

Adult NG2-Expressing Cells in Multiple Organs: A Novel Progenitor in Regenerative Medicine

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Abstract

Stem/progenitor cells have emerged as a promising tool for studying the mechanisms of cell development, tissue regeneration, and cell therapy for various disorders. Stem/progenitor cells are found not only in the embryo but also in most adult tissues for endogenous repair. They are capable of self renewal and differentiation into various cell types from their germ line, thus, are ideal candidates for cell-based therapy. Expression of neuron-glial antigen 2 (NG2) proteoglycan is found on several types of cell surface, majorly distributing on undifferentiated precursor cells in the Central Nervous System (CNS), named Oligodendrocyte Progenitor Cells (OPCs). NG2 proteoglycan has a widespread range of physiological roles and the cells that express chondroitin sulphate proteoglycan NG2/CSPG4 (NG2⁺cells) react to all forms of pathological insults. This cell population is abundant in the developing and mature organs with also pericytes PCs and mesenchymal stem cell (MSCs) potential. Our pilot studies demonstrated that NG2⁺cells are dedicated stem/precursors not only in the CNS as traditional thought, but also outside the CNS. Following injury, the repertoire of NG2⁺cells expands to become functional cells. Our experiment also showed the importance of NG2⁺cells as the reservoir for MSCs, which reaffirms the central role of these cells for their therapeutic potential. In this review, we provided some pilot evidence and briefly summarized the more recent progress on adult NG2⁺cell researches. We hypothesize that adult NG2⁺cells can be generated from not only the adult mammalian's CNS but also multiple outside organs including spinal cord, bone marrow, eyes, liver, heart, lung, pancreas and kidney. Therefore, development and research of adult NG2⁺cells may open a novel perspective in regenerative medicine in the future.

Keywords: NG2-expressing cells; Self-renewal; Tissue repair; Pericytes; Mesenchymal stem cells

Abbreviations: BM:MSCs: Bone Marrow: Derived Mesenchymal Stem Cells; BrdU: Bromodeoxyuridine; CNS: the Central Nervous System; EC: Endothelial cells; ECM: Extracellular Matrix; ERK1/2: The Extracellular Signal: Regulated Kinase 1/2; GABA_A: Gamma-Aminobutyric Acid-A; GFAP: Glial Fibrillary Acidic Protein; GluRs: Glutamate Receptors; GR: Glutamate Receptors; GS: Glutamine Synthetase; HSCs: Hepatic Stellate Cells; IC: Ischemic Cardiomyopathy; MAPK: Mitogen Associated Phosphate Kinases; MI: Myocardial Infarction; MS: Multiple Sclerosis; OPCs: Oligodendrocyte Precursor Cells; PC: Pericytes; PDGFR- β : Platelet-derived Growth Factor Receptor-Beta; α -SMAs: Alpha Smooth Muscle Actin; SMCs: Smooth Muscle Cells; Tkase: Tyrosine Kinase; Vwf: Von Willebrand Factor

NG2-Expressing Cells are Progenitors in the Adult Central Nervous System

The story of NG2 proteoglycan being as a valuable marker for identification of oligodendrocyte precursor cells (OPCs) in the Central Nervous System (CNS) began nearly thirty years ago [1]. Since then, scientists have postulated that NG2 proteoglycan expression might be the characteristic of immature neural cells capable of differentiating into either glia or neurons, hence the designation of the molecule as "nerve/glial antigen 2" [2]. However, the function of NG2-expressing cells (NG2⁺cells) in the adult CNS is still uncertain although that the NG2⁺cells are progenitor cells in the developing CNS is well documented [3]. In addition, the contribution of functional receptors of NG2⁺cells on neurons in response variety of injuries indicates NG2⁺cells plays a significant role in stimulating neural network as well as their potential importance in inflammation, neurodegeneration and regenerative medicine [4].

Phenotypic and physiological roles of NG2⁺cells in the adult CNS

Cells expressing the chondroitin sulphate proteoglycan NG2/CSPG4 are glia lineage which makes up 5-10% of all glia in the adult CNS. The morphology and distribution of NG2 proteoglycan are similar to, but distinct from, both microglia and astrocytes. The NG2⁺cells are present early in development and persist ubiquitously throughout CNS parenchyma where they exhibit multi-branched processes, big cell bodies and small nuclear. NG2 proteoglycan is a type 1-transmembrane protein expressed by a range of cell types within the mammalian nervous system. They are found in grey and white matter tracts of the developing and adult CNS. In vitro NG2⁺cells possess some functional receptors and contact neurons at the nodes of Ranvier or via synaptic terminals and some of them even have fire action potential. There has recently been much debate concerning the possible heterogeneous nature of the NG2⁺cells with respect to developmental

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origin and functional roles. The possibility of multiple and varied functions of NG2⁺ cells also suggest that such cell population may be heterogeneous [5]. Several studies have proposed that there may be two distinct populations of NG2⁺ cells in the developmental CNS, one is able to generate mature oligodendrocytes whereas the other has undefined roles [6-8]. For example, researchers have examined the response of NG2⁺ cells following demyelination in the rat spinal cord and identified the presence of two NG2⁺ cells populations, one responding to the insult by dividing and the other non-responsive, suggesting that the responsive population acts as oligodendrocyte progenitors [9]. However, whether the NG2⁺ cells residing in the adult CNS resemble to the ones in embryonic or neonatal CNS in terms of their morphology or proliferation characteristics remains unsolved [10].

