

Chemical Composition of *Hypericum laricifolium* Juss. Essential Oil Collected from Mérida-Venezuela

Janne Rojas*, Alexis Buitrago, Luis B Rojas and Antonio Morales

Organic Biomolecular Research Institute, Faculty of Pharmacy and Bioanalysis, University of Los Andes, Venezuela

Abstract

Hypericum is one of nine genera forming the family *Hypericaceae* distributed mainly in temperate regions and tropical mountains. Fresh aerial parts of *H. laricifolium* Juss. collected by the Piñango road, Mérida state, at 3900 m above sea level, yielded 1.2 mL (0.24% w/v) of essential oil by hydrodistillation. GC and GC/MS analysis showed the presence of 21 components being α -pinene (20.2%), verticilol (13.4%), 3-methyl-nonane (12.3%), 2-methyl-octane (9.6%) and nonane (7.6%) in major proportions. To the best of our knowledge, this is the first report on the chemical composition of the *Hypericum laricifolium* essential oil.

Keywords: *Hypericum laricifolium*; Essential oil; Hypericaceae; GC-MS; α -pinene; Verticilol

Introduction

Hypericum is one of nine genera forming the family *Hypericaceae*, comprises about 500 species distributed mainly in temperate regions and tropical mountains being most abundant in Europe, Asia, Northern Africa and America [1,2]. Many of these species have been used in traditional medicine to alleviate a number of diseases such as antispasmodic, diuretic, antimigraine, antiepileptic and cholagogue [3], and also for diarrhea, dyspepsia, parasite, neuralgia, sciatica and rheumatism [4,5].

Phytochemical studies carried out with different *Hypericum* species have revealed the presence of bioactive substances like phloroglucinol derivatives, naphthodianthrones, flavonoids, phenylpropanes, essential oils, benzophenones and xanthenes, showing a wide range of biological properties related to antimicrobial, anticancer, antidepressant, antianxiety, antiviral, cytotoxic and antifungal activities [6-9].

In the present investigation, the chemical composition of *Hypericum laricifolium* Juss essential oil collected from Mérida-Venezuela has been studied and compared to several *Hypericum* species reported worldwide.

Materials and Methods

Plant material

Fresh leaves of *Hypericum laricifolium* Juss were collected in May 2012 by the Piñango road, approximately 20 km from "Collado del Condor" peak, at 3900 m above sea level. A voucher specimen (JR33) was deposited in the Luis Terán Herbarium of the Faculty of Pharmacy and Bio analysis, University of Los Andes, Mérida, Venezuela.

Isolation of essential oil

The botanical material (500 g) was cut into small pieces and subjected to hydrodistillation for 4 h, using a Clevenger-type apparatus. The oil was dried over anhydrous sodium sulphate and stored at 4°C, until assays completed

Gas chromatography (GC)

GC analyses were performed on a Perkin-Elmer Auto System gas chromatograph equipped with flame ionization detectors. A 5% phenylmethyl polysiloxane fused-silica capillary column (AT-5, Alltech Associates Inc., Deerfield, IL), 60 m \times 0.25 mm, film thickness 0.25 μ m

was used for the GC analysis. The initial oven temperature was 60°C; this was then raised to 260°C at 4°C/min, and the final temperature maintained for 20 min. The injector and detector temperatures were 200°C and 250°C, respectively. The carrier gas was helium at 1.0 mL/min. The sample was injected using a split ratio of 1:100. Retention indices were calculated relative to C8-C24 n-alkanes, and compared with values reported in the literature [10,11].

Gas chromatography-mass spectrometry (GC-MS)

The GC-MS analyses were carried out on a Hewlett Packard GC-MS system, Model 5973, fitted with a 30 m long, cross-linked 5% phenylmethyl siloxane (HP-5MS, Hewlett Packard, USA) fused-silica column (0.25 mm, film thickness 0.25 μ m). Source temperature 230°C; quadrupole temperature 150°C; carrier gas helium, adjusted to a linear velocity of 34 m/s; ionization energy, 70 eV; scan range 40-500 amu; 3.9 scans/s. The injected volume was 1.0 μ L of a 2% dilution of oil in n-heptane. A Hewlett-Packard ALS injector was used with split ratio 1:100. The identification of the oil components was based on a Wiley MS Data Library (6th Edn.), followed by comparisons of MS data with published literature [11].

