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Chondrogenic Progenitors for Cartilage Repair and Osteoarthritis Treatment

Caroline N Dealy*

Editorial

Center for Regenerative Medicine and Skeletal Development, Department of Reconstructive Sciences, School of Dental Medicine, University of Connecticut Health Center, USA

Department of Orthopaedic Surgery, School of Medicine, University of Connecticut Health Center, USA

The challenge of injury to the articular cartilage of the joints is the inability of the tissue to effectively self-regenerate. Accordingly, most clinical cartilage repair strategies have focused on use of exogenous cells and/or materials to fill in localized acute defects. Problems still to be overcome using these approaches include attaining seamless integration of repair and host tissue, as well as faithful reproduction of true hyaline cartilage, without which permanent and durable repair is not achieved. A critical consideration in cartilage repair is the nature of the cells expected to do the job. Alternatives to the clinical paradigm of exogenous adult chondrocytes as repair cells need to be developed in order to achieve fully functional articular cartilage repair. Moreover, strategies must be devised to treat the widespread and chronic damage found in osteoarthritis, in which surgical intervention to achieve focal repair is not feasible.

Accumulating evidence demonstrates the existence of stem-like cells, possessing chondrogenic potential, residing within or adjacent to the articular cartilage. For example, a localized population of highly proliferative cells expressing progenitor markers is present in the perichondrium at the border of the growth plate and has been suggested to represent a stem cell-like niche for articular cartilage renewal [1]. Several studies have identified the superficial and/or middle layers of the articular cartilage as regions enriched in a mesenchymal stem cell- like population consisting of proliferating cells which express mesenchymal progenitor markers [2-5]. FACS sorting has been used to isolate mesenchymal progenitor cells from normal and/or osteoarthritic articular cartilage [4,6,7] and in in vitro differentiation assays have demonstrated chondrogenic, adipogenic and osteogenic potential by isolated progenitors [3,4,6,7]. Superficial zone progenitors may have an inherent and desirable bias towards differentiation into permanent articular cartilage, as isolated superficial zone progenitors underwent chondrogenic differentiation in vitro without concomitant expression of collagen type X, a marker of growth plate cartilage and hypertrophic maturation diagnostic of osteoarthritic disease [3]. Progenitor cells expressing mesenchymal stem cell markers have also been identified deep in the articular cartilage, as highly migratory cells associated with capillary invasion into the calcified zone past the tidemark [8], a characteristic of severe osteoarthritis. These cells were not observed in normal cartilage [8]. The migratory progenitor cells were found to possess enhanced in vitro chondrogenic potential relative to osteogenic or adipogenic lineages [8]. Mesenchymal progenitor cells have also been isolated from non-cartilage tissue in the joint including the synovium and fatpad [9], with the synovium-derived cells having greater chondrogenic differentiation potential than the adiposederived cells [9].

The existence of multiple endogenous chondrogenic progenitor cell populations in the joint and articular cartilage is exciting in terms of offering potential endogenous cell sources for cartilage repair. However, it is apparent that none of these endogenous progenitor populations are sufficient by themselves in halting osteoarthritic progression. The relative content and distribution of superficial and middle zone progenitors has been found to be similar in normal and osteoarthritic cartilage [4], although the profile of progenitor markers expressed by the cells differed [10]. This suggests that the presence of disease may alter endogenous progenitor populations and compromise their ability to accomplish self-repair. Consistent with this possibility, mesenchymal progenitors isolated from osteoarthritic cartilage underwent spontaneous osteogenic differentiation in vitro which was not observed in normal adult cartilage [11]. It is also possible that endogenous progenitors, and particularly the highly migratory cells associated with vascular invasion in osteoarthritis, may be part of a response to cartilage damage aimed to provide a temporary rather than permanent repair. For instance, surgical procedures such as micro fracture, which allow progenitors from the subchondral bone marrow to enter the articular cartilage, result in formation of fibro cartilage rather than hyaline cartilage [12]. The chondrogenic potential of bone marrow mesenchymal stem cells is further compromised by donor age [13]. Alterations in the local environment of the joint may also reduce the chondrogenic potential of endogenous progenitors. High levels of inflammatory cytokines are present in osteoarthritic or acutely injured joints, and treatment of mesenchymal progenitors with diseased synovial fluid reduces their in vitro chondrogenic differentiation potential [14,15].