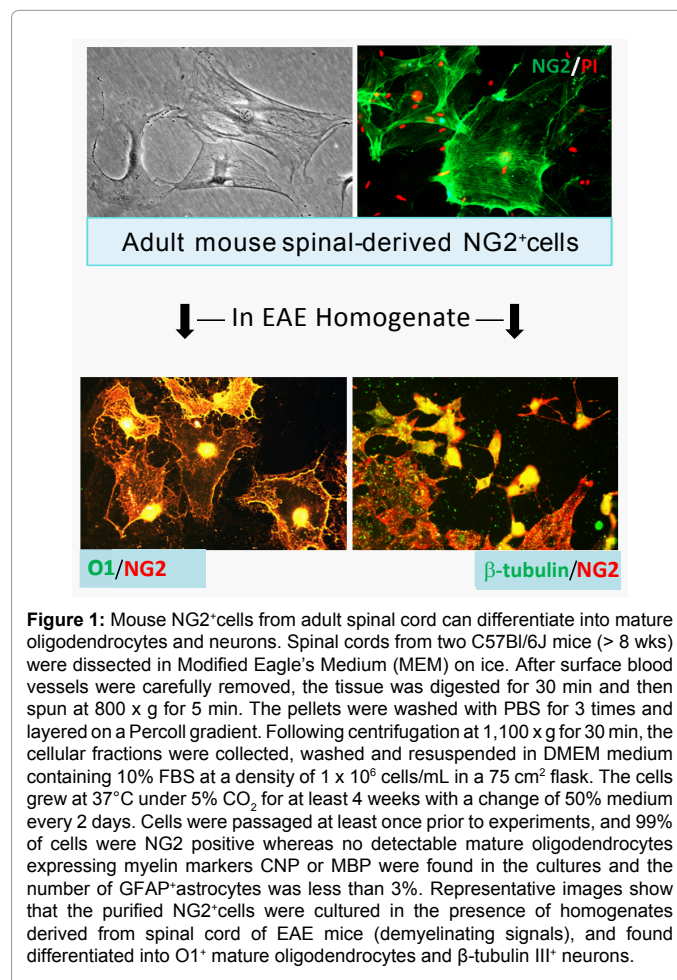
Defining the role of adult NG2⁺ cells *in vivo* CNS has remained elusive despite their phenotypic characterization as OPCs and are able to give rise to oligodendrocytes *in vitro* [11,12]. NG2 molecule possesses a large extracellular domain (300 kDa core protein) and was initially identified by a sequentially-absorbed rabbit antiserum [13,14]. The big molecule of NG2 includes sites near the single transmembrane domain which are readily cleaved by a variety of proteases [15], resulting in the deposition of the ectodomain in the extracellular matrix (ECM). Although having relatively slow rate of cell division adult NG2⁺ cells represent the major proliferative population in the adult CNS, accounting for 70% and 74% of bromodeoxyuridine (BrdU)-incorporating cells in the spinal cord and cerebral cortex respectively [16,17]. Generation of newly oligodendrocytes from adult CNS is an expected function, but it is likely that they also participate in homeostasis, glutamate signaling and interdigitate between pre- and post-synaptic terminals [18,19]. Experiments in some laboratories identified a serological epitope on human melanoma cells known as HMW-MAA [20-22] as the human homolog of NG2 proteoglycan [23]. Subsequently, detected excitatory synapses mediated by AMPA receptors on NG2⁺ cells and found respond to neuron released glutamate of such cells [18], indicating the expression of glutamate receptors (GR) on NG2⁺ cells. Furthermore, the presence of GR on NG2⁺ cells in the adult CNS regions has been recognized and reported as functional gamma-aminobutyric acid-A (GABA-A) receptors [24]. This is more likely to act as a rapid means of altering ion concentrations within NG2⁺ cells [25,26], thereby altering their physiological properties. In keeping with a role in glutamate neurotransmission suggesting oligodendrocytes and NG2⁺ cells are both to express glutamine synthetase [16,17]. Thus, there is good evidence for a role for NG2⁺ cells in glutamine cycle.

Stem/progenitor potential of NG2 cells for neurological disorders

NG2 proteoglycan can be detected in a variety of CNS diseases including demyelinating diseases of which multiple sclerosis (MS) is the best studied. Understanding the mechanisms underlying remyelination is essential to elucidate the reasons for the development of chronic demyelinated plaques seen in MS. In experimental models of demyelination, remodeling of NG2⁺ cells for denuded axons is usually an extremely rapid and efficient repair process, regardless of the method of induction of demyelination [27,28], suggesting NG2⁺ cells are largely responsible for myelin repair [29]. It has been demonstrated that cell proliferation is required in order to replenish lost oligodendrocytes [30] and the expression of Ki-67 antigen adult NG2⁺ cells in the presence of a toxicity indicating proliferation and undergo morphological changes of this cell population [15]. In human MS tissues, the antigenic profile and morphology of NG2⁺ cells are consistent with an oligodendrocyte progenitor function. By using

retroviral labeling as tracking tool demonstrated that NG2⁺ cells are the major cycling cell in the adult CNS not just least the progenitors of oligodendrocytes when cultured them *in vitro* [16,17].

Adult NG2⁺ cells will undergo well-characterized reactive changes in response to CNS pathology. Adult NG2⁺ cells has been tested by various CNS injury models and found that grafted cells adjacent to the damage site where increase in number and become hypertrophic [7,31,32]. A study reported that NG2⁺ cells generate mature oligodendrocytes, in transgenic mice neurons alone with subpopulations of astrocytes during normal development, indicating their stem/progenitor potential. Therefore, a new method is to be required for the development of NG2⁺ cell isolation or purification. Based on this line, we purified NG2⁺ cells for the first time from adult mouse spinal cord by using a novel procedure (Figure 1) and found that this purified NG2⁺ cells are highly homogenous (more than 98% of cells express NG2 proteoglycan). They display discoid, heterochromatic, irregular shape with multiple processes and importantly to response to injury. For instance, these transplanted cells could move into diseased conditions, protecting neural cell survival as well as increase remyelination in a mouse model of demyelinating disease such as in Experimental Allergic Encephalomyelitis (EAE) [33]. In addition, our study provided strong evidence that NG2⁺ cells are the highest dividing cell population comparing to other glial cell types when their receiving damage signals. Such phenomena were consistent with a study from Lee et al. laboratory. They have reported that injection intravenously of



NG2⁺cells resulted in grafted cells migrated into the lesion areas where they attenuated the lesion atrophy and induced a long-term functional improvement in an animal model [34]. Since differentiation potential is thought to be influenced by NG2⁺cell environment *in vitro* [35], it is possible NG2⁺cell may display a broader lineage potential following CNS disease [13,36]. They may participate in tissue repair in response to injury and promote regeneration in demyelinating insult by generating further differentiated functional oligodendrocytes and neurons. More importantly, these purified adult CNS-derived NG2⁺cells (CNS-NG2⁺cells) could be also applied in transplantation therapies for treating diverse other organ disorders such as eye diseases.

Are NG2-Expressing Cells Progenitors in the Adult Optic Nerve?

The optic nerve is a CNS white matter tract that connects eye to brain. It contains glia, astrocytes and axon of retinal ganglion cells. Among glia, the major populations are oligodendrocytes, astrocytes and microglia as well as NG2⁺cells.

