell populations with a rather blurred broad definition. MSCs were originally isolated from the bone marrow [7,8], and then considered to be present in virtually all postnatal organs and tissues [9]. The International Society for Cellular Therapy defines that human MSCs must be plastic adherent when maintained *in vitro* and be able to differentiate into osteoblasts, adipocytes, and chondroblasts in standard differentiating cell culture conditions [10]. In addition, the MSC population must express CD73, CD90, and CD105, and lack expression of hematopoietic markers such as CD14, CD34, CD45, and HLA-DR [10]. Thus, the definition of MSCs is not directly applicable to *in vivo* cells. Accordingly, the *in vivo* identity, physiologic function, and biologic properties of MSCs have been investigated mainly by systemic



*Corresponding author: Katsushi Tajima, MD, PhD, Program Leader, Research Program for Radiation Emergency Medicine, Research Center for Radiation Emergency Medicine, National Institute of Radiological Sciences, Anagawa 4-9-1, Inage, Chiba, 263-8555, Japan, Tel: 011-81-43-379-7808; Fax: 011-81-43-206-4094; E-mail: tajima@nirs.go.jp

Received January 07, 2013; Accepted February 01, 2013; Published February 16, 2013

Citation: Michikawa Y, Hazawa M, Saotome-Nakamura A, Yasuda T, Gotoh T, et al. (2013) Clinical Impact of Radiation-Resistant Mesenchymal Stem Cells in Bone Marrow Deduced from Preclinical Studies. J Bone Marrow Res 1: 101. doi:10.4172/2329-8820.1000101

Copyright: © 2013 Michikawa Y, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

transplantation of *in vitro*-cultured cells [11]. Clinical application also requires *ex vivo* cell amplification due to the low content of MSCs in the bone marrow (0.001%-0.01% of total nucleated cells) [12], even though the culture conditions may modify cellular properties, as reported for mouse MSCs [13]. The notion of *in vivo* MSCs, therefore, is rather indirect and hypothetical, leading to gaps in our understanding. Furthermore, unlike human MSCs, mouse bone marrow-derived MSCs have significant limitations in isolation by their adherence to plastic, mostly due to frequent contamination of cultures by hematopoietic cells [14-17]. To overcome this, a methodology was recently developed to selectively isolate a pure population of mouse MSCs directly from bone marrow cells based on surface marker expression [18,19]. An *in vivo* identification method of MSCs still needs to be developed.

Radiation Resistance of MSCs in Bone Marrow

Clinical observations

Differences in radiation sensitivity among bone marrow cells were originally noted in several clinical reports [20-25]. In patients surviving a long time after allogeneic bone marrow transplantation following whole body irradiation, the origin of the two stem cells, HSCs and MSCs, in the bone marrow tends to be significantly different. To identify the origin of the two types of stem cells, genetic markers such as a variable number of tandem repeats in the genomic DNA, which can discriminate origin of stem cells, were used for sex-matched cases [26] and Y-chromosome specific nucleotide sequences were used for sex-mismatched cases [27]. HSCs were found to derive mostly from donors, whereas MSCs derived mostly from the host. These observations were assumed to reflect an inherent difference in the radiation sensitivity of each type of stem cell, namely radio-sensitive host HSCs were effectively replaced by donor HSCs, whereas radio-resistant host MSCs rejected replacement by donor MSCs. Interestingly, pediatric patients who receive allogeneic bone marrow transplantation show mixed chimerism, indicating successful engraftment of donor MSCs [28]. The exact mechanism underlying the easier engraftment of donor MSCs in childhood compared with adulthood, however, remains unclear. MSCs in childhood are still actively proliferating to increase their number [29], and this might enhance their radio-sensitivity, leading to easier depletion of host MSCs and higher efficiency in the engraftment of donor MSCs.

