

# Application Prospects of the *Caenorhabditis Elegans* Model in Toxicology and Forensic Science

Yushu Zhang, Zhuo Li, Wenxing Yang, Kui Zhang\*

West China School of Basic Medical Sciences and Forensic Medicine, Sichuan University, Chengdu 610041, China

## ABSTRACT

This review summarizes the current applications of *Caenorhabditis elegans* (referred to as *C. elegans*) as a model organism in toxicological research and explores its prospects in the field of forensic science. *C. elegans*, with its short life cycle, clear genetic background and highly conserved genome, has demonstrated unique advantages in toxicological studies. Starting from its biological characteristics, this paper introduces the toxicological research methods and progress based on the *C. elegans* model and focuses on its applications in environmental forensic medicine and forensic toxicology.

**Keywords:** Forensic toxicology; Environmental forensic medicine; *Caenorhabditis elegans*; Model organism

## DESCRIPTION

### Biological characteristics of *C. elegans*

*C. elegans* is a nonparasitic, soil-dwelling nematode with the following notable biological characteristics:

**Short life cycle:** Under laboratory conditions, *C. elegans* can complete a generation turnover in about 4 days, with low maintenance costs [1].

**Clear genetic background:** The genome of *C. elegans* has been fully sequenced, with 60%-80% of its genes being homologous to those of humans [2]. The experimental data obtained are highly consistent with human detection data.

**Complete nervous system:** The nervous system of *C. elegans* has been fully mapped, with some neurotransmitters showing high similarity to those in higher mammals [3-7].

**Rich behavioral traits:** *C. elegans* exhibits clear, diverse, and easily quantifiable behavioral traits.

**Transgenic strains:** Additionally, since the *C. elegans* genome has been fully annotated, its model has been widely used in genetics, molecular biology and cell biology. Transgenic strains based on the Green Fluorescent Protein (GFP) gene, such as CL2166 (*gst4::GFP*) and SJ4005 (*hsp4::GFP*), have been widely used in toxicological research. These strains are mostly preserved

in the *Caenorhabditis* Genetics Center (CGC) and are available for open access, providing convenience for relevant research [8,9].

### Toxicological research methods using the *C. elegans* model

Toxicological studies using the *C. elegans* model typically simulate real world exposure patterns of toxicants, including liquid-phase, solid-phase and gas-phase exposures. The toxic effects of substances are assessed through various biological indicators, such as survival rate, reproductive capacity, growth and development and behavioral phenotypes [10-19]. In recent years, the *C. elegans* model, with its unique biological characteristics, has shown high compatibility and synergistic effects with various cutting-edge techniques. These include high-throughput technologies, fluorescence screening, metabolomics, microfluidic chip technology, and gene-editing techniques (such as CRISPR/Cas9) to precisely modify the *C. elegans* genome, thereby revealing the toxic effects of substances at the molecular level [20-22].

### Research achievements of the *C. elegans* model in forensic science

**Environmental forensic medicine:** Significant achievements have been made in environmental toxicology using the *C. elegans*

**Correspondence to:** Zhuo Li, West China School of Basic Medical Sciences and Forensic Medicine, Sichuan University, Chengdu 610041, China, E-mail: leezhuo2017@163.com

**Received:** 02-Sep-2025, Manuscript No. JCT-25-38651; **Editor assigned:** 04-Sep-2025, PreQC No. JCT-25-38651 (PQ); **Reviewed:** 18-Sep-2025, QC No. JCT-25-38651; **Revised:** 23-Sep-2025, Manuscript No. JCT-25-38651 (R); **Published:** 30-Sep-2025, DOI: 10.35248/2475-3181.25.15.602

**Citation:** Zhang Y, Li Z, Yang W, Zhang K (2025). Application Prospects of the *Caenorhabditis Elegans* Model in Toxicology and Forensic Science. J Clin Toxicol.15:602.

**Copyright:** © 2025 Zhuo Li, et.al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

model, covering pesticides, nano industrial materials, organic pollutants and heavy metals [14,15,23-36]. These studies provide a scientific basis for environmental risk monitoring and the assessment of the correlation between pollutants and human health. For instance, the toxicity effects of trifluralin, such as increased oxidative stress levels and mitochondrial damage, are evaluated by observing the body length and pharyngeal pumping frequency, as well as the head swing frequency of *C. elegans* exposed to different concentrations of trifluralin [23].