Physiological and pathological functions of NG2⁺cells in adult optic nerve

In the developmental optic nerve, NG2⁺cells are highly identified in grey and white matter [37]. NG2⁺cells are a small branch originated in the row of oligodendrocytes that make connection with the nodes of Ranvier in white matter and function (synapse) in grey matter [18]. In the adult optic nerve, only 7% of NG2⁺cells exist glial cells and their composition are only limited within optic nerve head in demyelinating lesion. NG2⁺cells share some similarity with astrocytes in morphology and biological characteristics [18] but do not express astrocyte markers like GFAP, vimentin, calcium-binding protein S-100b and glutamine synthetase (GS) [38].

It is well known that the cells expressing NG2 in the CNS were considered as OPCs [10], because of their being able to develop into oligodendrocytes or oligodendrocytes-type-2 astrocytes (O-2A) progenitors *in vitro* [39]. It is demonstrated that the stage of generating newly oligodendrocytes from NG2⁺cells in developmental CNS [40-42] would express transition marker, a tetraspanin protein CD9 [43], while CD9 was lost on NG2⁺cells due to remarkable decreased proliferative rate in the optic nerve [41,44,45]. However, much work has been done for the differentiation of oligodendrocytes from NG2⁺OPC [46,47], but the problem of whether the newly NG2⁺OPCs differentiated oligodendrocytes have function remains opened.

NG2⁺cells in the adult optic nerve are regenerative stem/progenitors

The presence of NG2⁺cells in the optic nerve may have implications for efforts to enhance endogenous repair in a wide range of disorders [27]. However, the factors that regulate the diverse responses of NG2⁺cells in optic nerve to damage are elusive. Several lines of evidence indicate a potential role of NG2⁺cells in optic nerve for the neurotransmitter glutamate [48]. Firstly, NG2⁺cells express AMPA-type glutamate receptors (GluRs) [18,49] by axons in white matter [50,51]. Secondly, there is evidence that AMPA receptors on NG2⁺cells are permeable to Ca²⁺ [51-53], and this study provided a mechanism by which glutamate could regulate intracellular pathways that control cell proliferation, growth, differentiation and death, such as the calcium-dependent mitogen-associated phosphate kinases (MAPK), extracellular signal-regulated kinases (ERK1/2). Finally, glutamate has a well-defined role in the pathology of oligodendrocyte lineage cells

[54]. Therefore, NG2⁺cells like NG2⁺OPCs and oligodendrocytes are extremely susceptible to glutamate-mediated calcium-dependent excitotoxicity [49,55]. Some experimental results showed that injection of the potent glutamate receptor agonist (kainite) into the cerebral cortex and white matter of the optic nerve induced an extensive gliosis [19,56] and caused the loss of oligodendrocytes demyelinating injury [56,57].

It is very important to address whether existed NG2⁺cells in optic nerve have comparable biological feature with CNS-NG2⁺cells, we used the same procedure as adult CNS-NG2⁺cells and purified successfully NG2⁺cells from adult mouse eyes (Iris) (Iris-NG2⁺cells). Our experimental results showed that NG2⁺cells have similar morphology and biological characteristics with CNS-NG2⁺cells. They exhibit a more polarized appearance, radial morphology, extending processes and fine multi-branching processes and lack expression of GFAP [43]. Iris-NG2⁺cells involved in a number of crucial cellular processes. When grown them in the injured conditioned media (retinal homogenate) collected from retinal pigment mutation mice with tyrosine kinase (TKase) mutation (lack TKase function), the purified Iris-NG2⁺cells differentiated into Opsin⁺ functional cone neurons (Figure 2).

Are NG2-Expressing Cells Progenitors in the Adult Bone Marrow?

The contribution of NG2⁺cells to tissue repair in brain and eye disorders has been highlighted in recent studies, but far from being restricted NG2⁺cells as progenitors is OPC marker. It is also an effective progenitor marker in bone marrow (BM) [58] existing as pericytes (PCs) or mesenchymal stem cells (MSCs) [59,60]. We propose the existence of these “adult NG2 progenitors” in BM may immediately recognize to have important implications for tissue repair during

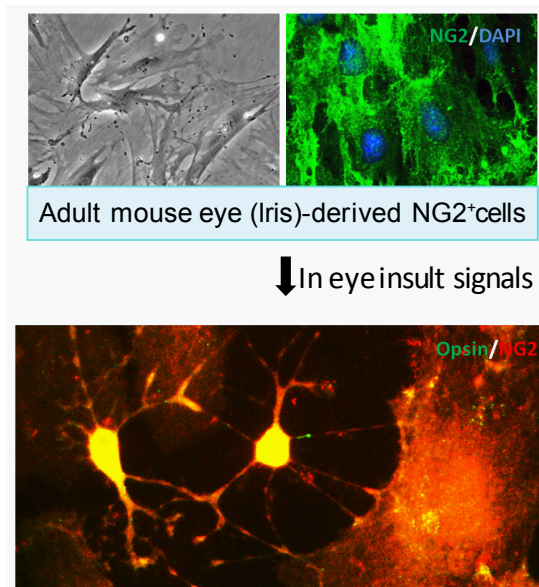


Figure 2: Mouse eye (Iris)-derived NG2-expressing progenitors (eye (Iris)-NG2⁺cells) differentiate into functional Opsin⁺core neuron in response to injured eye homogenates. Mouse eye Iris from two C57Bl/6J mice (> 8 wks) was used to obtain NG2⁺cells. The rest of procedures to isolate NG2⁺cells were performed as described above. NG2⁺cells were cultured in the presence of homogenates derived from eye injured mice. Representative images show that the isolated NG2⁺cells differentiated into functional Opsin⁺core neuron when cultured in the presence of homogenates derived from Iris mice (eye insults).

diseases of diverse tissues. Hopefully, an increase in the pace of our acquisition of knowledge about the stem/progenitor functional role played by adult NG2⁺ cells in both normal and pathological tissues for regenerative medicine will be seen in the coming years.

Are NG2⁺ Cells the reservoir for mesenchymal stem cells?