In vitro culture study

The radiation resistance of MSCs isolated from human bone marrow has also been demonstrated in *in vitro* culture studies [4]. In these studies, cells showed considerable *in vitro* radiation resistance compared with so-called radiation resistant cell lines, such as A 549 lung cancer cells [4]. MSCs have several cellular mechanisms, such as highly efficient reactive oxygen species-scavenging ability and avoidance of cell death by active DNA damage response pathways, including cell cycle arrest and DNA repair [4,5,30]. Interestingly, MSCs isolated from different anatomic bone marrow sites display variable responses to ionizing radiation treatment [31]. MSCs derived from maxillary and mandibular trabecular bones are more radiation-resistant than those derived from the iliac crest. MSCs isolated from the maxilla and mandibular trabecular bones induce higher p21 expression, which is known to inhibit apoptosis and harborless DNA damage after ionizing radiation exposure. The induction of p21 expression is considered to activate a robust G1 arrest and DNA repair mechanisms. It remains unclear, however, whether or not MSCs isolated from other organs or tissues have similar radiation resistance.

In vivo animal model study

The radiation resistance of MSCs has been also demonstrated in *in vivo* animal model studies. In a pig model, the mandible was subjected to fractionized radiation of 2 × 9 Gy within 1 week [32]. This treatment corresponds with that of a standardized clinical treatment regimen of head and neck cancer patients fractionally-irradiated with 30 × 2 Gy. Isolation of MSCs at different time-points post-irradiation revealed no significant differences regarding the proliferation capacity and osteogenic differentiation potential. These findings imply that MSCs can effectively cope with higher doses of irradiation *in vivo*. In a murine model, the radiation sensitivity of HSCs and MSCs was compared using a flow cytometry-mediated prospective identification method [18]. HSCs were almost undetectable at 10 days after whole-body irradiation at a dose of 10.5 Gy, whereas a significant number of MSCs remained in bone marrow on the same day. Approximately 71% of freshly isolated MSCs of non-irradiated bone marrow were in the G₀ phase, which could have protected them from lethal irradiation by escaping the cell cycle-mediated apoptotic program [18]. In another murine model, mouse MSCs isolated from flushed bone marrow aspirates were more radio-sensitive than those isolated from collagenase-digested bone marrow [33]. These findings suggest that MSCs in mouse bone marrow are not uniform, but heterogeneous. Therefore, more attention should be paid to the isolation procedures of MSCs from mouse bone marrow.

Clinical Relevance of Radiation Resistance of MSCs Deduced from Preclinical Study

Uncertainty of the functional quality of surviving endogenous MSCs

The detailed properties of radiation-surviving endogenous MSCs are not well documented in human or animal studies. It is quite uncertain whether they can perform usual functions *in vivo* even after exposure to life-threatening ionizing radiation. For example, in an *in vitro* study, MSCs are considered to be sources of tumorigenic cells due to the acquisition of some genetic modifications such as telomere shortening by non-life-threatening low dose radiation exposure, though these findings were not confirmed *in vivo* [34]. The physiologic properties of surviving MSCs after a life-threatening dose of radiation are more likely to differ significantly from those before radiation exposure, despite their having an active DNA damage responding pathway.

Consequences of functionally depleting endogenous HSC niches in bone marrow

MSCs are considered to be part of the HSC niche in the bone marrow [6]. Although selective depletion of MSC functions in bone marrow has not yet been achieved, an interesting observation was made using diphtheria toxin receptor-mediated selective depletion of other defined HSC niche cells, which are called CXC chemokine ligand 12- abundant reticular (CAR) cells [35]. The CAR cells are primary mesenchymal cells with the ability to differentiate into adipocytes as well as osteoblasts, which may be functionally identical to MSCs. HSCs from CAR cell-depleted mice were reduced in number and cell size, were more quiescent, and had increased expression of early myeloid selector genes. Thus, a niche composed of adipo-osteogenic progenitors is required for the proliferation of HSCs and lymphoid and erythroid progenitors, as well as maintenance of HSCs in an undifferentiated state [35]. Accordingly, radiation-damaged host MSCs may similarly influence the donor HSCs.

Lack of evidence regarding direct interactions between surviving host MSCs and donor MSCs

Although many reports indicate *in vivo* or *in vitro* interactions of exogenous MSCs with various endogenous surviving cell types, there is no report concerning direct interaction between host MSCs and donor cells, including MSCs. This is probably due to technical difficulties or due to a lack of interest in this subject. It is worthwhile to point out here a current trend in stem cell biology to intensively investigate interactions between host stem cells and donor stem cells. Emerging concepts propose that transplanted-stem cells act to initiate and stimulate host stem cell-based tissue repair, rather than directly participating in the repair processes [36,37].