**Forensic toxicology:** In forensic toxicology, the *C. elegans* model is used to study the molecular mechanisms of toxicant action, such as addiction mechanisms, teratogenic effects on offspring and the impact of neurotoxins on neurodegenerative diseases [37-40]. In recent years, at least 50 genes related to ethanol poisoning have been identified in the *C. elegans* model and some of these homologous genes have been confirmed to be associated with human ethanol addiction and metabolic disorders [41,42]. In addition, the *C. elegans* model is also used in genetic toxicology research to explore the transgenerational and multigenerational toxic effects of heavy metals and organic pollutants. Studies have shown that the mechanism of transgenerational toxic effects induced by heavy metals may be related to the overactivation of oxidative stress [43,44]. Toxicity from multigenerational exposure is often associated with the accumulation of toxicants in the body and phenotypic changes such as growth inhibition, reproductive damage, neuronal injury, and oxidative stress induction can usually be observed in *C. elegans* models exposed to toxicants over multiple generations.

**Outlook for the *C. elegans* model in forensic science:** The application of the *C. elegans* model in forensic identification faces challenges. In the future, it is necessary to further strengthen the integration with forensic practice. Metabolic mechanism data obtained from *C. elegans* toxicology experimental models should be used to establish open and updated standardized databases of metabolic mechanisms, which should then be correlated with human data from forensic identification. It is also necessary to improve the judicial interpretation rules for toxicological data and the construction of the evidence chain so that research findings can serve forensic identification.

## CONCLUSION

As the demand for elucidating the mechanisms of toxicant action and their metabolic pathways in the human body increases in forensic science, toxicological research based on the *C. elegans* model will offer a new perspective for judicial identification. Establishing a comprehensive database of metabolic mechanisms and continuously accumulating and updating relevant toxicological data will provide stronger support for interdisciplinary research in forensic and toxicological sciences.

## REFERENCES

1. Yunli Zhao JC, Rui Wang, Xiaoxiao Pu, Dayong Wang. A review of transgenerational and multigenerational toxicology in the *in vivo* model animal *Caenorhabditis elegans*. *J Appl Toxicol*. 2023;43:122-145.

2. Dolese DA, Junot MP, Ghosh B, Butsch TJ, Johnson AE, Bohnert KA, et al. Degradative tubular lysosomes link pexophagy to starvation and early aging in *C. elegans*. *Autophagy*. 2021;18:1522-1533.
3. Cook SJ, Jarrell TA, Brittin CA, Wang Y, Bloniarz AE, Yakovlev MA, et al. Whole-animal connectomes of both *Caenorhabditis elegans* sexes. *Nature*. 2019;571:63-71.
4. White JG, Southgate E, Thomson JN, Brenner S. The structure of the nervous system of the nematode *Caenorhabditis elegans*. *Phil Trans R Soc Lond B*. 1986;314:1-340.
5. Setty H, Salzberg Y, Karimi S, Berent-Barzel E, Krieg M, Oren-Suissa M, et al. Sexually dimorphic architecture and function of a mechanosensory circuit in *C. elegans*. *Nature Communications*. 2022;13:6825.
6. Moroz LL, Romanova DY. Chemical cognition: Chemoconnectomics and convergent evolution of integrative systems in animals. *Anim. Cogn*. 2023;26:1851-1864.
7. Dag U, Nwabudike I, Kang D, Gomes MA, Kim J, Atanas AA, et al. Dissecting the functional organization of the *C. elegans* serotonergic system at whole-brain scale. *Cell*. 2023;186:2574-2592.
8. Wu J, Gao Y, Xi J, You X, Zhang X, Zhang X, et al. A high-throughput microplate toxicity screening platform based on *Caenorhabditis elegans*. *Ecotoxicol Environ Saf*. 2022:245.
9. Wu CW, Wimberly K, Pietras A, Dodd W, Atlas MB, Choe KP, et al. RNA processing errors triggered by cadmium and integrator complex disruption are signals for environmental stress. *BMC Biology*. 2019;17:56.
10. Wang Y, Zhang H, Wu X, Xue C, Hu Y, Khan A, et al. Ecotoxicity assessment of sodium dimethyldithiocarbamate and its micro-sized metal chelates in *Caenorhabditis elegans*. *Sci Total Environ*. 2020;720:137666.
11. Wang S, Chu Z, Zhang K, Miao G. Cadmium-induced serotonergic neuron and reproduction damages conferred lethality in the nematode *Caenorhabditis elegans*. *Chemosphere*. 2018;213:11-18.
12. Ngo LT, Huang WT, Chan MH, Su TY, Li CH, Hsiao M, et al. Comprehensive neurotoxicity of lead halide perovskite nanocrystals in nematode *Caenorhabditis elegans*. *Small*. 2023;20:2306020.
13. Currie SD, Doherty JP, Xue KS, Wang JS, Tang L. The stage-specific toxicity of per- and Polyfluoroalkyl Substances (PFAS) in nematode *Caenorhabditis elegans*. *Environ Pollut*. 2023;336:122429.
14. Abbass M, Chen Y, Arlt VM, Stürzenbaum SR. Benzo[a]pyrene and *Caenorhabditis elegans*: Defining the genotoxic potential in an organism lacking the classical CYP1A1 pathway. *Arch Toxicol*. 2021;95:1055-1069.
15. Bargmann CI, Hartwig E, Horvitz HR. Odorant-selective genes and neurons mediate olfaction in *C. elegans*. *Cell*. 1993;74:515-527.
16. Hiroki S, Yoshitane H, Mitsui H, Sato H, Umatani C, Kanda S, et al. Molecular encoding and synaptic decoding of context during salt chemotaxis in *C. elegans*. *Nat Commun*. 2022;13:2928.
17. Yang W, Wu T, Tu S, Qin Y, Shen C, Li J, et al. Redundant neural circuits regulate olfactory integration. *PLOS Genetics*. 2022;18:e1010029.
18. Sun Q, Liu C, Jiang K, Fang Y, Kong C, Fu J, et al. A preliminary study on the neurotoxic mechanism of harmine in *Caenorhabditis elegans*. *Comp Biochem Physiol C Toxicol Pharmacol*. 2021;245:109038.
19. Jeong A, Park SJ, Lee EJ, Kim KW. Nanoplastics exacerbate Parkinson's Mater. 2024;465:133289.
20. Peter E, Candido M, Jones D. Transgenic *Caenorhabditis elegans* strains as biosensors. *Trends Biotechnol*. 1996;14:125-129.

21. Paavanen-Huhtala S, Kalichamy K, Pessi AM, Häkkinen S, Saarto A, Tuomela M, et al. Biomonitoring of indoor air fungal or chemical toxins with *Caenorhabditis elegans* nematodes. *Pathogens*. 2023;12:161.
22. Wang Z. *Caenorhabditis elegans* as an *in Vivo* model organism to elucidate teratogenic effects. *Teratogenicity Testing*. 2024;283-306.
23. Liu H, Fu G, Li W, Liu B, Ji X, Zhang S, et al. Oxidative stress and mitochondrial damage induced by a novel pesticide fluopimomide in *Caenorhabditis elegans*. *Environmental Science and Pollution Research*. 2023;30:91794-91802.
24. Jacques MT, Soares MV, Farina M, Bornhorst J, Schwerdtle T, Ávila DS, et al. Impaired physiological responses and neurotoxicity induced by a chlorpyrifos-based formulation in *Caenorhabditis elegans* are not solely dependent on the active ingredient. *Environ Toxicol Pharmacol*. 2023;101:104196.
25. Bora S, Vardhan GS, Deka N, Khataniar L, Gogoi D, Baruah A, et al. Paraquat exposure over generation affects lifespan and reproduction through mitochondrial disruption in *C. elegans*. *Toxicol*. 2021;447:152632.
26. Gonzalez-Hunt CP, Luz AL, Ryde IT, Turner EA, Ilkayeva OR, Bhatt DP, et al. Multiple metabolic changes mediate the response of *Caenorhabditis elegans* to the complex I inhibitor rotenone. *Toxicol*. 2021;447:152630.
27. Qu M, Chen H, Lai H, Liu X, Wang D, Zhang X, et al. Exposure to nanoplastics and its 4 chemically modified derivatives at predicted environmental concentrations causes differently regulatory mechanisms in nematode *Caenorhabditis elegans*. *Chemosphere*. 2022;305:135498.
28. Jiang W, Yan W, Tan Q, Xiao Y, Shi Y, Lei J, et al. The toxic differentiation of micro- and nanoplastics verified by gene-edited fluorescent *Caenorhabditis elegans*. *Sci Total Environ*. 2023;856:159058.
29. Wang M, Feng Y, Cao Z, Yu N, Wang J, Wang X, et al. Multiple generation exposure to ZnO nanoparticles induces loss of genomic integrity in *Caenorhabditis elegans*. *Ecotoxicol Environ Saf*. 2023;249:114383.
30. Chen J, Chen C, Wang N, Wang C, Gong Z, Du J, et al. Cobalt nanoparticles induce mitochondrial damage and  $\beta$ -amyloid toxicity via the generation of reactive oxygen species. *NeuroToxicology*. 2023;95:155-163.
31. Zhang F, You X, Zhu T, Gao S, Wang Y, Wang R, et al. Silica nanoparticles enhance germ cell apoptosis by inducing Reactive Oxygen Species (ROS) formation in *Caenorhabditis elegans*. *J Toxicol Sci*. 2020;45:117-129.
32. Wang Y, Gai T, Zhang L, Chen L, Wang S, Ye T, et al. Neurotoxicity of bisphenol A exposure on *Caenorhabditis elegans* induced by disturbance of neurotransmitter and oxidative damage. *Ecotoxicol environ saf*. 2023;252:114617.
33. Qu Z, Liu L, Wu X, Guo P, Yu Z, Wang P, et al. Cadmium-induced reproductive toxicity combined with a correlation to the oogenesis process and competing endogenous RNA networks based on a *Caenorhabditis elegans* model. *Ecotoxicol environ saf*. 2023;268:115687.
34. Nicolai MM, Weishaupt AK, Baesler J, Brinkmann V, Wellenberg A, Winkelbeiner N, et al. Effects of manganese on genomic integrity in the multicellular model organism *Caenorhabditis elegans*. *Int J Mol Sci*. 2021;22:10905.
35. Ke T, Santamaria A, Junior FB, Rocha JB, Bowman AB, Aschner M, et al. Methylmercury exposure-induced reproductive effects are mediated by dopamine in *Caenorhabditis elegans*. *Neurotoxicol Teratol*. 2022;93:107120.
36. Zhang J, Yang W, Li Z, Huang F, Zhang K. Multigenerational exposure of cadmium trans-generationally impairs locomotive and chemotactic behaviors in *Caenorhabditis elegans*. *Chemosphere*. 2023;325:138432.
37. Zhenglu Wang, S. D., Jinze Wang, Wei Du, Lin Zhu. Assessment on chronic and transgenerational toxicity of methamphetamine to *Caenorhabditis elegans* and associated aquatic risk through Toxicity Indicator *Environ Pollut*. 2021;288:117696.
38. Qian Lu, Y. B., Lingyi Ma, Ran Liu. Transgenerational reproductive and developmental toxicity of tebuconazole in *Caenorhabditis elegans*. *J Appl Toxicol*. 2020;40:578-591.
39. Zeng XS, Geng WS, Jia JJ. Neurotoxin-induced animal models of Parkinson disease: Pathogenic mechanism and assessment. *ASN Neuro*. 2018;10:1759091418777438.
40. da Silva LP, da Cruz Guedes E, Fernandes IC, Pedroza LA, da Silva Pereira GJ, Gubert P, et al. Exploring *caenorhabditis elegans* as Parkinson's disease model: Neurotoxins and genetic implications. *Neurotox Res*. 2024;42:11.
41. Engleman EA, Katner SN, Neal-Beliveau BS. *Caenorhabditis elegans* as a model to study the molecular and genetic mechanisms of drug addiction. *Progress in molecular biology and translational science*. 2016;137:229-252.
42. Salim C, Kan AK, Batsaikhan E, Patterson EC, Jee C. Neuropeptidergic regulation of compulsive ethanol seeking in *C. elegans*. *Sci Rep*. 2022;12:1804.
43. Yang YF, Chen PJ, Liao VH. Nanoscale zerovalent iron (nZVI) at environmentally relevant concentrations induced multigenerational reproductive toxicity in *Caenorhabditis elegans*. *Chemosphere*. 2016;150:615-623.
44. Hu C, Hou J, Zhu Y, Lin D. Multigenerational exposure to TiO<sub>2</sub> nanoparticles in soil stimulates stress resistance and longevity of survived *C. elegans* via activating insulin/IGF-like signaling. *Environ Pollut*. 2020;263:114376.