Much of the work conducted on adult stem/progenitor cells has focused on mesenchymal stem cells (MSCs) which is a leading candidate stem cell population for clinical applications. Adult MSCs can be isolated from a range of stroma tissues including bone marrow (BM-MSCs) and fat [61]. One capacity of MSCs that is well established is their ability to release a wide range of cellular modulators that can modulate the host tissue [62] and immune responses [63-65]. The capacity of BM-MSCs modulate immune response combined with the release of a range of trophic factors contributing injured tissue regeneration, thus MSCs have being brought into a broader therapeutic scenario. Recent studies have demonstrated that MSCs can be generated from NG2-expressing PCs within injured area and create a regenerative environment by releasing bioactive trophic factor. For example expression of NG2, PDGF receptor beta (PDGFR-β) and α-smooth muscle actin (α-SMA) [66], strongly supporting MG2-expressing MSCs possess PC potential. The term “MSCs” was first used to describe as adherent, marrow-derived homogeneous cells. They can divide in *ex vivo* and generated a diversity of connective cells [67]. Although a body of studies verified that BM-MSCs can differentiate along mesodermal lineages to form bone muscle and cartilage [68] the mechanism underlying their differentiation is not clear. There are detailed and elegant studies in the literature to support the fact that MSCs can be observed for almost every perivascular locations (on both arterial and venous vessels) and PC antigens can be identified by anti-NG2 and anti CD146 antibodies [69,70].

More work has been done about BM-derived NG2⁺ cells but still no conclusion can be drawn regarding the real foothold of PCs and MSCs, and also the relationship between NG2-expressing PCs and MSCs remains to be established, there are several open questions: (1) Can NG2-expressing MSCs be named PCs and how to picture NG2-expressing MSC progenitor potential? (2) If NG2⁺ cells were truly BM-MSC progenitors, can NG2-expressing MSCs be functional as vascular PCs? (3) Do the multiple organ-derived NG2⁺ cells share the same biological characteristics? (4) are NG2-expressing PCs from bone marrow ‘better’ than the ones from other organs?

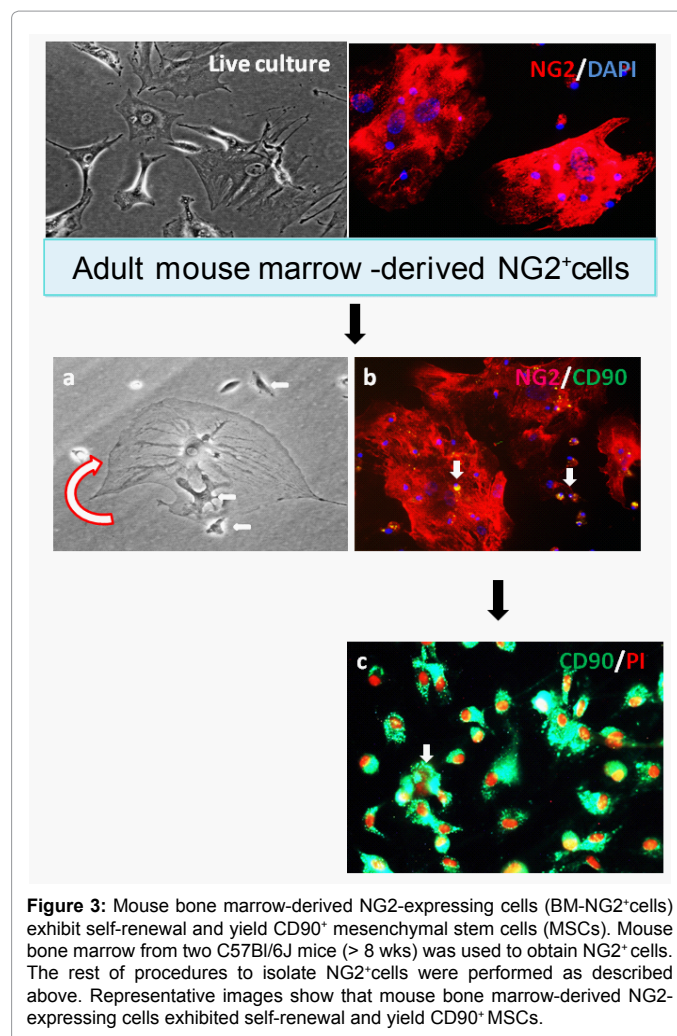
The challenge of translating bench studies on BM-NG2⁺ MSC therapies to the bed application

The last decade has seen a remarkable advance of BM-MSCs therapeutic benefit from studies on animal models and pre-clinical investigations. The disorders and conditions appeared to benefit from infused BM-MSCs result from two generalizable therapeutic activities, immunomodulation and trophic activities [71]. Immunomodulation involves dendritic cells, B- and T-cells [72-75], while trophic activities involve MSC-released bioactive factors that shift the inflammatory Th cells from Th1 to Th2 [76], prohibit inflammatory cytokine production, scar formation [77-79], and angiogenesis [80], resulted in tissue repair by stimulating intrinsic progenitor cell activation, proliferation and appropriately differentiate [62,81].

A major challenge in moving MSC therapies into the clinic is to understand the mechanism that MSC existed. In this regard, encouraging our current researches came from the study on animal models of MS and demonstrated that human BM-MSC-secreted

hepatocyte growth factor (HGF) is a critical molecule that contributed to immunomodulatory and direct regeneration. Treatment of mice with EAE and lysolecithin (LPC)-induced demyelination with human MSC-HGF contained conditioned media (MSC-HGF-CM) resulted in a dramatic reduction in functional deficits, improved histological appearance and alteration in the relative levels of pro and anti-inflammatory cytokines [82]. However, current knowledge regarding the immunobiology and clinical application of BM-MSCs or HGF need to be strengthened further to establish BM-MSCs or HGF as effective therapeutic tool in regenerative medicine. A further useful detail of BM-MSCs with particular reference to their derivation from NG2-expressing PCs and their potential for possible use in tissue regeneration and repair warrants further study. At present, we are working on a critical issue: Can NG2⁺ cells purified from BM (BM-NG2⁺ cells) generate newly MSCs?

To address this question, it is a considerable value to purified NG2⁺ cells from adult mammals BM. we have used the same procedure as CNS-NG2⁺ cells and eye (Iris)-NG2⁺ cells and purified this cell population. We found that NG2-expressing cells can be purified from BM. Based on our experiments we presented a remarkable outcome. As in images showed (Figure 3) that approximately >95% of BM isolated cells express NG2 proteoglycan (BM-MG2⁺ cells) and performed not only self renewal but also differentiated into CD90-expressing



MSCs. We propose that these cells may like NG2⁺ cells located other organs have local functions in the tissue microenvironment beyond mesenchymal differentiation and functional outcome in response to insult signals. This study strongly supported our hypothesized: BM-NG2⁺ cells are the reservoir of MSCs. The NG2-derived MSCs could secrete a variety of beneficial factors that modulates immune response and promote endogenous regeneration. We also proposed that the possibility of a reservoir of NG2-expressing PCs for MSCs that would have the similar biological and functional capacities that survey tissue repair [69,83-87].