Non-hematopoietic antigen-presenting cells are sufficient to induce lethal acute Graft-versus-Host Disease

Graft-versus-Host-Disease (GVHD) is a potentially life-threatening complication and a major limitation of allogeneic hematopoietic stem cell transplantation outcome [38]. This complication is thought to be initiated by the activation of mature donor CD4⁺ T-cells that are co-infused with the hematopoietic stem cell transplant. CD4⁺ T-cells recognize target alloantigens presented on major histocompatibility complex molecules expressed on the antigen-presenting cells that reside somewhere within the host tissues [39]. Upon alloantigen recognition, donor CD4⁺ T-cells become activated, expand, and induce cytotoxic effects on target organs, including the skin, gut, and liver [40]. Although it has been established that recipient dendritic cells are the major antigen-presenting cells expressing allogeneic peptides on their surface [39,41], recent evidence indicates that only non-hematopoietic recipient cells surviving radiation exposure can induce experimental lethal acute GVHD by expressing allogeneic antigens [42]. Because of the properties of MSCs described in previous reports, some bone marrow MSCs surviving after ionizing radiation exposure might represent the GVHD-causing antigen presenting

cells. Importantly, the immunomodulatory functions of MSCs can be converted to suppressing or promoting functions, depending on the conditions surrounding them [43,44]. Thus, the clinical relevance of surviving endogenous MSCs might be stochastic in bone marrow, as illustrated in figure 2.

Future Perspectives Regarding the Radiation Resistance of MSCs

MSCs are now considered as advanced therapy medical products by the European Medicines Agency by regulation No. (EC) 1394/2007 of the European Commission [45,46]. Product approval has been granted for the treatment of pediatric GVHD in Canada and New Zealand (Prochymal[®]; Osiris Therapeutics, Columbia, MD). Clinical-grade large-scale expansion of MSCs is currently available [47-49], supporting more than 100 MSC-related clinical trials registered at <http://www.clinicaltrials.gov/>. To improve the clinical effectiveness of the MSCs, it will be important to understand the interactions between these therapeutic exogenous MSCs and endogenous MSCs. In particular, the immunomodulatory properties of MSCs are suggested to be a double edged-sword [43,44]. Thus, it might be necessary to carefully monitor the functional status of endogenous MSCs and to precisely control them to achieve maximal therapeutic effects of the exogenous MSCs.

Besides hematopoietic regeneration, the radiation resistance of MSCs might be problematic in cases of radiation therapy. MSCs are also regarded as a key component of the tumor stroma, which promotes not only tumor growth, but also angiogenesis and metastasis [50,51]. Thus, the regulation and control of radio-resistant MSCs in the bone marrow might be a critical issue to be addressed in radiation therapy.

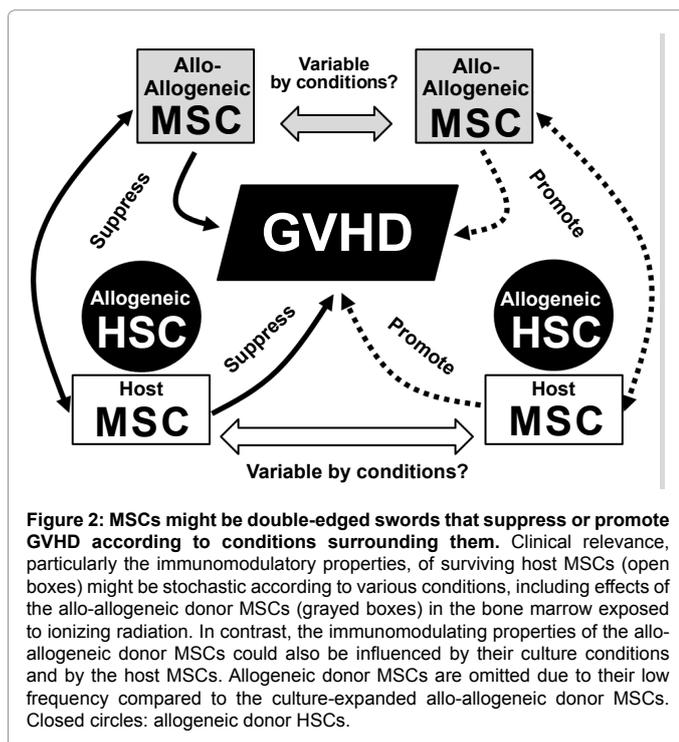
Adult mouse MSCs prospectively purified from bone marrow by flow cytometry were recently demonstrated to be suitable sources for highly efficient generation of high quality induced pluripotent stem (iPS) cells compared to other somatic cells such as fibroblast cells [52]. The iPS cells derived from MSCs appear to be the closest equivalent of the embryonic stem cells based on the gene expression profile and germline-transmission efficiency. Therefore, in the case of severe radiation emergency accidents, especially whole body irradiated cases, radiation-resistant MSCs with high efficiency in the generation of iPS might be the best choice for autonomous regenerative medicine in the future.