Novel approaches for articular cartilage repair may lie in strategies to enhance the effectiveness of resident chondrogenic progenitors by promoting progenitor recruitment, expansion or chondrogenic differentiation. Endogenous progenitors may be recruited to regions of damage via signals which induce cell homing. Remarkably, TGFβ-3 released by a bioscaffold is sufficient, in the absence of exogenous cells, to induce cartilaginous resurfacing of the joint in vivo [16], and also induces recruitment of adipose-, synovium- and mesenchymalderived progenitors into scaffolds in vitro while promoting cartilage characteristic gene expression [17]. Exogenous factors may be introduced into the joint with the goal of promoting chondrogenic differentiation by endogenous progenitor cells in the superficial zones. Intra-articular injection of BMP [18] or hyaluronan (reviewed in [19]) may be particularly promising in this regard as both agents possess well-established pro-chondrogenic activities, and both also promote formation of hyaline cartilage, instead of fibrocartilage, by progenitor cells from bone marrow in joints subjected to microfracture [20-22].

Modification of the extracellular matrix and increased chondrocyte proliferation are characteristics of osteoarthritis typically considered to be part of the disease pathology. Paradoxically, stimulation of

*Corresponding author: Caroline N Dealy, Center for Regenerative Medicine and Skeletal Development (MC-3705), Department of Reconstructive Sciences and Orthopaedic Surgery, University of Connecticut Health Center, 263 Farmington Avenue, Farmington CT 06032, USA, Tel: 860-679-1193; Fax: 860-679-2910; E-mail: deay@nso2.uchc.edu

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these processes may be useful in promoting targeted migration of progenitors to damaged sites [23], or may promote expansion or prime subsequent chondrogenic differentiation by resident progenitor pools [5]. For example, transient treatment of mixed host/donor cartilage explants with the pro-inflammatory cytokine IL1- β caused matrix remodeling via induction of matrix catabolic activity, but ultimately enhanced integration between the two tissues [24]. Moreover, surprisingly, blocking matrix metallo protease activity has been found to suppress chondrogenic differentiation of mesenchymal stem cells *in vitro* [25]. Further, transient stimulation of β -catenin signaling, a signal typically associated with osteoarthritic progression [26], instead caused thickening of the articular cartilage *in vivo* [27] and increased proliferation of isolated superficial zone progenitors *in vitro* while promoting subsequent differentiation of the cells towards permanent articular cartilage *in vivo* [5].

Strategies to augment the response of endogenous progenitor populations to cartilage injury may involve not only exogenous factors, but also exogenous progenitor cells. In particular the potential use of mesenchymal stem cells for treatment of articular cartilage damage has been intensely investigated (reviewed in [28-30]). Outcomes from studies in which progenitor cells including mesenchymal stem cells were implanted or injected into damaged joints in animal models [31-36] or humans [37,38] have been encouraging, however, the mechanisms by which the exogenous progenitor cells restore cartilage integrity and/or function are poorly understood. Intriguingly, a recent study examining articular cartilage repair by human embryonic stem cell-derived chondrocytes, which were implanted into full thickness focal defects in rat articular cartilage in vivo and monitored using a human-specific antibody, revealed gradual replacement of the human cells with rat cells during repair of the defect region [39]. This suggests the exogenous chondrocytes provide paracrine signals which can enable the endogenous host tissue accomplish cartilage repair. Signals from the host tissue may in turn influence differentiation by exogenous progenitors introduced into the damaged cartilage or joint. The injured or osteoarthritic joint is classically considered hostile due to the presence of inflammatory cytokines in the joint fluid. However, the local microenvironment adjacent to the damaged cartilage may provide access to chondrocyte-produced pro-chondrogenic factors that may positively influence chondrogenic potential of engrafted exogenous cells. Indeed, while intra-articular injection of human embryonic stem cells leads to teratomas which ultimately destroy the joint, when introduced as implants directly into a focal defect the cells instead form cartilage [40]. Mutually beneficial interactions between progenitors and chondrocytes have also been shown. For instance, chondrogenesis in co-cultures of mesenchymal stem cells and articular chondrocytes is mediated by progenitor-stimulated chondrocyte proliferation, in conjunction with chondrocytestimulated progenitor cell differentiation into the chondrocyte lineage [41]. In addition, participation of mesenchymal progenitors in repair of articular cartilage was accompanied by strong induction of collagen type II staining around both host and transplanted cells [36]. Thus, exogenous progenitor cells may function in promoting cartilage repair through interaction with endogenous chondrocytes or chondrogenic progenitors, in addition to themselves serving as a supplemental source of cells for reconstruction of damaged cartilage.