Are NG2⁺ Cells from Outside CNS Organs the Originators in Tissue Repair?

Similarity of functional properties of NG2⁺ cells, MSCs and PCs exist in multiple diseased organs is entirely unclear. The MSCs isolated from multiple adult organs proliferate extensively in the cultures of unselected. PCs, as a monolayer of dendritic-like cells closely ensheath endothelial cells within capillaries and microvessels (arterioles and venules), have been recently identified as possible originators of MSCs [88]. It is clear that purified perivascular cells express NG2 which exhibit multiple mesodermal developmental potentials, and become indistinguishable from conventionally derived MSCs after *in vitro* culture [89]. However, the possible roles played by these blood vessel-bound NG2-expressing PCs and MSCs in organogenesis and adult tissue repair remain elusive. Better knowledge of the lineage affiliation of these cells will contribute to the development of more efficient and tissue repair.

Adult cardiovascular NG2⁺ cells in heart failure repair

Homeostasis of different cell types in cardiovascular wall is essential for maintaining heart function. In physiological conditions, the turnover rate of vascular cells is low but greatly increased in diseased situations, e.g. vascular injury after angioplasty. It is believed that mature vascular cells have an ability to proliferate to replace lost cells and recent evidence indicates stem/progenitor cells may participate in vascular repair in damaged vessels.

In general, there are three layers (intima, media, and adventitia) of immature cardiovascular vessels. Some progenitor cells also reside in cardiovascular vessel such as endothelial progenitors, MSCs, Sca-1⁺, CD34⁺ and NG2⁺ cells [90-92]. These resident progenitor cells quietly persist in adult healthy heart and waiting for the opportunity for cell therapy in response to injury signals. Below we discuss the data suggesting the presence of NG2-expressing cells that reside in the adult heart exhibit PCs characteristics. Mature cardiovascular vessels have three cellular parts: PCs, endothelial cells (ECs) and smooth muscle cells (SMCs) that have developed independently during development [93]. After maturation, the role of progenitors in healthy and pathological processes has been well documented in different animal models [94,95]. It takes long time to understand how those progenitors contributed to vascular homeostasis, proliferation, angiogenesis and tissue repair. An interesting study recently showed that PCs being as progenitors that response to local injury by differentiating into functional cells [96].

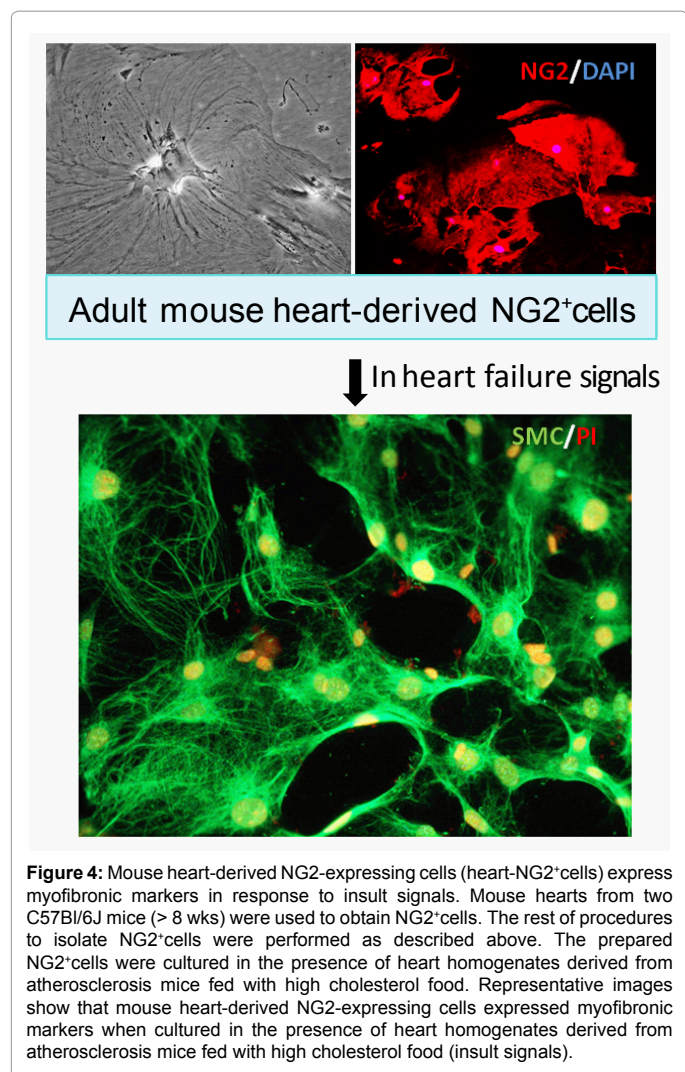
Several aspects contribute to defining a suitable source of MSCs for treatment of different insult conditions. Tissue injury may require relatively few cells places in a specific location adjacent to the area of damage, while PCs in the wall of blood vessels may require damage signal stimulation to give rise to MSCs. A study suggested that aortic arch is the major source of PCs and aortic arch-derived PCs express NG2 [97]. Further study revealed that NG2⁺ cells that isolated from

aorta arch were provided with vasculogenesis, suggesting PC potential [98]. Notably, PCs jointing with capillary ECs, indicating NG2-expressing PCs can be regarded as the structural components of blood vessels to regulate vascular contractility and perform functional task through generated MSCs [60,70,99-102]. We propose NG2 expressing PCs cells in adult heart blood vessel are an attracted cells. They may endue characteristics with PC and generate MSCs in response to insult with broad developmental potential for heart failure [103].

PCs distribute not only in veins but arteries as well. They do not express markers of hematopoietic stem cells (CD34, CD45) and endothelial cells (vWF) [104]. A study showed that cultured a cell population that generated from rat aortic in a collagen and bFGF containing conditioned media, the cells formed spheroid colonies and cell surface antigens were identified by NG2, PDGFR- β and nestin antibodies [105], suggesting cardiovascular vessel wall process PC-like NG2⁺ cells. Recent histological observations showed that blood vessel wall-derived NG2-expressing PCs can also differentiate into adipocytes, chondrocytes, osteoblasts within calcified atherosclerotic lesions [106,107], suggesting their mesenchymal origin and adipogenic and osteogenic potential.

