

Open Access

Characterization of the Essential Oil Composition of *Isodon rugosus* (Wall. ex Benth.) Codd. from Garhwal region of Western Himalaya

Verma RS^{1*}, Pandey V², Chauhan A¹ and Tiwari R²

¹CSIR-Central Institute of Medicinal and Aromatic Plants (CSIR-CIMAP), Research Centre, Pantnagar, P.O.-Dairy farm Nagla, Udham Singh Nagar, Uttarakhand-263149, India

²CSIR-Central Institute of Medicinal and Aromatic Plants (CSIR-CIMAP) Lucknow, 226015, Uttar Pradesh, India

Abstract

Background: *Isodon rugosus* belongs to the family Lamiaceae and is an important aromatic shrub, distributed widely throughout the northern temperate regions of the Himalaya. Regardless of the various traditional usage and diverse biological activities, little data exists on the essential oil composition of *I. rugosus* populations. This study aimed to characterize the essential oil of *I. rugosus* collected from two locations (Ghangaria and Tapovan) of Garhwal region of western Himalaya for occurrence of new composition.

Methods: The shade dried plant materials were hydrodistilled in a Clevenger's type apparatus for 3 h for isolation of their essential oils. The resulting essential oils were analysed using gas chromatography-flame ionization detector (GC-FID) and GC-mass spectrometry (GC-MS).

Result: Altogether, 92 constituents, comprising 83.9-92.6% of the total oil compositions were identified. Major constituents of the oil were α -pinene (0.8-19.3%), *trans*-ferruginol (3.0-17.1%), α -phellandrene (2.6-10.5%), (*E*)-caryophyllene (4.0-9.3%), germacrene D (1.9-8.8%), abietatriene (1.9-5.3%), β -phellandrene (3.1-4.4%), δ -cadinene (2.9-3.6%), limonene (1.3-3.6%), myrcene (0.2-3.6%) and *p*-cymene (2.9-3.5%). The examined oil was considerably different in respect of their qualitative and quantitative compositions compared to earlier studies. Moreover, to the best of our knowledge, 52 constituents, including *trans*-ferruginol, abietatriene, totarene, phyllocladene, 4-*epi*-abietal, caryophyllene acetate, cubenol, allo-hedycaryol, δ -amorphene, *trans*-cadina⁻¹ (6),4-diene, *cis*-cadina⁻¹ (6),4-diene, α -muurolol, amorpha-4,9-dien-2-ol, (*Z*)-nuciferol acetate and viridiflorol were reported for the first time in *I. rugosus* essential oil.

Conclusion: The dominant presence of sesquiterpenoids and diterpenoids in the 'Ghangaria' population and monoterpenoids and sesquiterpenoids in the 'Tapovan' population made them novel compositions. The newly identified constituents, especially diterpenoids can be of chemotaxonomic significance.

Keywords: Isodon rugosus; Lamiaceae; Essential oil composition; a-Pinene; trans-Ferruginol

Introduction

Isodon rugosus (Wall. ex Benth.) Codd (syn. Plectranthus rugosus Wall. ex. Benth.) belongs to the family Lamiaceae and is an aromatic branched shrub, with erect stem possessing ovate and opposite leaves of notched margin and covered with dense small hairs on ventral side. It is distributed widely throughout the northern temperate regions of the Himalaya at the elevation of 1500-2500 m [1,2]. The plant flowers from July to September and the seeds ripen from August to October [3]. The extracts of fresh and dried leaves have shown to be efficacious against worm infestations. It emanates a characteristic aroma and enjoys a reputation of being an antiseptic, germicidal and cardiac stimulant in folk medicine [4]. The plant is used in Pakistani traditional medicine for toothache and is claimed to be effective as an antiseptic, a hypoglycaemic, an antidiarrheal and a bronchodilator [5,6]. A topical administration of fresh leaf extract is used to treat scabies for its immediate effect, while 1-2 drops of this extract are used to treat earache [7]. An extract of the leaves is also used to treat hypertension, fevers, rheumatism and toothache. Branches are used for making dusters [8,9]. The plant extracts and fractions of different solvents exhibited antifungal [3], antibacterial, phytotoxic [10], antioxidant, and lipoxygenase inhibitory activities [11,12]. Besides various medicinal properties, it is also be used for phytoremediation [13]. Phytochemical studies carried out on the plant revealed the presence of steroids, flavonoids, terpenoids, saponin, tannins, cardiac glycosides, coumarins, reducing sugars and β -cyanin. Diterpenoids, namely, rugosinin, effusanin-A, effusanin-B, effusanin-E, lasiokaurin and oridonin have been isolated from the plant [14,15]. In addition to these, triterpenoids, namely, plectranthoic acid A and B, acetyl plectranthoic acid and plectranthadiol have also been isolated from the plant [16,17]. Moreover, studies on the essential oil composition indicate the presence of sesquiterpene hydrocarbons, including β -caryophyllene, germacrene-D and α -humulene as the major constituents [18,19]. Despite the multipurpose usage, there exists merger data on the chemical composition of *I. rugosus* populations growing in temperate Himalayan region. Therefore, in continuation of our research on the essential oil constituents of underexplored aromatic species of Indian flora, the present study was planned to carry out detailed GC-FID and GC-MS investigation of *I. rugosus* essential oils extracted from the populations growing at higher altitudes of Garhwal region of Uttarakhand, India. This study also aimed to characterize several new