In order to exploit the potential of exogenous signals and cells in cartilage repair strategies, means for efficient delivery and retention of the factors or cells at the region of damage will need to be devised. This may be achieved for localized defects via direct surgical implantation of bioscaffolds, with or without incorporated factors or cells. Alternate approaches such as direct injection into the joint will be required for treatment of non-focal osteoarthritic lesions [42]. A non-invasive approach would be preferable for the patient and convenient for the health care provider even for local damage due to acute injury. Clinical trials are underway to evaluate the safety and efficacy for osteoarthritis treatment of direct intra-articular injection of recombinant BMP [43], or of genetically-modified chondrocytes expressing TGF-β1 [44]. Encapsulation of growth factors or genetically-transduced cells releasing such factors, in biomaterial microspheres is also being investigated as a way to achieve sustained delivery into the joint [45-47]. Several studies in which mesenchymal progenitor cells were labeled and tracked following direct intra-articular injection in damaged or osteoarthritic joints have demonstrated presence of labeled cells within the tissues of the joint including the surface and interior regions of the articular cartilage [32,33,48,49]. In one study injected mesenchymal stem cells were found to repopulate fibro cartilage and synovium but not articular cartilage [31]. However, the cells were found to attach and populate deep fissures of human osteoarthritic cartilage explants, and subsequently formed a regenerated cartilage surface in vitro [50]. Thus physical as well as molecular cues may promote graft-host tissue integration. Incorporation of exogenous progenitors into the articular cartilage may be enhanced by increasing the number of injected cells [51], or by co-injecting them with hyaluronan [36]. One novel approach for targeting injected cells to damaged articular cartilage utilized iron-labeled synovial-derived progenitors which were injected into the joint and homed to the damaged region via implantation of an intra-articular magnet [52].

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This issue of Rheumatology: Current Research is focused on chondrogenic progenitor responses to cartilage injury. These encouraging studies suggest the field can expect exciting progress in utilization of exogenous signals or progenitor cells for cartilage repair, as well as development of strategies to enhance repair by endogenous chondrogenic progenitors already present in articular cartilage. Importantly, these approaches may offer new promise for treatment of cartilage injury and osteoarthritis.

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References

- Karlsson C, Thornemo M, Henriksson HB, Lindahl A (2009) Identification of a stem cell niche in the zone of Ranvier within the knee joint. J Anat 215: 355-363.
- Dowthwaite GP, Bishop JC, Redman SN, Khan IM, Rooney P, et al. (2004) The surface of articular cartilage contains a progenitor cell population. J Cell Sci 117: 889-897.
- McCarthy HE, Bara JJ, Brakspear K, Singhrao SK, Archer CW (2011) The comparison of equine articular cartilage progenitor cells and bone marrowderived stromal cells as potential cell sources for cartilage repair in the horse. Vet J.
- Pretzel D, Linss S, Rochler S, Endres M, Kaps C, et al. (2011) Relative percentage and zonal distribution of mesenchymal progenitor cells in human osteoarthritic and normal cartilage. Arthritis Res Ther 13: R64.
- Yasuhara R, Ohta Y, Yuasa T, Kondo N, Hoang T, et al. (2011) Roles of β-catenin signaling in phenotypic expression and proliferation of articular cartilage superficial zone cells. Lab Invest 91: 1739-1752.
- Fickert S, Fiedler J, Brenner RE (2004) Identification of subpopulations with characteristics of mesenchymal progenitor cells from human osteoarthritic cartilage using triple staining for cell surface markers. Arthritis Res Ther 6: R422-432.
- Hattori S, Oxford C, Reddi AH (2007) Identification of superficial zone articular chondrocyte stem/progenitor cells. Biochem Biophys Res Commun 358: 99-103.