NG2-expressing PCs per se serve as blood vessel progenitor cells [108,109] and can generate cardiomyocytes, smooth muscles and vascular EC in homeostatic conditions and after myocardial injuries for constitution of myocardium within the adult heart. Isolation and stimulation of PCs in adult heart may represent a potential therapeutic strategy to treat heart failures [110-112]. Consistent with this notion a number of studies of cardiac injury in animal model have suggested the clinical potential of heart-derived NG2-expressing PCs as progenitors in regeneration following cardiac injury [113]. Another study showed that grafted NG2-expressing cells that isolated from blood vessels into animal model, improved cardiac fabric construction and function through the combined effect of myogenesis and angiogenesis has been observed [100,114]. For example, the limited capacity of the adult human heart to spontaneously repair its damaged muscle following myocardial infarction (MI), an ischemia animal model, leads to loss of cardiac function, ventricular remodeling and progressive dysfunction frequently leading to heart failure. So prolonged heart ischemia caused by atherosclerosis or thromboemboli results in ischemic cardiomyopathy (IC) and myocardial infarction (MI), which may in turn elicit heart failure and death [115].

In clinic, despite improvements in emergency treatment, myocardial infarction often leads to congestive heart failure. Other than heart transplantation, current therapies are aimed at enabling the patient to survive with a heart working at a fraction of its original capacity. It is therefore no surprise that cardiac stem cell therapy has raised many hope, especially MSCs are better studies in animal models [116]. Unfortunately, neither the ideal source and type of stem cell nor the critical cell number and mode of application have been defined so far. If it is the case that NG2⁺ cells can be the source of MSCs that would cross a variety of faced obstacles in current clinical settings. A large gap in our understanding of what factors control NG2-expressing PCs differentiate into functional cells. In order to bridge this gap, we have purified NG2⁺ cells from adult mouse heart aortic arch (heart-NG2⁺ cells) by using the same procedure as CNS-NG2⁺ cells, eye (Iris)-NG2⁺ cells and BM-NG2⁺ cells and cultured them with myocardial infarction (MI) homogenate (Figure 4). Interestingly, we found that the heart-NG2⁺ cells turn on SMCs-expressing myofibronic progenitor cells when they present in MI homogenate.



NG2⁺ cells in other multiple organs for tissue repair

Existed NG2⁺ cells have also been recognized in other adult organs including heart, liver, pancreas, lung and kidney where they maintain specialized functions [117], suggest these cells may act as contractile cells and obligatory regulators for vascular development, stabilization, maturation and tissue remodeling.

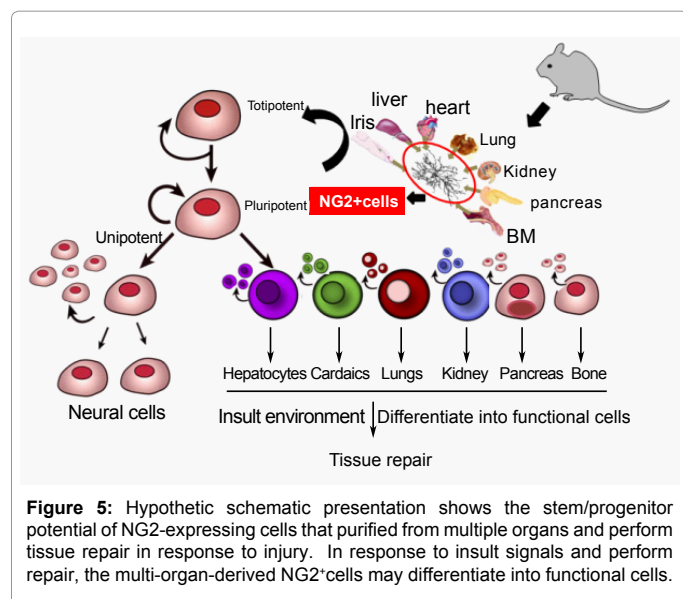
NG2-expressing cells in adult rodent liver share similar characteristics with PCs [99]. They express both PC (PDGFR- β , CD146) and MSC markers (CD105, CD72) but do not express EC, astrocyte (GFAP) and hematopoietic stem cell (CD34) markers. The NG2-expressing PCs also shared some cell phenotypic markers with the MSCs that isolated from other organs [118] and possess an intermediate phenotype between vascular SMCs and fibroblasts with a capacity to differentiate into a myofibroblast phenotype. Morphologically, PCs in adult mouse liver possess long processes embracing the abluminal endothelium wall in pre-capillary arterioles, capillaries, and post-capillary venues. Long cytoplasmic processes that extend along and encircle the endothelial tube ideally position these cells for paracrine signaling with ECs. These varied morphologies have been translated into diverse functions. Interestingly, anatomic characteristics of hepatic stellate cells (HSCs) have led to the distinction of HSCs as an

adult liver-specific PCs [107,119]. This new concept is recapitulated by studies of PCs in other organs that identify the protein role and relevant mechanisms of this cell type for vascular development and function.

In adult mammalian liver, the complementary processes of angiogenesis (vascular growth from preexisting vessels) and vasculogenesis (*de novo* blood vessel development) regulate vascular development and homeostasis. Liver regenerative medicine for liver disease repair is much related to this regulation and focused majorly on three major areas: (1) Liver cirrhosis, to improve regeneration and reduce scarring by modulating the liver's own regenerative processes, (2) Immunity-mediated liver damage to down-regulate inflammatory immune response, and (3) to offer hepatic cell differentiation from transplanted stem/progenitor to supplement or replace hepatocyte function. There were several studies reported that NG2-expressing cells in blood vessels express NG2 and CD146 proteoglycan, suggesting PC progenitor potential [120,121] although these rely mostly on indirect evidence. Of clinical interest, the role of PC-like HSCs in hepatic angiogenesis and remodeling comes from work using a pre-clinical animal model (liver fibrosis or cirrhosis) for liver regeneration [122-125].