*Corresponding author: Verma RS, CSIR-Central Institute of Medicinal and Aromatic Plants (CSIR-CIMAP), Research Centre, Pantnagar, P.O.-Dairy farm Nagla, Udham Singh Nagar, Uttarakhand-263149, India, Tel: +91-5944234445; E-mail: rs.verma@cimap.res.in; rswaroop1979@yahoo.com

Received May 12, 2015; Accepted June 17, 2015; Published June 22, 2015

Citation: Verma RS, Pandey V, Chauhan A, Tiwari R (2015) Characterization of the Essential Oil Composition of *Isodon rugosus* (Wall. ex Benth.) Codd. from Garhwal region of Western Himalaya. Med Aromat Plants S3: 002. doi:10.4172/2167-0412. S3-002

Copyright: © 2015 Verma RS, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

constituents for *I. rugosus* essential oils in addition to earlier reported constituents. The newly identified constituents, especially diterpenoids of the essential oil can be of immense chemotaxonomic value.

Materials and Methods

Plant material and isolation of essential oil

Plant materials (*I. rugosus*) were collected from Ghangaria (30.681851° N and 79.589786° E; altitude 2687.26 m) and Tapovan (30.491288° N and 79.628392° E; altitude: 1917.67 m) regions of Chamoli, Uttarakhand during the third week of August, 2012. The populations growing in Ghangaria and Tapovan regions were in flowering and vegetative phases, respectively during collection time. The plant materials were authenticated at Botany Department of CSIR-CIMAP Research Centre Pantnagar by one of the authors (AC). The shade dried plant materials were hydrodistilled in a Clevenger's apparatus for 3 h to collect their essential oils. The essential oil content (% v/w) was estimated on dry weight basis. The oils obtained were dehydrated over anhydrous sodium sulphate and kept in a cool and dark place until further analyses.

Gas chromatography (GC/FID)

GC analysis of the essential oils was carried out on a PerkinElmer AutoSystem XL gas chromatograph, equipped with DB-5 capillary column (60 m \times 0.32 mm i.d., film thickness 0.25 µm) and flame ionization detector (FID). The oven column temperature ranged from 70-250°C, programmed at 3°C minute⁻¹, with initial and final hold time of 2 minute, using H₂ as carrier gas at 10 psi constant pressure, a split ratio of 1:35, an injection size of 0.03 µL neat, and injector and detector temperatures were maintained at 250°C and 280°C, respectively.

Gas chromatography-mass spectrometry (GC-MS)

GC-MS analysis of the essential oil samples was carried out on a Clarus 680 GC interfaced with a Clarus SQ 8C mass spectrometer of PerkinElmer fitted with Elite-5 MS fused-silica capillary column (30 m \times 0.25 mm i.d., film thickness 0.25 µm). The oven temperature program was from 60-240°C, at 3°C minute⁻¹, and programmed to 270°C at 5°C minute⁻¹; injector temperature was 250°C; transfer line and source temperatures were 220°C; injection size 0.03 µL neat; split ratio 1:50; carrier gas He at 1.0 mL minute⁻¹; ionization energy 70 eV; mass scan range 40-450 amu.