- Koelling S, Kruegel J, Irmer M, Path JR, Sadowski B, et al. (2009) Migratory chondrogenic progenitor cells from repair tissue during the later stages of human osteoarthritis. Cell Stem Cell 4: 324-335.
- Lee SY, Nakagawa T, Reddi AH (2010) Mesenchymal progenitor cells derived from synovium and infrapattellar fat pad as a source of superficial zone cartilage tissue engineering: analysis of superficial zone protein/lubricin expression. Tissue Eng Part A 16: 317-325.
- Grogan SP, Miyaki S, Asahara H, D'Lima DD, Lotz MK (2009) Mesenchymal progenitor cell markers in human articular cartilage: normal distribution and changes in osteoarthritis. Arthritis Res Ther 11: R85.
- Chang HX, Yang L, Li Z, Chen G, Dai G (2011) Age-related biological characterization of mesenchymal progenitor cells in human articular cartilage. Orthopedics 34: e382-388.
- LaPrade RF, Bursch LS, Olson EJ, Havlas V, Carlson CS (2008) Histologic and immunohistochemical characteristics of failed articular cartilage resurfacing procedures for osteochondritis of the knee: a case series. Am J Sports Med 36: 360-368.
- Murphy JM, Dixon K, Beck S, Fabian D, Feldman A, et al. (2002) Reduced chondrogenic and adipogenic activity of mesenchymal stem cells from patients with advanced osteoarthritis. Arthritis Rheum 46: 704-713.
- 14. Ando W, Heard BJ, Chung M, Nakamura N, Frank CB, et al. (2012) Ovine synovial membrane-derived mesenchymal progenitor cells retain the phenotype of the original tissue that was exposed to in-vivo inflammation: evidence for a suppressed chondrogenic differentiation potential of the cells. Inflamm Res.
- 15. Kruger JP, Endres M, Neumann K, Stuhlmuller B, Morawietz L, et al. (2012) Chondrogenic differentiation of human subchondral progenitor cells is affected by synovial fluid from donors with osteoarthritis or rheumatoid arthritis. J Orthop Surg Res 7: 10.
- Lee CH, Cook JL, Mendelson A, Moioli EK, Yao H, et al. (2010) Regeneration of the articular surface of the rabbit synovial joint by cell homing: a proof of concept study. Lancet 376: 440-448.
- Mendelson A, Frank E, Allred C, Jones E, Chen M, et al. (2011) Chondrogenesis by chemotactic homing of synovium, bone marrow, and adipose stem cells in vitro. FASEB J 25: 3496-3504.
- Hayashi M, Muneta T, Takahashi T, Ju YJ, Tsuji K, et al. (2010) Intra-articular injections of bone morphogenetic protein-7 retard progression of existing cartilage degeneration. J Orthop Res 28: 1502-1506.
- Bannuru RR, Natov NS, Dasi UR, Schmid CH, McAlindon TE (2011) Therapeutic trajectory following intra-articular hyaluronic acid injection in knee osteoarthritis--meta-analysis. Osteoarthritis Cartilage 19: 611-619.
- Kuo AC, Rodrigo JJ, Reddi AH, Curtiss S, Grotkopp E, et al. (2006) Microfracture and bone morphogenetic protein 7 (BMP-7) synergistically stimulate articular cartilage repair. Osteoarthritis Cartilage 14: 1126-1135.
- 21. Yang HS, La WG, Bhang SH, Kim HJ, Im GI, et al. (2011) Hyaline cartilage regeneration by combined therapy of microfracture and long-term bone morphogenetic protein-2 delivery. Tissue Eng Part A 17: 1809-1818.
- Kang SW, Bada LP, Kang CS, Lee JS, Kim CH, et al. (2008) Articular cartilage regeneration with microfracture and hyaluronic acid. Biotechnol Lett 30: 435-439.
- Gerter R, Kruegel J, Miosge N (2012) New insights into cartilage repair the role of migratory progenitor cells in osteoarthritis. Matrix Biol 31: 206-213.
- 24. Khan IM, Gonzalez LG, Francis L, Conlan RS, Gilbert SJ, et al. (2011) Interleukin-1 β enhances cartilage-to-cartilage integration. Eur Cell Mater 22: 190-201.
- Bertram H, Boeuf S, Wachters J, Boehmer S, Heisel C, et al. (2009) Matrix metalloprotease inhibitors suppress initiation and progression of chondrogenic differentiation of mesenchymal stromal cells in vitro. Stem Cells Dev 18: 881-892.
- 26. Zhu M, Tang D, Wu Q, Hao S, Chen M, et al. (2009) Activation of beta-catenin signaling in articular chondrocytes leads to osteoarthritis-like phenotype in adult beta-catenin conditional activation mice. J Bone Miner Res 24: 12-21.
- 27. Yuasa T, Kondo N, Yasuhara R, Shimono K, Mackem S, et al. (2009) Transient activation of Wnt/{beta}-catenin signaling induces abnormal growth plate