The development of advanced fibrosis presents three further animal models that also showed similarities with NG2-expressing PC behaviors. In the first, the development of fibrosis and its spontaneous resolution, there is a progressive activation of HSCs to become myofibroblast-like progenitor cells. In the second, a form of sinusoidal remodeling occurs in more advanced cirrhotic liver. These scars contain myofibroblasts, likely derived from activated HSCs. Intriguingly, if a comparison is drawn between armature scar within a cirrhotic liver and a less mature scar, the cells within the mature scar express markers more commonly associated with HSCs than myofibroblasts progenitor cells, such as glial fibrillary acidic protein and desmin in the absence of α -SMA [126]. Lastly, good evidence now exists in the models of advanced human and rodent fibrosis for bone marrow stem cell contribution to the myofibroblast progenitor and HSC population [127,128]. However, each of the aforementioned differentiation experiments was performed on unpurified PCs. Take together with those *in vivo* experiments may support to understand mechanisms by which HSCs operate as "contractile machinery" and relax in response to toxic agents [129-131]. Good evidence supports a contraction-based vasoconstrictive function of HSCs in the setting of cirrhosis but the role of HSCs as a regulator of vascular tone in normal liver is less established. These results demonstrated that modulation of vasoactive agents such as endothelin and carbon monoxide cause changes in sinusoidal diameter at locations where HSCs reside [132]. The process of sinusoidal remodeling could be distinct from the more characterized role of HSCs in the process of collagen deposition and fibrosis [133], suggesting that these vasoregulatory changes occur outside of the hepatic sinusoids [134,135]. In liver cirrhosis, partial cirrhotic reversal is highly related to the level of fibrotic resolution and the state of cirrhotic progress [136] and these primary phenomena raise the possibility of slowing the development of liver cirrhosis [137].

According to a range of studies described above about NG2⁺ cell stem/progenitor potential, we propose adult NG2⁺ cells that purified from multiple organs such as CNS-NG2⁺ cells, eye (Iris-NG2), BM-NG2⁺ cells, liver-NG2⁺ cells, heart-NG2⁺ cells, pancreas-NG2⁺ cells, lung-NG2⁺ cells and kidney-NG2⁺ cells may all share the similar biological characteristics. They express both PCs (PDGFR- β , CD146) and MSC markers (CD73/CD105). They may have stem/progenitor potential in response to insult signals and involve in repair processes (Figure 5).



Unanswered Questions and Perspectives

Based on in the search for proof of resident NG2-expressing cells in the adult CNS, outside CNS organs and our preliminary studies demonstrate that the NG2⁺cells would be a novel progenitor cells in regenerative medicine.

Much work has been done in the developmental system of the NG2⁺cells, but many problems remain unsolved in the adult system. Although we have developed a novel approach for gleaning a sufficient number of NG2⁺cell from adult multiple mouse organs, we still don't completely understand what is the potential role of these NG2-expressing stem/progenitors in adult mammalian organs. For example: (1) the possibility of tumorigenesis has not been tested. We propose, like other adult stem/progenitor cells such as MSCs, these adult NG2⁺cells may be relatively safe compared to ESCs and iPSCs which readily form teratomas. (2) What is the nature of these adult NG2⁺cells residing in the multiple organs? Are they "permanent resident" or just a "crossing guest" in blood vessel wall? We propose in normal condition, these adult NG2⁺cells like traditional progenitors that has capacity to generate newly different cell types and reside "quietly" in blood vessel wall but can be activated in response to different insult semaphores not only injury alone to differentiate rapidly into repair cells. (3) Although a body of evidences provided by the review that these multiple adult organ-derived NG2⁺cells share characteristics with PCs and MSCs, further research still need to be done on their different in unique niches, the contours underlying their genomics, proteomics and glycomics compare to traditional stem/progenitor cells, as well as the degree of immune rejection.

In sum, appeared adult NG2⁺cells on the stage are the population that locates not only in the CNS as traditional thought but also outside CNS multiple organs such as eyes, bone marrow, heart, liver, lung, pancreas and kidney (multi-NG2⁺cells). More analysis of multi-NG2⁺cells will provide valuable insights into the cellular and molecular pathways that mediate recovery from multiple organ diseases. Such researches are fundamentally important for setting novel cell-based therapy in regenerative medicine.

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References

- Raff MC, Miller RH, Noble M (1983) A glial progenitor cell that develops in vitro into an astrocyte or an oligodendrocyte depending on culture medium. *Nature* 303: 390-396.
- Robinson S, Miller RH (1999) Contact with central nervous system myelin inhibits oligodendrocyte progenitor maturation. *Dev Biol* 216: 359-368.
- Dawson MR, Levine JM, Reynolds R (2000) NG2-expressing cells in the central nervous system: are they oligodendroglial progenitors? *J Neurosci Res* 61: 471-479.
- Blum R, Heinrich C, Sánchez R, Lepier A, Gundelfinger ED, et al. (2011) Neuronal network formation from reprogrammed early postnatal rat cortical glial cells. *Cereb Cortex* 21: 413-424.
- Faber-Zuschratter H, Hüttmann K, Steinhäuser C, Becker A, Schramm J, et al. (2009) Ultrastructural and functional characterization of satellitosis in the human lateral amygdala associated with Ammon's horn sclerosis. *Acta Neuropathol* 117: 545-555.
- Belachew S, Chittajallu R, Aguirre AA, Yuan X, Kirby M, et al. (2003) Postnatal NG2 proteoglycan-expressing progenitor cells are intrinsically multipotent and generate functional neurons. *J Cell Biol* 161: 169-86.
- Keirstead HS, Levine JM, Blakemore WF (1998) Response of the oligodendrocyte progenitor cell population (defined by NG2 labelling) to demyelination of the adult spinal cord. *Glia* 22: 161-170.
- Mallon BS, Shick HE, Kidd GJ, Macklin WB (2002) Proteolipid promoter activity distinguishes two populations of NG2-positive cells throughout neonatal cortical development. *J Neurosci* 22: 876-885.
- Oderfeld-Nowak B, Zaremba M, Kwiatkowska-Patzer B, Lipkowski AW, Kurkowska-Jastrzebska I, et al. (2009) NG2 positive cells of rat spinal cord activated during experimental autoimmune encephalomyelitis are spatially associated with radially oriented astroglia and express p75 receptor: a role for nerve growth factor in oligodendrocyte progenitor migration? *Arch Ital Biol* 147: 105-15.
- Polito A, Reynolds R (2005) NG2-expressing cells as oligodendrocyte progenitors in the normal and demyelinated adult central nervous system. *J Anat* 207: 707-716.
- Levine JM, Stincone F, Lee YS (1993) Development and differentiation of glial precursor cells in the rat cerebellum. *Glia* 7: 307-321.
- Tang DG, Tokumoto YM, Raff MC (2000) Long-term culture of purified postnatal oligodendrocyte precursor cells. Evidence for an intrinsic maturation program that plays out over months. *J Cell Biol* 148: 971-984.
- Stallcup WB, Beasley L (1987) Bipotential glial precursor cells of the optic nerve express the NG2 proteoglycan. *J Neurosci* 7: 2737-2744.
- Wilson PO, Barber PC, Hamid QA, Power BF, Dhillon AP, et al. (1988) The immunolocalization of protein gene product 9.5 using rabbit polyclonal and mouse monoclonal antibodies. *Br J Exp Pathol* 69: 91-104.
- Nishiyama A, Komitova M, Suzuki R, Zhu X (2009) Polydendrocytes (NG2 cells): multifunctional cells with lineage plasticity. *Nat Rev Neurosci* 10: 9-22.
- Dawson MR, Polito A, Levine JM, Reynolds R (2003) NG2-expressing glial progenitor cells: an abundant and widespread population of cycling cells in the adult rat CNS. *Mol Cell Neurosci* 24: 476-488.
- Horner PJ, Power AE, Kempermann G, Kuhn HG, Palmer TD, et al. (2000) Proliferation and differentiation of progenitor cells throughout the intact adult rat spinal cord. *J Neurosci* 20: 2218-2228.
- Bergles DE, Roberts JD, Somogyi P, Jahr CE (2000) Glutamatergic synapses on oligodendrocyte precursor cells in the hippocampus. *Nature* 405: 187-191.
- Ong WY, Levine JM (1999) A light and electron microscopic study of NG2 chondroitin sulfate proteoglycan-positive oligodendrocyte precursor cells in the normal and kainate-lesioned rat hippocampus. *Neuroscience* 92: 83-95.
- Bumol TF, Chee DO, Reisfeld RA (1982) Immunochemical and biosynthetic analysis of monoclonal antibody-defined melanoma-associated antigen. *Hybridoma* 1: 283-292.
- Bumol TF, Walker LE, Reisfeld RA (1984) Biosynthetic studies of proteoglycans