Identification of essential oil constituents

Identification of the essential oil constituents was done on the basis of retention index (RI) determined using a homologous series of *n*-alkanes (C_8 - C_{30} , Supelco Bellefonte, PA, USA) under identical experimental conditions, co-injection with known essential oil constituents, mass spectra library search (NIST/EPA/NIH, version 2.0 g, and Wiley registry of mass spectral data 9th edition) and by comparing the mass spectral and retention data with literature [20]. The relative amounts of individual components were calculated based on the GC peak area (FID response) without using a correction factor.

Results and Discussion

The essential oil yield was observed to be 0.15% and 0.25% (v/w) on dry weight basis in 'Ghangaria' and 'Tapovan' populations of *I. rugosus*, respectively. These variations were probably due to the variations of plant stages and geographical conditions. The resulting essential oils were subjected to GC-FID and GC-MS analyses. A total of 81 constituents, forming 92.6% of the total oil composition

of 'Ghangaria' population and 80 constituents, comprising 83.9% of total oil composition of 'Tapovan' population were identified using retention index (RI) and mass spectrum of the individual components (Table 1). Major constituents of the oils were α -pinene (0.8-19.3%), *trans*-ferruginol (3.0-17.1%), α -phellandrene (2.6-10.5%), (*E*)-caryophyllene (4.0-9.3%), germacrene D (1.9-8.8%), abietatriene (1.9-

S. no.	Compound	RI	Content		Identification
			(70) I	, II	methods⁰
1	α-Thuiene [†]	926	t	0.1	RI. MS
2	α-Pinene	932	0.8	19.3	RI. MS
3	Camphene	948	t	0.2	RI, MS
4	Sabinene	974	-	0.3	RI, MS
5	1-Octen-3-ol [†] + β-pinene	978	0.1	0.4	RI. MS
6	Mvrcene	988	0.2	3.6	RI. MS
7	α-Phellandrene	1005	2.6	10.5	RI, MS
8	δ-3-Carene	1010	0.3	1.0	RI, MS
9	α-Terpinene	1016	0.2	0.3	RI, MS
10	p-Cymene	1022	3.5	2.9	RI, MS
11	o-Cymene [†]	1024	t	t	RI, MS
12	Limonene	1027	1.3	3.6	RI, MS
13	β-Phellandrene	1028	3.1	4.4	RI, MS
14	1,8-Cineole	1030	0.2	0.2	RI, MS
15	(Z)-β-Ocimene	1034	-	0.1	RI, MS
16	(<i>E</i>)-β-Ocimene	1045	t	0.4	RI, MS
17	γ-Terpinene	1053	0.3	0.5	RI, MS
18	cis-Sabinene hydrate [†]	1063	0.4	0.3	RI, MS
19	Terpinolene	1089	0.6	1.3	RI, MS
20	p-Cymenene [†]	1091	t	-	RI, MS
21	Linalool	1097	0.3	0.2	RI, MS
22	trans-Sabinene hvdrate [†]	1099	t	-	RI, MS
23	<i>n</i> -Undecane [†]	1102	-	t	RI, MS
24	cis-Thujone	1104	t	-	RI, MS
25	cis-p-Menth-2-en-1-ol	1122	t	t	RI, MS
26	trans-p-Menth-2-en-1-ol	1139	t	t	RI, MS
27	Camphor [†]	1145	0.1	-	RI, MS
28	Terpinen-4-ol	1177	0.6	0.3	RI, MS
29	p-Cymen-8-ol [†]	1182	t	t	RI, MS
30	α-Terpineol	1190	t	t	RI, MS
31	<i>trans</i> -p-Mentha ⁻¹ (7),8- dien-2-ol [†]	1191	t	t	RI, MS
32	γ-Terpineol [†]	1197	t	t	RI, MS
33	β-Cyclocitral [†]	1215	0.2	t	RI, MS
34	Carvacrol methyl ether [†]	1245	t	-	RI, MS
35	Piperitone [†]	1255	t	-	RI, MS
36	Bornyl acetate	1286	t	t	RI, MS
37	Thymol [†]	1288	t	t	RI, MS
38	Carvacrol [†]	1300	t	t	RI, MS
39	α-Cubebene	1353	0.3	0.1	RI, MS
40	α-Copaene	1378	0.5	0.3	RI, MS
41	β-Bourbonene	1390	0.7	0.2	RI, MS
42	Isolongifolene [†]	1393	0.2	0.3	RI, MS
43	β-Elemene	1394	0.2	0.3	RI, MS
44	(Z)-Caryophyllene [†]	1409	-	0.1	RI, MS
45	α-Gurjunene [†]	1411	0.3	0.6	RI, MS
46	β-Funebrene [†]	1415	-	0.1	RI, MS
47	(E)-Caryophyllene	1420	9.3	4.0	RI, MS
48	β-Cedrene [†]	1421	-	t	RI, MS
49	β-Gurjunene	1435	1.7	0.2	RI, MS
50	cis-Muurola-3,5-diene ⁺	1452	-	t	RI, MS
51	α-Humulene	1454	1.2	1.0	RI, MS

Citation: Verma RS, Pandey V, Chauhan A, Tiwari R (2015) Characterization of the Essential Oil Composition of *Isodon rugosus* (Wall. ex Benth.) Codd. from Garhwal region of Western Himalaya. Med Aromat Plants S3: 002. doi:10.4172/2167-0412.S3-002

52	allo-Aromadendrene [†]	1461	0.3	-	RI, MS			
53	cis-Cadina-1(6),4-diene†	1465	0.9	1.2	RI, MS			
54	trans-Cadina-1(6),4-dienet	1478	2.2	1.2	RI, MS			
55	Germacrene D	1483	8.8	1.9	RI, MS			
56	β-Selinene [†]	1494	0.2	0.1	RI, MS			
57	γ-Amorphene [†]	1500	0.6	0.3	RI, MS			
58	α-Muurolene	1502	2.8	0.7	RI, MS			
59	<i>trans</i> -β-Guaiene [†]	1504	0.7	0.5	RI, MS			
60	Germacrene A [†]	1508	-	t	RI, MS			
61	α-Bulnesene [†]	1509	-	t	RI, MS			
62	δ-Amorphene [†]	1513	1.5	0.8	RI, MS			
63	γ-Cadinene	1515	1.0	0.2	RI, MS			
64	Cubebol	1518	1.7	2.8	RI, MS			
65	δ-Cadinene	1524	3.6	2.9	RI, MS			
66	α-Cadinene [†]	1542	0.1	0.2	RI, MS			
67	Germacrene-D-4-ol	1578	0.7	1.0	RI, MS			
68	Spathulenol	1582	1.1	0.1	RI, MS			
69	Caryophyllene oxide	1587	0.1	0.2	RI, MS			
70	allo-Hedycaryol [†]	1589	1.4	-	RI, MS			
71	Globulol [†]	1594	0.1	-	RI, MS			
72	Viridiflorol [†]	1596	0.4	0.7	RI, MS			
73	1,10-di-epi-Cubenol [†]	1620	0.4	0.1	RI, MS			
74	<i>epi</i> -α-Cadinol	1642	0.9	0.5	RI, MS			
75	α-Muurolol [†]	1650	0.5	0.9	RI, MS			
76	Cubenol [†]	1651	1.4	0.2	RI, MS			
77	α-Cadinol	1656	1.0	1.0	RI, MS			
78	Amorpha-4,9-dien-2-ol [†]	1698	0.8	-	RI, MS			
79	Caryophyllene acetate [†]	1703	1.0	1.2	RI, MS			
80	(Z)-Nuciferol [†]	1728	0.1	t	RI, MS			
81	(E)-Nuciferol [†]	1760	t	-	RI, MS			
82	(Z)-Nuciferol acetate [†]	1835	0.8	-	RI, MS			
83	Totarene [†]	1925	2.0	0.6	RI, MS			
84	Sandaracopimara- 8(14),15-diene [†]	1970	0.3	0.2	RI, MS			
85	13-epi-Dolabradiene [†]	2003	0.3	0.2	RI, MS			
86	Phyllocladene [†]	2019	1.9	1.2	RI, MS			
87	Abietatriene [†]	2060	5.3	1.9	RI, MS			
88	Abietadiene [†]	2090	0.4	0.3	RI, MS			
89	Nezukol [†]	2137	-	0.4	RI, MS			
90	Abieta-8(14),13(15)- diene [†]	2151	-	0.1	RI, MS			
91	4- <i>epi</i> -Abietal⁺	2302	1.0	0.2	RI, MS			
92	trans-Ferruginol [†]	2336	17.1	3.0	RI, MS			
Class composition								
	Monoterpene hydrocarbons		9.4	45.6				
	Oxygenated monoterpenes		1.8	1.0				
	Sesquiterpene hydrocarbons		37.1	17.2				
	Oxygenated		12.4	8.7				
	Diterpene hydrocarbons		10.2	4 5				
	Oxygenated ditemenes		18.1	3.6				
	Benzenoid compounds		3.5	2.9				
	Aliphatic compounds		0.0	0.4				
	Total identified		92.6	83.9				

RI: Retention Index determined on DB-5 gas chromatography column (30 m \times 0.25 mm) using a homologous series of *n*-alkanes; I: Ghangaria; II: Tapovan; Identification of the compound based on RI (Retention Index), MS (Mass Spectrometry); †Reported for the first time in *Isodon rugosus* essential oil; t: Trace (<0.05%).

Table 1: Essential oil composition of *Isodon rugosus* collected from Garhwal region of western-Himalaya, India.

5.3%), β-phellandrene (3.1-4.4%), δ-cadinene (2.9-3.6%), limonene (1.3-3.6%), myrcene (0.2-3.6%), p-cymene (2.9-3.5%), cubebol (1.7-2.8%), α-muurolene (0.7-2.8%), *trans*-cadina⁻¹ (6),4-diene (1.2-2.2%), totarene (0.6-2.0%), phyllocladene (1.2-1.9%), β-gurjunene (0.2-1.7%), and δ -amorphene (0.8-1.5%). The oil of 'Ghangaria' population was dominated by sesquiterpenoids (sesquiterpene hydrocarbons: 37.1%; oxygenated sesquiterpenes: 12.4%), followed by diterpenoids (sesquiterpene hydrocarbons: 10.2%; oxygenated sesquiterpenes: 18.1%). However, the oil of 'Tapovan' population was characterised by mainly monoterpenoids (monoterpene hydrocarbons: 45.6%; oxygenated monoterpenes: 1.0%) and sesquiterpenoids (sesquiterpene hydrocarbons: 17.2%; oxygenated sesquiterpenes: 8.7%). Major constituents of the oil of 'Ghangaria' population were trans-ferruginol (17.1%), (E)-caryophyllene (9.3%), germacrene D (8.8%), abietatriene (5.3%), δ-cadinene (3.6%), p-cymene (3.5%), β-phellandrene (3.1%), α-muurolene (2.8%), α-phellandrene (2.6%), *trans*-cadina⁻¹(6),4-diene (2.2%), totarene (2.0%), phyllocladene (1.9%), β-gurjunene (1.7%), cubebol (1.7%), δ -amorphene (1.5%), allo-hedycaryol (1.4%), cubenol (1.4%), limonene (1.3%) and a-humulene (1.2%). On the other hand, the characteristic constituents of 'Tapovan' population were α-pinene (19.3%), α-phellandrene (10.5%), β-phellandrene (4.4%), (E)-caryophyllene (4.0%), myrcene (3.6%), limonene (3.6%), transferruginol (3.0%), p-cymene (2.9%), δ-cadinene (2.9%), cubebol (2.8%), germacrene D (1.9%), abietatriene (1.9%), terpinolene (1.3%), cis-cadina-1 (6),4-diene (1.2%), trans-cadina-1 (6),4-diene (1.2%), caryophyllene acetate (1.2%) and phyllocladene (1.2%). Thus, comparison of the results clearly showed that I. rugosus populations growing in two different locations of Himalaya had considerable quantitative variation in their essential oil compositions.

Page 3 of 4

The essential oil composition of *I. rugosus* have been studied in past by few researchers. All the reported compositions of I. rugosus (syn P. rugosus) were characterised by higher amounts of sesquiterpenoids. Weyerstahl et al (1983) reported caryophyllene (22.0%) as main constituents along with germacrene D, a-phellandrene, a-pinene, caryophyllene oxide, α -cadinol, δ -cadinene, limonene, β -phellandrene, myrcene and *p*-cymene [21]. Tiwari et al reported β -caryophyllene (38.4%), germacrene D (23.8%), spathulenol (3.2%), a-cadinol (2.2%), p-cymene (3.6%), y-terpinene (2.8%) and limonene (2.7%) as major constituents of the oil [18]. Similarly, the essential oil of the population collected from Nainital (Kumaon region, western Himalaya) was dominated by sesquiterpene hydrocarbons, represented by β-caryophyllene (29.8-36.2%), germacrene D (25.2-28.2%), α-humulene (6.6-8.6%) and (E)-β-farnesene (2.3-3.8%) as the major constituents [19]. However, recently Irshad et al reported spathulenol (21.0%), β-caryophyllene (10.6%), germacrene D (20.0%), 3-carene (4.8%), myrcene (4.0%), a-myrecene (4.0%) and limonene (2.7%) as major constituents of I. rugosus essential oil [22]. Thus, the chemical compositions of I. rugosus examined in this study showed remarkable qualitative and quantitative differences as compared to the compositions reported earlier from other countries / locations. These variations are likely to be due to both biotic and abiotic factors affecting plant growth and biosynthesis.

Ferruginol (meroterpene), an oxidized abietane diterpene, has led to several synthetic and biological studies due to their interesting structure and potent biological profile [23,24]. The α -pinene, a monoterpene hydrocarbon, possesses antimalarial activity [25]. It is used as an important substance in the manufacture of a variety of synthetic aroma chemicals and its epoxide is isomerised to produce campholenic aldehyde, which is an intermediate for the sandalwood fragrance, santalol [26]. Beside the presence of several common terpenoids, *I*. Citation: Verma RS, Pandey V, Chauhan A, Tiwari R (2015) Characterization of the Essential Oil Composition of *Isodon rugosus* (Wall. ex Benth.) Codd. from Garhwal region of Western Himalaya. Med Aromat Plants S3: 002. doi:10.4172/2167-0412.S3-002

rugosus growing in Garhwal region of Himalayan can be a good source of rarely occurring terpenoids such as *trans*-ferruginol and therefore can be further explored for sustainable utilization. According to the literature review, a total of 52 constituents are reported in *I. rugosus* essential oil till now [18,19,21,22]. However, this study identified a total of 92 constituents, which included 40 previously reported constituents. Thus, to the best of our knowledge, about 52 constituents were being reported for the first time for *I. rugosus* essential oil (marked in Table 1). The newly identified constituents of the oil can play an important role in oil authentication and chemotaxonomic studies of this Himalayan *Isodon* species. Moreover, the dominant presence of sesquiterpenoids and diterpenoids in the 'Ghangaria' population and monoterpenoids and sesquiterpenoids in the 'Tapovan' population made them novel compositions of *I. rugosus*.

Acknowledgments

CSIR, New Delhi is thankfully acknowledged for the financial support through XII-FYP project (BSC-0203) to carrying out the work. Authors are also thankful to the Director, CSIR-Central Institute of Medicinal and Aromatic Plants for encouragement and to the Central Chemical Facility (CSIR-CIMAP) for providing facility for GC and GC-MS analyses.

References

- Gupta RK (1968) Flora of Nainitalensis, a handbook of the flowering plants of Nainital, Navyug Publication, New Delhi 270.
- 2. Polunin O, Stainton A (1984) Flowers of the Himalaya, Oxford University Press, New Delhi 321.
- Rauf A, Khan A, Rasool S, Shah ZA, Saleem M (2012) In-vitro antifungal activity of three selected Pakistani medicinal plants. Middle East Journal of Medicinal Plants Research 1: 41-43.
- Singh V (1994) Herbal remedies for worm infestation in Kashmir Himalaya. Fitoterapia 65: 354-357.
- Ajmal SM, Mohammad S, Zahid K, Bakht Z (2012) Ethnomedicinal and phytoeconomic elaboration of Lilownai valley, district Shangla Pakistan. International Research Journal of Pharmacy 3: 164-169.
- Sher Z, Khan Z, Hussain F (2011) Ethnobotanical studies of some plants of Chagharzai valley, district Buner, Pakistan. Pakistan Journal of Botany 43: 1445-1452.
- Sabeen M, Ahmad SS (2009) Exploring the folk medicinal flora of Abbotabad city, Pakistan. Ethnobotany Leaflets 13: 810-833.
- Akhtar N, Rashid A, Murad W, Bergmeier E (2013) Diversity and use of ethnomedicinal plants in the region of Swat, North Pakistan. J Ethnobiol Ethnomed 9: 25.
- Khan SW, Khatoon S (2007) Ethnobotanical studies on useful trees and shrubs of Haramosh and Bugrote valleys, in Gilgit northern areas of Pakistan. Pakistan Journal of Botany 39: 699-710.

10. Rauf A, Muhammad N, Khan A, Uddin N, Atif M (2012) Antibacterial and phytotoxic profile of selected Pakistani medicinal plants. World Applied Sciences Journal 20: 540-544.

Page 4 of 4

- Janbaz KH, Arif J, Saqib F, Imran I, Ashraf M, et al. (2014) In-vitro and in-vivo validation of ethnopharmacological uses of methanol extract of Isodon rugosus Wall. ex Benth. (Lamiaceae). BMC Complement Altern Med 14: 71.
- Rauf A, Uddin G, Ali M, Muhammad N, Gul S (2013) Phytochemical screening and antioxidant activity of Pakistani medicinal plants. Wudpecker Journal of Medicinal Plants 2: 1-6.
- Muhammad S, Shah MT, Khan S, Saddique U, Gul N, et al. (2013) Wild plant assessment for heavy metal phytoremediation potential along the mafic and ultramafic terrain in Northern Pakistan. BioMed Research International.
- Abbaskhan A, Choudhary MI, Tsuda Y, Parvez M, Atta-ur-Rahman, et al. (2003) A new diepoxy-ent-kauranoid, rugosinin, from Isodon rugosus. Planta Med 69: 94-96.
- Sun HD, Huang SX, Han QB (2006) Diterpenoids from Isodon species and their biological activities. Nat Prod Rep 23: 673-698.
- Razdan TK, Kachroo V, Harkar S, Koul GL (1982) Plectrantholic acids A and B - two new triterpenoids from Plectranthus rugosus. Tetrahedron 38: 991-992.
- Razdan TK, Kachroo V, Harkar S, Koul GL, Dhart KL (1982) Plectranthoic acid, acetyl plectranthoic acid and plectranthadiol-three new triterpenoids from Plectranthus rugosus. Phytochemistry 21: 409-412.
- Tiwari A, Padalia RC, Mathela CS (2008) Sesquiterpene rich essential oil from Plectranthus rugosus Wall. Journal of Essential Oil Bearing Plants 11: 58-61.
- Padalia RC, Verma RS (2011) Comparative study of volatile oil compositions of two Plectranthus species from northern India. Nat Prod Res 25: 1727-1732.
- Adams RP (2007) Identification of essential oil components by gas chromatography/mass spectrometry. Allured Publishing Corp, Carol Stream, Illinois, USA.
- Weyerstahl P, Kaul VK, Meier N, Weirauch M, Marschall H (1983) Volatile constituents of Plectranthus rugosus leaf oil. Planta Med 48: 99-102.
- 22. Irshad M, Aziz S, Rehman H, Hussain H (2012) GC-MS analysis and antifungal activity of essential oils of Angelica glauca, Plectranthus rugosus, and Valeriana wallichii. Journal of Essential Oil Bearing Plants 15: 15-21.
- Gonzalez MA, Perez-Guaita D (2012) Short syntheses of (+)-ferruginol from (+) dehydroabietylamine. Tetrahedron 68: 9612-9615.
- Son KH, Oh HM, Choi SK, Han DC, Kwon BM (2005) Anti-tumor abietane diterpenes from the cones of Sequoia sempervirens. Bioorg Med Chem Lett 15: 2019-2021.
- Van Zyl RL, Seatlholo ST, Van Vuuren SF, Viljoen AM (2006) The biological activity of 20 nature identical essential oil constituents. Journal of Essential Oil Research 18: 129-133.
- Suh YW, Kim NK, Ahn WS, Rhee HK (2003) One-pot synthesis of campholenic aldehyde from a-pinene over Ti-HMS catalyst II: effects of reaction conditions. Journal of Molecular Catalysis A: Chemical 198: 309-316.

This article was originally published in a special issue, **Plant Medicines** Applications in Therapeutics handled by Editor(s). Dr. Ana Isabel Faria Ribeiro, Tropical Research Institute (IICT), Portugal