closure and articular cartilage thickening in postnatal mice. Am J Pathol 175: 1993-2003.

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- Khan WS, Johnson DS, Hardingham TE (2010) The potential of stem cells in the treatment of knee cartilage defects. Knee 17: 369-374.
- O'Sullivan J, D'Arcy S, Barry FP, Murphy JM, Coleman CM (2011) Mesenchymal chondroprogenitor cell origin and therapeutic potential. Stem Cell Res Ther 2: 8.
- Qi Y, Feng G, Yan W (2012) Mesenchymal stem cell-based treatment for cartilage defects in osteoarthritis. Mol Biol Rep 39: 5683-5689.
- Murphy JM, Fink DJ, Hunziker EB, Barry FP (2003) Stem cell therapy in a caprine model of osteoarthritis. Arthritis Rheum 48: 3464-3474.
- Lee KB, Hui JH, Song IC, Ardany L, Lee EH (2007) Injectable mesenchymal stem cell therapy for large cartilage defects--a porcine model. Stem Cells 25: 2964-2971.
- 33. Matsumoto T, Cooper GM, Gharaibeh B, Meszaros LB, Li G, et al. (2009) Cartilage repair in a rat model of osteoarthritis through intraarticular transplantation of muscle-derived stem cells expressing bone morphogenetic protein 4 and soluble Flt-1. Arthritis Rheum 60: 1390-1405.
- Chang CH, Kuo TF, Lin FH, Wang JH, Hsu YM, et al. (2011) Tissue engineeringbased cartilage repair with mesenchymal stem cells in a porcine model. J Orthop Res 29: 1874-1880.
- Toghraie FS, Chenari N, Gholipour MA, Faghih Z, Torabinejad S, et al. (2011) Treatment of osteoarthritis with infrapatellar fat pad derived mesenchymal stem cells in Rabbit. Knee 18: 71-75.
- 36. Sato M, Uchida K, Nakajima H, Miyazaki T, Rodriguez Guerrero A, et al. (2012) Direct transplantation of mesenchymal stem cells into the knee joints of Hartley Strain guinea pig with spontaneous osteoarthritis. Arthritis Res Ther 14: R31.
- 37. Wakitani S, Okabe T, Horibe S, Mitsuoka T, Saito M, et al. (2011) Safety of autologous bone marrow-derived mesenchymal stem cell transplantation for cartilage repair in 41 patients with 45 joints followed for up to 11 years and 5 months. J Tissue Eng Regen Med 5: 146-150.
- 38. Pak J (2011) Regeneration of human bones in hip osteonecrosis and human cartilage in knee osteoarthritis with autologous adipose-tissue-derived stem cells: a case series. J Med Case Reports 5: 296.
- Toh WS, Lee EH, Guo XM, Chan JK, Yeow CH, et al. (2010) Cartilage repair using hyaluronan hydrogel-encapsulated human embryonic stem cell-derived chondrogenic cells. Biomaterials 31: 6968-6980.
- Wakitani S, Aoki H, Harada Y, Sonobe M, Morita Y, et al. (2004) Embryonic stem cells form articular cartilage, not teratomas, in osteochondral defects of rat joints. Cell Transplant 13: 331-336.