- in human melanoma cells with a monoclonal antibody to a core glycoprotein of chondroitin sulfate proteoglycans. *J Biol Chem* 259: 12733-12741.
22. Houghton AN, Eisinger M, Albino AP, Cairncross JG, Old LJ (1982) Surface antigens of melanocytes and melanomas. Markers of melanocyte differentiation and melanoma subsets. *J Exp Med* 156: 1755-1766.
23. Desai SA, Wang X, Noronha EJ, Kageshita T, Ferrone S (1998) Characterization of human anti-high molecular weight-melanoma-associated antigen single-chain Fv fragments isolated from a phage display antibody library. *Cancer Res* 58: 2417-2425.
24. Lin SC, Bergles DE (2004) Synaptic signaling between neurons and glia. *Glia* 47: 290-298.
25. Kirchhoff F, Kettenmann H (1992) GABA Triggers a $[Ca^{2+}]_i$ Increase in Murine Precursor Cells of the Oligodendrocyte Lineage. *Eur J Neurosci* 4: 1049-1058.
26. Yuan X, Eisen AM, McBain CJ, Gallo V (1998) A role for glutamate and its receptors in the regulation of oligodendrocyte development in cerebellar tissue slices. *Development* 125: 2901-2914.
27. Levine JM, Reynolds R, Fawcett JW (2001) The oligodendrocyte precursor cell in health and disease. *Trends Neurosci* 24: 39-47.
28. Zawadzka M, Rivers LE, Fancy SP, Zhao C, Tripathi R, et al. (2010) CNS-resident glial progenitor/stem cells produce Schwann cells as well as oligodendrocytes during repair of CNS demyelination. *Cell Stem Cell* 6: 578-590.
29. Kucharova K, Chang Y, Boor A, Yong VW, Stallcup WB (2011) Reduced inflammation accompanies diminished myelin damage and repair in the NG2 null mouse spinal cord. *J Neuroinflammation* 8: 158.
30. Keirstead HS, Blakemore WF (1999) The role of oligodendrocytes and oligodendrocyte progenitors in CNS remyelination. *Adv Exp Med Biol* 468: 183-197.
31. Di Bello IC, Dawson MR, Levine JM, Reynolds R (1999) Generation of oligodendroglial progenitors in acute inflammatory demyelinating lesions of the rat brain stem is associated with demyelination rather than inflammation. *J Neurocytol* 28: 365-381.
32. Watanabe M, Toyama Y, Nishiyama A (2002) Differentiation of proliferated NG2-positive glial progenitor cells in a remyelinating lesion. *J Neurosci Res* 69: 826-836.
33. Bai L, Hecker J, Kerstetter A, Miller RH (2013) Myelin repair and functional recovery mediated by neural cell transplantation in a mouse model of multiple sclerosis. *Neurosci Bull* 29: 239-250.
34. Lee ST, Chu K, Park JE, Lee K, Kang L, et al. (2005) Intravenous administration of human neural stem cells induces functional recovery in Huntington's disease rat model. *Neurosci Res* 52: 243-249.
35. Kondo T, Raff M (2000) Oligodendrocyte precursor cells reprogrammed to become multipotential CNS stem cells. *Science* 289: 1754-1757.
36. Levine JM, Stallcup WB (1987) Plasticity of developing cerebellar cells in vitro studied with antibodies against the NG2 antigen. *J Neurosci* 7: 2721-2731.
37. Butt AM, Duncan A, Hornby MF, Kirvell SL, Hunter A, et al. (1999) Cells expressing the NG2 antigen contact nodes of Ranvier in adult CNS white matter. *Glia* 26: 84-91.
38. Nishiyama A, Watanabe M, Yang Z, Bu J (2002) Identity, distribution, and development of polydendrocytes: NG2-expressing glial cells. *J Neurocytol* 31: 437-455.
39. Hampton DW, Asher RA, Kondo T, Steeves JD, Ramer MS, et al. (2007) A potential role for bone morphogenetic protein signalling in glial cell fate determination following adult central nervous system injury in vivo. *Eur J Neurosci* 26: 3024-3035.
40. Ffrench-Constant C, Raff MC (1986) Proliferating bipotential glial progenitor cells in adult rat optic nerve. *Nature* 319: 499-502.
41. Greenwood K, