

Antibiotic Susceptibility Patterns of Salmonella Sp. Isolated From Cattle Dungs and Slaughter Slabs, Ile-Ife, Osun State

J. Omololu-Aso^{1*}, D. Adekunle¹, O. O. Omololu-Aso², O. Adesunloro Azeez K¹

1Department of Microbiology, Obafemi Awolowo University, Osun, Nigeria; ²Department of Obstetrics Gyneacology, University Orita-mefa Ibadan, Nigeria

ABSTRACT

Introduction: Bacteria of the genus Salmonella are significant food safety concern in many parts of the world, especially in developing countries like Nigeria.

Materials and methods: A total of Sixty-seven (67) samples were obtained from different abattoirs in all the four local government Areas in Ile-Ife. Samples collected from the different abattoirs include; slaughter slab and swab of freshly killed cow dung, with the aim of isolating and determining Salmonella species. The samples were transported into the microbiological laboratory and isolated according to conventional, cultural and serological methods, alongside further biochemical test was also carried out.

Result: Out of the 67 samples, 21 were positive for Salmonella. The antibiotic susceptibility pattern of theisolates was subjected to 7 routinely prescribed antibiotics namely; chloramphenicol, ciprofloxacin, amoxicillin, Augmentin, gentamycin, pefloxacin, streptomycin.

Conclusion: This study confirms the presence of multi-drug resistant Salmonella species in cattle dung and slaughter slab at the abattoirs in Ile-Ife, Osun state Nigeria. Proper hygiene methods should be encouraged in the study area. To prevent cross infection between the meat vendors and the consumers.

Keywords: Salmonella; Infections; Antibiotic resistant; Antibiotics; Abattoir

INTRODUCTION

The demand and consumption of animal products such as meat (especially raw meat) is high in particular and in the country in general. Nevertheless, reports on the hygienic status abattoir and butcher shops are fragmented because no comparable data are available regarding the assessment of food safety practice, foodborne diseases, and microbial load of meat in the abattoir and butcher shops of the study area. Top among hygiene slaughter challenges in developing countries, including Nigeria, is the lack of necessary infrastructure that enhances hygiene processing such as potable water supply, drainage, and proper waste disposal facilities [1-3].

This is exacerbated by poor knowledge, unhygienic methods of slaughter and processing activities among slaughterhouse personnel as well as government's lack of political will to enforce

hygiene slaughter and meat inspection laws in slaughterhouses. In most Nigeria slaughterhouses, activities such as slaughter, flaying, carcass splitting, and intestinal evisceration are usually performed on the floor resulting in profound contamination of the floor and carcasses with feces. Livestock feces are heavily populated with Enterobacteriaceae, which are commensal flora of the gastrointestinal tract.

They are always present in the slaughterhouse environment, and being harbored and shed by animals and slaughterhouse personnel, are the most common contaminating aerobic bacteria in slaughterhouses. Salmonella is a zoonotic foodborne pathogen and the etiologic agent of salmonellosis. The emergence of antimicrobial resistant Salmonella recovered from meat products has become a source of major concern. Pathogens are reported to inflict more serious damage when they are antimicrobialresistant. Resistance results in longer stay in hospital as well as

Correspondence to: Joseph Omololu-Aso, Department of Microbiology, Obafemi Awolowo University, Osun, Nigeria; E-mail: omololuaso@oauife.edu.ng

Received: 27-Sep-2022, Manuscript No. JADPR-22-19373; Editor assigned: 29-Sep-2022, PreQC No. JADPR-22-19373 (PQ); Reviewed: 10-Oct-2022, QC No. JADPR-22-19373; Revised: 01-Feb-2023, Manuscript No. JADPR-22-19373 (R); Published: 09-Feb-2023, DOI: 10.35248/2329-8731.23.11.291

Citation: Aso JO, Adekunle D, Aso OOO, Azeez OK (2023) Antibiotic Susceptibility Patterns of Salmonella Sp. Isolated From Cattle Dungs and Slaughter Slabs, Ile-Ife, Osun State. J Infect Dis Preve Med. 11:291.

Copyright: © 2023 Aso IO, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. J Infect Dis Preve Med, Vol.11 Iss.2 No:1000291

increased cost and less effective chemotherapy that may become toxic to the patient. Infections due to resistant Salmonella are reported to result in increase in morbidity and mortality, especially in immune compromised patients.

Obviously, the presence of resistant *Salmonella* in meat is a threat to public health. Salmonellae resist drugs through enzymatic degradation, alteration of site of actions, blocking cell permeability, efflux pumps, and horizontal transfer of resistant genes. Drug-resistant *Salmonella* are capable of transmitting antibacterial resistance genes to other bacteria, thereby maintaining the dissemination of antimicrobial resistance among bacteria in the food chain. In Nigeria, multidrugresistant *Salmonella* spp. has been isolated from various sources: Water, farm produce and environment, abattoir environment poultry [4-6].

In this study, we investigated the antibiotic susceptibility patterns of *Salmonella* sp. isolated from cattle dungs and slaughter slabs, ile-Ife, osun state.

MATERIALS AND METHODS

Study area

The study was carried out in slaughter houses located in Ile-ife, Osun state. The city is located in present-day Osun state with a population of 509,813 according to population census of 2006 (Figure 1).

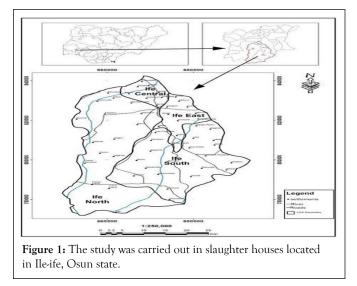


Table 1: Morphological reaction of isolate to the selected media.

Morphology	Results				
Growth in nutrient agar	Positive for all isolates				
Growth in macconkey agar	Positive for Salmonella species				
Growth in Salmonella-shigella agar	Positive for Salmonella species				
Motility testing	Positive for Salmonella species				
Gram staining	Negative for Salmonella species				

In order to get the cattle slaughter house, 15 locations were selected randomly out of (29 wards local government) located in Ile-ife.

Sample collection

The surface swabs from abattoir were collected aseptically using sterile moistened swab sticks by rubbing firmly over the predetermined surface area using parallel stroke lines with slow rotation with respectively chosen template surface area to be swabbed. In the abattoir the moistened sterile swab stick was used to swab 60 cm² from the slaughter slabs. The swabs were transferred to the respective capped sterile tubes and then transferred into the tube containing nutrient broth aseptically.

Isolation, identification and characterization

Each sample swab was inoculated into prepared sterile nutrient broth (Oxoid, UK) and incubated at 37°C for 24 h for enrichment. Discrete colonies were later sub-cultured on a solid differential media; MacKonkey agar (*E.coli*) and *salmonella*shigella agar (SS Agar). All the plates were incubated at 37°C for 24 hr. Discrete colonies were further sub cultured onto freshly prepared plates of nutrient agar. Gram staining, morphological identification, catalase and coagulase tests, sugar fermentation analysis and antimicrobial susceptibility trends of the isolate were conducted [7-10].

RESULTS

A total of twenty-one (21) *Salmonella* isolates were isolated from 67 samples obtained from cattle dung and slaughter slab (Tables 1-4).

 Table 2: Microscopic examination of isolates.

Isolates	Gram reaction	Shape	Arrangement
Mc22	-ve	Rod	Clustered
Kc4	-ve	Rod	Clustered
Ac4	-ve	Rod	Clustered
Ac2	-ve	Rod	Clustered
Hc2	-ve	Rod	Clustered
Hc4	-ve	Rod	Clustered
Gc4	-ve	Rod	Clustered
Ic2	-ve	Rod	Clustered
Ic4	-ve	Rod	Clustered
Jc2	-ve	Rod	Clustered
Hb	ve	Rod	Clustered
Fb2	-ve	Rod	Clustered
Fb3	-ve	Rod	Clustered
Gb1	-ve	Rod	Separated
Ga	-ve	Rod	Clustered
Ia1	-ve	Rod	Clustered
Ia2	-ve	Rod	Clustered
Ma	-ve	Rod	Clustered
Fa	-ve	Rod	Clustered
Ea2	-ve	Rod	Separated
Ea1	-ve	Rod	Clustered

 Table 3: Biochemical test of the Salmonella specie.

Sample Code	Biochemical test									
	Catalase	Indole	Citrate	Motility	Motility Methyl red	Gram staining	Triple sugar iron			
					·		glucose	Lactose	H_2S	Gas
Kc4	+	-	+	+	+	-	+	-	+	
Mc22	+	-	+	+	+	-	+	-	+	+
Ac4	+	-	+	+	+	-	+	-	-	-
Ac2	+	-	+	+	+	-	+	+	+	-
Hc2	+	-	+	+	+	-	+	-	+	-
Hc4	+	-	+	+	+	-	+	-	-	-

Gc4	+	-	+	+	+	-	+	-	+	-	
Ic2	+	-	+	+	+	-	+	-	+	-	
Ic4	+	-	+	+	+	-	+	-	+	-	
Jc2	+	-	+	+	+	-	+	-	+	-	
Hb	+	-	+	+	+	-	+	-	+	-	
Fb2	+	-	+	+	+	-	+	-	+	-	
Fb3	+	-	+	+	+	-	+	-	+	-	
Gb1	+	-	+	+	+	-	+	-	+	-	
Ga	+	-	+	+	+	-	+	-	-	-	
Ia1	+	-	+	+	+	-	+	-	+	-	
Ia2	+	-	+	+	+	-	+	-	+	+	
Ma	+	-	+	+	+	-	+	-	-	+	
Fa	+	-	+	+	+	-	+	-	+	+	
Ea2	+	-	+	+	+	-	+	-	+	-	
Ea1	+	-	+	+	+	-	+	-	+	-	

Table 4: Antibiotic susceptibility index of Salmonella.

Antibiotics	Total isolates	Susceptible n (%)	Intermediate n (%)	Resistant n (%)	Total n (%)
Chloramphenicol	15	14 (93.33)	1 (6.67)	0 (0)	15 (100)
Ciprofloxacin	15	15 (100)	0 (0)	0 (0)	15 (100)
Amoxicillin	15	10 (66.67)	1 (6.67)	4 (26.67)	15 (100)
Augmentin	15	9 (60)	2 (13.33)	4 (26.67)	15 (100)
Gentamycin	15	2 (13.33)	0 (0)	13 (86.67)	15 (100)
Pefloxacin	15	15 (100	0 (0)	0 (0)	15 (100)
Streptomycin	15	15 (100)	0 (0)	0 (0)	15 (100)

DISCUSSION

The study revealed the antibiotic susceptibility pattern of *Salmonella* specie isolated from slaughter slabs and cattle dungs in Ile-Ife, Osun state and how it poses a significant risk on the consumers. From the 67 samples collected from abattoirs 21 samples showed *Salmonella* growth and the rest showed other enteric bacteria such as *Escherichia coli* and *Shigella*. There is thus,

urgent need for applying proper hygienic practices among the food vendors as we equally recommend the use disposable aprons by the meat vendors as these will restrict transfer and the spread of food borne pathogens (Table 5) [11-14].

Antibiotics	Disc potency	Susceptible	intermediate	Resistance
Chloramphenicol	30 µg	≥ 18	13-17	≤12
Ciprofloxacin	10 µg	2 31	21-30	≤20
Amoxicillin	30 µg	≥ 18	14-17	≤13
Augmentin	30 µg	≥ 18	14-17	≤13
Gentamycin	10 µg	≤ 15	13-14	212
Pefloxacin	30 µg	≥15	12-14	≤11
Streptomycin	30 µg	≥15	12-14	≤11

Based on this study *Salmonella* showed resistance to Gentamycin, Amoxicillin and Augmentin but susceptible to Ciprofloxacin, Chloramphenicol, Pefloxacin and Streptomycin. The ability of some of these *Salmonella* isolates to show high level of resistance to some of the antimicrobial agents used is an indication that these antibiotics have been abused, hence the possibility they have acquired resistance.

CONCLUSION

Knowing the prevalent resistance of these microorganisms to antibiotics, different antibiotics must be used and tested to know which actually is potent to treat salmonella infections so as to reduce excessive cost of health services, reduce morbid services, reduce mortality rate and overall lead to a general wellbeing. However, preventive measures should be taken especially daily washing of the slaughter slab and proper disposal of feacal matters, availability of clean drinking water, promotion of safe food handling practices and public health education in order to maintain a proper hygiene level. The knowledge and understanding about factors that contribute to the occurrence, distribution and establishment of the salmonella disease may be helpful in eliminating its survival in the environment, thereby protecting public health.

REFERENCES

- Abakpa GO, Umoh VJ, Ameh JB, Yakubu SE, Kwaga JKP, Kamaruzaman S. Diversity and antimicrobial resistance of Salmonella enterica isolated from fresh produce and environmental samples. Environ Nanotechnol Monit Manag. 2015;3:38-46.
- Adesiyun AA, Oni OO. Prevalence and antibiograms of salmonellae in slaughter cattle, slaughter areas and effluents in Zaria Abattoir. J Food Prot. 1989;52(4):232-235.
- Akinyemi KO, Iwalokun BA, Foli F, Oshodi K, Coker AO. Prevalence of multiple drug resistance and screening of enterotoxin (stn) gene in *Salmonella* enterica serovars from water sources in Lagos, Nigeria. Public Health. 2010;125(2):65-71.

- Babatunde SK, Kolawole DO, Adedayo MR, Ajiboye AE, Ajao AT, Mustapha ON. Prevalence and characterization of Salmonella isolates from poultry farms in Ilorin. Niger J Life Sci. 2017;4(1): 1-4.
- Botteldoorn N, Heyndrickx M, Rijpens N, Grijspeerdt K, Herman L. Salmonella on pig carcasses: Positive pigs and cross contamination in the slaughterhouse. J Appl Microbiol. 2003;95(5):891-903.
- Cosby de, Cox NA, Harrison MA, Wilson JL, Buhr RJ, Fedorka-Cray PJ. Salmonella and antimicrobial resistance in broilers: A review. J Appl Poult Res. 2015;24(3):408-426.
- Cosgrove SE, Sakoulas G, Perencevich EN, Schwaber MJ, Karchmer AW, Carmeli Y. Comparison of mortality associated with methicillin-resistant and methicillin- susceptible *Staphylococcus aureus* bacteremia: A meta-analysis. Clin Infect Dis. 2003;6(1):53-59.
- Dammo DJM, Sangodoyin MNW. Impacts of effluents on river Nggada water quality in Maiduguri, Nigeria. Adv Sci Eng Res. 2017;3(1):1-9.
- El-Tayeb MA, Ibrahim ASS, Al-Salamah AA, Almaary KS, Elbadawi YB. Prevalence, serotyping and antimicrobials resistance mechanism of Salmonella enterica isolated from clinical and environmental samples in Saudi Arabia. Braz J Microbiol. 2017;48(3):499-508.
- Fasanmi GO, Olukole SG, Kehinde OO. Microbial studies of table scrapings from meat stalls in Ibadan metropolis, Nigeria: Implications on meat hygiene. Afr J Biotech. 2010;9(21):3158-3162.
- Igbinosa EO, Beshiru A. Isolation and characterization of antibiotic susceptibility profile of *Salmonella* species isolated from abattoir environment. J Sci. 2017;19(2):389-397.
- 12. Iheanacho O, Delia G, Hussni M, Dipeolu M, Poole J, Gachohi J, Makita K. Assessment of risks to human health associated with meat from different value chains in Nigeria: Using the example of the beef value chain. Report for Nigeria Intergrated Animal and Human Health Management Project. 2012;1-111.
- Istifanus V, Bwala HB. Infrastructure challenges: The review of environmental and health implication of abattoir operation in a developing country. Global J human-social Sci Res. 2017;5(6):60-72.
- Quinn PJ, Markey BK. Concise review of veterinary microbiology. 2nd ed. Blackwell Publishing Limited, Oxford. 2003;1-50.