Acknowledgement

The authors express their gratitude to Ms. Tomoko Fukuzaki, Ms. Nozomi Gotoh, Ms. Katsuko Noshiro, Ms. Yoko Satoh, Mr. Michio Hama, and Ms. Rie Morihana for their assistance with the manuscript preparation. This work was supported by a grant of NIRS.

Reference

- Koide Y, Morikawa S, Mabuchi Y, Mugeruma Y, Hiratsu E, et al. (2007) Two distinct stem cell lineages in murine bone marrow. *Stem Cells* 25: 1213-1221.
- Mauch P, Constine L, Greenberger J, Knospe W, Sullivan J, et al. (1995) Hematopoietic stem cell compartment: acute and late effects of radiation therapy and chemotherapy. *Int J Radiat Oncol Biol Phys* 31: 1319-1339.
- Fliedner TM, Graessle D, Paulsen C, Reimers K (2002) Structure and function of bone marrow hemopoiesis: mechanisms of response to ionizing radiation exposure. *Cancer Biother Radiopharm* 17: 405-426.
- Chen MF, Lin CT, Chen WC, Yang CT, Chen CC, et al. (2006) The sensitivity of human mesenchymal stem cells to ionizing radiation. *Int J Radiat Oncol Biol Phys* 66: 244-253.
- Sugrue T, Lowndes NF, Ceredig R (2013) Mesenchymal stromal cells: radio-resistant members of the bone marrow. *Immunol Cell Biol* 91: 5-11.
- Méndez-Ferrer S, Michurina TV, Ferraro F, Mazloom AR, Macarthur BD, et



- al. (2010) Mesenchymal and haematopoietic stem cells form a unique bone marrow niche. *Nature* 466: 829-834.
7. Friedenstein AJ, Deriglasova UF, Kulagina NN, Panasuk AF, Rudakowa SF, et al. (1974) Precursors for fibroblasts in different populations of hematopoietic cells as detected by the in vitro colony assay method. *Exp Hematol* 2: 83-92.
8. Prockop DJ (1997) Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 276: 71-74.
9. Nombela-Arrieta C, Ritz J, Silberstein LE (2011) The elusive nature and function of mesenchymal stem cells. *Nat Rev Mol Cell Biol* 12: 126-131.
10. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, et al. (2006) Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 8: 315-317.
11. da Silva Meirelles L, Caplan AI, Nardi NB (2008) In search of the in vivo identity of mesenchymal stem cells. *Stem Cells* 26: 2287-2299.
12. Caplan AI (1994) The mesengenic process. *Clin Plast Surg* 21: 429-435.
13. Rombouts WJ, Ploemacher RE (2003) Primary murine MSC show highly efficient homing to the bone marrow but lose homing ability following culture. *Leukemia* 17: 160-170.
14. Phinney DG, Kopen G, Isaacson RL, Prockop DJ (1999) Plastic adherent stromal cells from the bone marrow of commonly used strains of inbred mice: variations in yield, growth, and differentiation. *J Cell Biochem* 72: 570-585.
15. Peister A, Mellad JA, Larson BL, Hall BM, Gibson LF, et al. (2004) Adult stem cells from bone marrow (MSCs) isolated from different strains of inbred mice vary in surface epitopes, rates of proliferation, and differentiation potential. *Blood* 103: 1662-1668.
16. Soleimani M, Nadri S (2009) A protocol for isolation and culture of mesenchymal stem cells from mouse bone marrow. *Nat Protoc* 4: 102-106.
17. Zhu H, Guo ZK, Jiang XX, Li H, Wang XY, et al. (2010) A protocol for isolation and culture of mesenchymal stem cells from mouse compact bone. *Nat Protoc* 5: 550-560.
18. Morikawa S, Mabuchi Y, Kubota Y, Nagai Y, Niibe K, et al. (2009) Prospective identification, isolation, and systemic transplantation of multipotent mesenchymal stem cells in murine bone marrow. *J Exp Med* 206: 2483-2496.
19. Houlihan DD, Mabuchi Y, Morikawa S, Niibe K, Araki D, et al. (2012) Isolation of mouse mesenchymal stem cells on the basis of expression of Sca-1 and PDGFR- α . *Nat Protoc* 7: 2103-2111.
20. Simmons PJ, Przepiorka D, Thomas ED, Torok-Storb B (1987) Host origin of marrow stromal cells following allogeneic bone marrow transplantation. *Nature* 328: 429-432.
21. Laver J, Jhanwar SC, O'Reilly RJ, Castro-Malaspina H (1987) Host origin of the human hematopoietic microenvironment following allogeneic bone marrow transplantation. *Blood* 70: 1966-1968.
22. Raskind WH, Singer JW, Morgan CA, Fialkow PJ (1988) Host origin of marrow stromal cells obtained from marrow transplant recipients and transformed in vitro by simian virus-40. *Exp Hematol* 16: 827-830.
23. Athanasou NA, Quinn J, Brenner MK, Prentice HG, Graham A, et al. (1990) Origin of marrow stromal cells and haemopoietic chimaerism following bone marrow transplantation determined by in situ hybridisation. *Br J Cancer* 61: 385-389.
24. Stute N, Fehse B, Schröder J, Arps S, Adamietz P, et al. (2002) Human mesenchymal stem cells are not of donor origin in patients with severe aplastic anemia who underwent sex-mismatched allogeneic bone marrow transplant. *J Hematother Stem Cell Res* 11: 977-984.
25. Dickhut A, Schwerdtfeger R, Kuklick L, Ritter M, Thiede C, et al. (2005) Mesenchymal stem cells obtained after bone marrow transplantation or peripheral blood stem cell transplantation originate from host tissue. *Ann Hematol* 84: 722-727.
26. Gatti RA, Nakamura Y, Nussmeier M, Susi E, Shan W, et al. (1989) Informativeness of VNTR genetic markers for detecting chimerism after bone marrow transplantation. *Dis Markers* 7: 105-112.
27. Hutchinson RM, Pringle JH, Potter L, Patel I, Jeffreys AJ (1989) Rapid identification of donor and recipient cells after allogeneic bone marrow transplantation using specific genetic markers. *Br J Haematol* 72: 133-140.
28. Pozzi S, Lisini D, Podestà M, Bernardo ME, Sessarego N, et al. (2006) Donor multipotent mesenchymal stromal cells may engraft in pediatric patients given either cord blood or bone marrow transplantation. *Exp Hematol* 34: 934-942.
29. Tokalov SV, Grüner S, Schindler S, Wolf G, Baumann M, et al. (2007) Age-related changes in the frequency of mesenchymal stem cells in the bone marrow of rats. *Stem Cells Dev* 16: 439-446.
30. Prendergast AM, Cruet-Hennequart S, Shaw G, Barry FP, Carty MP (2011) Activation of DNA damage response pathways in human mesenchymal stem cells exposed to cisplatin or γ -irradiation. *Cell Cycle* 10: 3768-3777.
31. Damek-Poprawa M, Stefanik D, Levin LM, Akintoye SO (2010) Human bone marrow stromal cells display variable anatomic site-dependent response and recovery from irradiation. *Arch Oral Biol* 55: 358-364.
32. Singh S, Kloss FR, Brunauer R, Schimke M, Jamnig A, et al. (2012) Mesenchymal stem cells show radioresistance in vivo. *J Cell Mol Med* 16: 877-887.
33. Carbonneau CL, Despars G, Rojas-Sutterlin S, Fortin A, Le O, et al. (2012) Ionizing radiation-induced expression of INK4a/ARF in murine bone marrow-derived stromal cell populations interferes with bone marrow homeostasis. *Blood* 119: 717-726.
34. Christensen R, Alsner J, Brandt Sorensen F, Dagnaes-Hansen F, Kolvraa S, et al. (2008) Transformation of human mesenchymal stem cells in radiation carcinogenesis: long-term effect of ionizing radiation. *Regen Med* 3: 849-861.
35. Omatsu Y, Sugiyama T, Kohara H, Kondoh G, Fujii N, et al. (2010) The essential functions of adipo-osteogenic progenitors as the hematopoietic stem and progenitor cell niche. *Immunity* 33: 387-399.
36. Dong F, Caplan AI (2012) Cell transplantation as an initiator of endogenous stem cell-based tissue repair. *Curr Opin Organ Transplant* 17: 670-674.
37. Chistiakov DA (2010) Endogenous and exogenous stem cells: a role in lung repair and use in airway tissue engineering and transplantation. *J Biomed Sci* 17: 92.
38. Blazar BR, Korngold R, Valleria DA (1997) Recent advances in graft-versus-host disease (GVHD) prevention. *Immunol Rev* 157: 79-109.
39. Markey KA, Banovic T, Kuns RD, Olver SD, Don AL, et al. (2009) Conventional dendritic cells are the critical donor APC presenting alloantigen after experimental bone marrow transplantation. *Blood* 113: 5644-5649.
40. Beilhack A, Schulz S, Baker J, Beilhack GF, Wieland CB, et al. (2005) In vivo analyses of early events in acute graft-versus-host disease reveal sequential infiltration of T-cell subsets. *Blood* 106: 1113-1122.
41. Duffner UA, Maeda Y, Cooke KR, Reddy P, Ordemann R, et al. (2004) Host dendritic cells alone are sufficient to initiate acute graft-versus-host disease. *J Immunol* 172: 7393-7398.
42. Koyama M, Kuns RD, Olver SD, Raffelt NC, Wilson YA, et al. (2011) Recipient nonhematopoietic antigen-presenting cells are sufficient to induce lethal acute graft-versus-host disease. *Nat Med* 18: 135-142.
43. Vianello F, Dazzi F (2008) Mesenchymal stem cells for graft-versus-host disease: a double edged sword? *Leukemia* 22: 463-465.
44. Li W, Ren G, Huang Y, Su J, Han Y, et al. (2012) Mesenchymal stem cells: a double-edged sword in regulating immune responses. *Cell Death Differ* 19: 1505-1513.
45. European Commission regulation (EC) No 1394/2007 of the European Parliament and of The Council of 13 November 2007 on advanced therapy medicinal products and amending Directive 2001/83/EC and Regulation (EC) No 726/2004. *Official Journal of the European Union* 50: 121-137.
46. Klug B, Celis P, Carr M, Reinhardt J (2012) Regulatory structures for gene therapy medicinal products in the European Union. *Methods Enzymol* 507: 337-354.
47. Fekete N, Rojewski MT, Fürst D, Kreja L, Ignatius A, et al. (2012) GMP-compliant isolation and large-scale expansion of bone marrow-derived MSC. *PLoS One* 7: e43255.
48. Nold P, Brendel C, Neubauer A, Bein G, Hackstein H (2013) Good manufacturing practice-compliant animal-free expansion of human bone marrow derived mesenchymal stroma cells in a closed hollow-fiber-based bioreactor. *Biochem Biophys Res Commun* 430: 325-330.

49. Rojewski MT, Fekete N, Baila S, Nguyen K, Fürst D, et al. (2012) GMP-compliant isolation and expansion of bone marrow-derived MSCs in the closed, automated device Quantum Cell Expansion system. *Cell Transplant*.
50. Huang WH, Chang MC, Tsai KS, Hung MC, Chen HL, et al. (2012) Mesenchymal stem cells promote growth and angiogenesis of tumors in mice. *Oncogene*.
51. Tsukamoto S, Honoki K, Fujii H, Tohma Y, Kido A, et al. (2012) Mesenchymal stem cells promote tumor engraftment and metastatic colonization in rat osteosarcoma model. *Int J Oncol* 40: 163-169.
52. Niibe K, Kawamura Y, Araki D, Morikawa S, Miura K, et al. (2011) Purified mesenchymal stem cells are an efficient source for iPS cell induction. *PLoS One* 6: e17610.