Results and Discussion

Fresh aerial parts of *H. laricifolium* yielded 1.2 mL (0.24% w/v) of essential oil, which showed by GC/MS (Figure 1), the presence of 21 components being α -pinene (20.2%), verticilol (13.4%), 3-methyl-nonane (12.3%), 2-methyl-octane (9.6%) and nonane (7.6%) in major proportions. Table 1 summarizes a complete list of identified components. According to previous reports, α -pinene is a common monoterpene identified in several *Hypericum* species, where it is found among the main components along with its isomer, β -pinene.

A study from Turkey reported for *H. triquetrifolium* essential

*Corresponding author: Janne Rojas, Organic Biomolecular Research Institute, Faculty of Pharmacy and Bioanalysis, University of Los Andes, Venezuela, Tel: +58-274-2403557; Fax: +58-274-2403455; E-mail: janner@ula.ve

Received July 02, 2013; Accepted August 05, 2013; Published August 08, 2013

Citation: Rojas J, Buitrago A, Rojas LB, Morales A (2013) Chemical Composition of *Hypericum laricifolium* Juss. Essential Oil Collected from Mérida-Venezuela. Med Aromat Plants 2: 132. doi: 10.4172/2167-0412.1000132

Copyright: © 2013 Rojas J, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

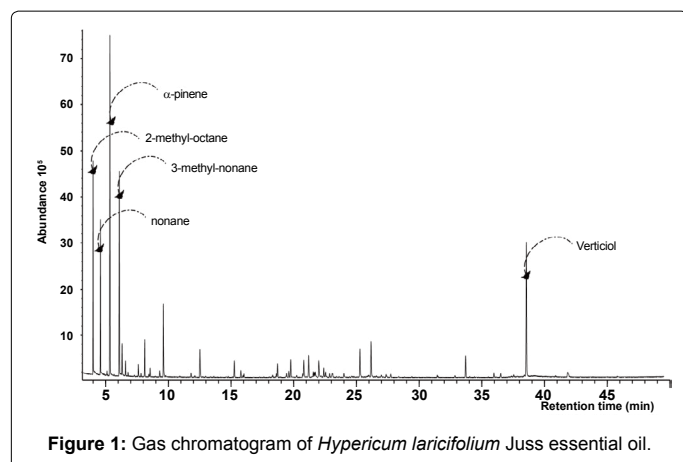


Figure 1: Gas chromatogram of *Hypericum laricifolium* Juss essential oil.

Compounds	%	RI
2-methyl-octane	9.6	866
nonane	7.6	906
α -pinene	20.2	945
3-methyl-nonane	12.3	977
β -pinene	1.9	986
sabinene	1.0	996
limonene	1.0	1039
(E)- β -ocymene	2.4	1058
2-methyl-decane	0.6	1073
terpinolene	0.5	1098
undecane	5.4	1311
α -terpineol	2.4	1424
2-undecanone	0.6	1430
(E)- α -bergamotene	0.5	1467
aromadendrene	1.5	1481
α -humulene	1.7	1496
β -acoradiene	2.0	1499
curcumene	0.4	1614
(E)- β -farnesene	0.5	1652
cembrene	2.0	1938
verticilol	13.4	2166

The composition of the essential oil was determined by comparison of the MS of each component with Wiley GC/MS library data and also from its retention index (RI).

Table 1: Essential oil composition of *Hypericum laricifolium* Juss.

oil α -pinene (12.3%) as one of the main components, while *H. aviculariifolium* subsp. *depilatum* var. *depilatum* showed high concentrations of α -pinene (52.1%), but β -pinene (3.6%) was present in minor proportions [12]. *H. annulatum* from Serbia also reported high concentrations of α -pinene (56.3%) and lower amount of β -pinene (4.2%) [13]. Similarly from Serbia, another investigation revealed that *H. richeri* subsp. *grisebachii* contained β -pinene (5.8%) among its components [14]. *H. elongatum* also showed a large amount of α -pinene (80.43%) and determined β -pinene (2.59%) between the minor compounds [14]. However, *H. rumeliacum* showed higher concentrations of β -pinene (26.1%) comparing to α -pinene (7.3%) [15].

On the other hand, *H. perforatum*, known as the most commercially important species within the genus *Hypericum*, showed the presence of α -pinene (8.6%) and β -pinene (3.8%), along with germacrene-D (22.1%), β -caryophyllene (11.3%), α -cadinol (4.4%), 2-methyl-octane (3.7%), terpinen-4-ol (3.3%), caryophyllene oxide (3.3%), α -muurolol

(2.9%) and spathulenol (2.8%) [16]. Other investigation from Tajikistan showed that essential oil of *H. perforatum* was mainly composed by germacrene-D (13.7%), α -pinene (5.1%), trans-caryophyllene (4.7%), *n*-dodecanol (4.5%), caryophyllene oxide (4.2%), bicycle-germacrene (3.8%) and spathulenol (3.4%), while for *H. scabrum*, the main components were α -pinene (44.8%), spathulenol (7.1%), verbenone (6.0%), trans-verbenol (3.9%) and γ -muurolene (3.5%) [17].

A recent investigation carried out with several *Hypericum* species collected from different locations in Europe, Asia and North Africa reported for *H. uniglandulosum* 2,6-Dimethyl-3,5-heptadien-2-one (40.7%), nonacosane (3.2%), hexadecanoic acid (2.7%) and α -pinene (2.7%); for *H. scabroides*, hexadecanoic acid (17.7%), spathulenol (5.3%), nonacosane (4.4%), dodecanoic acid (4.1%), baeckeol (4.1%) and γ -muurolene (3.9%); for *H. kotschyianum*, α -pinene (14.4%), nonacosane (11.1%), hexadecanoic acid (9.2%), β -pinene (8.7%), spathulenol (6.3%) and limonene (5.1%). *H. salsugineum* showed nonacosane (42.7%), hexadecanoic acid (23.2%) and baeckeol (6.1%), and for *H. thymopsis* α -pinene (44.0%), baeckeol (32.9%), spathulenol (8.0%), limonene (7.6%) and camphene (5.2%) [18].

Hydrocarbons, such as 3-methyl-nonane (12.3%), 2-methyl-octane (9.6%) and nonane (7.6%) were also present among main components of *H. laricifolium* essential oil in the current investigation; related components have also been reported, previously, in high concentrations for other *Hypericum* species [12,13,15-18].

However, one particular component identified in this study caught the attention, since is very rarely detected in essential oils. Verticilol is a cembranoid diterpene that has been reported in the essential oil of several species belonging to *Burseraceae*, *Lauraceae* and *Cupressaceae* families [19-21]. Particularly, *B. elongata* (*Burseraceae*) oil is mainly composed by verticilol (52.4%); authors revealed that this particular compound has also been previously reported for *Boswellia elongata* collected from a different location in Yemen [19]. However, to date this component has not been identified in any *Hypericum* species. To the best of our knowledge, this is the first report on the chemical composition of the *Hypericum laricifolium* essential oil.

Acknowledgments

Authors are grateful to Dr. Alfredo Usubillaga and Dr. Rosa Aparicio, Faculty of Pharmacy and Biomedical Sciences, University of Los Andes, for helping in performing the GC/MS chromatogram. We also want to express our gratitude to CDCHT-ULA for the partial financial support (SE-FA-02-12-03).

References

- Stevens PF (2007) *Hypericaceae*. In: Kubitzki K (Ed), *The families and genera of vascular plants*, Springer Verlag, Berlin, Germany 194-201.
- APG III (2009) An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Bot J Linn Soc* 161: 105-121.
- Babakhanlu P, Mirza M, Sefidkon F, Ahmadi L, Barazande MM, et al. (1999) Identification of essential oil of *Hypericum perforatum*. *Med Aromat Plant Res* 200: 93-102.
- Rotblatt M, Ziment I (2002) Evaluation of 65 selected herbs: Evidence-based herbal medicine. Hanley and Belfus, Philadelphia, USA 66-379.
- Butterweek V (2003) Mechanism of action of St. John's wort in depression. What is known? *CNS Drugs* 17: 539-562.
- Greeson JM, Sanford B, Monti DA (2001) St. John's wort (*Hypericum perforatum* L.): A review of the current pharmacological, toxicological and clinical literature. *Psychopharmacology* 153: 402-414.
- Patocka J (2003) The chemistry, pharmacology and toxicology of the biologically active constituents of the herbs *Hypericum perforatum* L. *J Appl Biomed* 1: 61-70.

8. Wabo HK, Kowa TK, Lonfouo AH, Tchinda AT, Kikuchi H, et al. (2012) Phenolic compounds and terpenoids from *Hypericum lanceolatum*. Rec Nat Prod 6: 94-100.
9. Ghasemi Y, Khalaj A, Mohagheghzadeh A, Reza A, Hossein M (2007) Composition and antimicrobial activity of the essential oil and extract of *Hypericum elongatum*. J Appl Sci 7: 2671-2675.
10. Adams RP (2007) Identification of essential oil components by Gas chromatography/Mass spectrometry. (4th Ed.), Allured Publishing Corporation. Carol Stream IL, USA.
11. Davies NW (1990) Gas chromatographic retention indices of monoterpenes and sesquiterpenes on methyl silicon and carbowax 20 M phases. J Chromatogr A 503: 1-24.
12. Yuce E, Bagci E (2012) The essential oils of the aerial parts of two *Hypericum* taxa (*Hypericum triquetrifolium* and *Hypericum aviculariifolium* subsp. *depilatum* var. *depilatum* (Clusiaceae)) from Turkey. Nat Prod Res 26: 1985-1990.
13. Radulovića N, Đorđevića A, Palića R, Zlatkovićb B (2010) Essential oil composition of *Hypericum annulatum* Moris (Hypericaceae) from Serbia. J Essent Oil Res 22: 619-624.
14. Đorđević A, Zlatković B, Lazarević J, Palić R (2011) A detailed chemical composition and antimicrobial activity of *Hypericum richeri* Vill. Subsp. *grisebachii* (Boiss.) Nyman essential oil from Serbia. J Med Plants Res 5: 5486-5492.
15. Radulović NS, Blagojević PD (2012) Chemical composition of *Hypericum rumeliacum* Boiss. essential oil. A new chemotype of this pharmacologically valuable species?. Chem Biodivers 9: 2324-2341.
16. Chauhan RS, Vashistha RK, Nautiyal MC, Tava A, Cecotti R (2011) Essential oil composition of *Hypericum perforatum* L. from cultivated source. J Essent Oil Res 23: 20-25.
17. Sharopov SF, Gulmurodov IS, Setzer WN (2010) Essential oil composition of *Hypericum perforatum* L. and *Hypericum scabrum* L. growing wild in Tajikistan. J Chem Pharm Res 2: 284-290.
18. Özkan EE, Demirci B, Gürer ÇÜ, Kültür Ş, Mat A, et al. (2013) Composition of essential oils from five endemic hypericum species of Turkey. Organic Chem Curr Res 2: 1-4.
19. Awadh-Ali NA, Wurster M, Arnold N, Teichert A, Schmidt J, et al. (2008) Chemical composition and biological activities of essential oils from the *Oleogum resins* of three endemic *Soqotraen boswellia* species. Rec Nat Prod 2: 6-12.
20. Cheng SS, Liu JY, Hsui YR, Chang ST (2006) Chemical polymorphism and antifungal activity of essential oils from leaves of different provenances of indigenous cinnamon (*Cinnamomum osmophloeum*). Bioresour Technol 97: 306-312.
21. Khajjak MH, Raza AM, Shawani MN, Ahmed F, Shaheen G, et al. (2012) Comparative analysis of essential oil contents of *Juniperus excelsa* (M. Beib.) found in Balochistan, Pakistan. Afr J Biotechnol 11: 8154-8159.

This article was originally published in a special issue, **Bioactive essential oils: Essential oil as a source of bioactive constituents** handled by Editor(s).
Dr. RC Padalia, CCSIR- Central Institute of Medicinal & Aromatic, India