- Acharya C, Adesida A, Zajac P, Mumme M, Riesle J, et al. (2012) Enhanced chondrocyte proliferation and mesenchymal stromal cells chondrogenesis in coculture pellets mediate improved cartilage formation. J Cell Physiol 227: 88-97.
- 42. Kwon DR, Park GY (2012) Intra-articular injections for the treatment of osteoarthritis: focus on the clinical use of several regimens. In: Osteoarthritis-Diagnosis, Treatment and Surgery. Chen Q (Ed), In Tech 67-100.
- 43. Ha CW, Noh MJ, Choi KB, Lee KH (2012) Initial phase I safety of retrovirally transduced human chondrocytes expressing transforming growth factor-beta-1 in degenerative arthritis patients. Cytotherapy 14: 247-256.
- 44. Hunter DJ, Pike MC, Jonas BL, Kissin E, Krop J, et al. (2010) Phase 1 safety and tolerability study of BMP-7 in symptomatic knee osteoarthritis. BMC Musculoskelet Disord 11: 232.
- Siu RK, Zara JN, Hou Y, James AW, Kwak J, et al. (2012) NELL-1 promotes cartilage regeneration in an in vivo rabbit model. Tissue Eng Part A 18: 252-261.
- 46. Zhang Z, Bi X, Li H, Huang G (2011) Enhanced targeting efficiency of PLGA microspheres loaded with Lornoxicam for intra-articular administration. Drug Deliv 18: 536-544.
- 47. Eswaramoorthy R, Chang CC, Wu SC, Wang GJ, Chang JK, et al. (2012) Sustained release of PTH(1-34) from PLGA microspheres suppresses osteoarthritis progression in rats. Acta Biomater .
- 48. Mokbel A, El-Tookhy O, Shamaa AA, Sabry D, Rashed L, et al. (2011) Homing and efficacy of intra-articular injection of autologous mesenchymal stem cells in experimental chondral defects in dogs. Clin Exp Rheumatol 29: 275-284.

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- 49. Mokbel AN, El Tookhy OS, Shamaa AA, Rashed LA, Sabry D, et al. (2011) Homing and reparative effect of intra-articular injection of autologus mesenchymal stem cells in osteoarthritic animal model. BMC Musculoskelet Disord 12: 259.
- Coleman CM, Curtin C, Barry FP, O'Flatharta C, Murphy JM (2010) Mesenchymal stem cells and osteoarthritis: remedy or accomplice? Hum Gene Ther 21: 1239-1250.
- 51. Agung M, Ochi M, Yanada S, Adachi N, Izuta Y, et al. (2006) Mobilization of bone marrow-derived mesenchymal stem cells into the injured tissues after intraarticular injection and their contribution to tissue regeneration. Knee Surg Sports Traumatol Arthrosc 14: 1307-1314.
- Hori J, Deie M, Kobayashi T, Yasunaga Y, Kawamata S, et al. (2011) Articular cartilage repair using an intra-articular magnet and synovium-derived cells. J Orthop Res 29: 531-538.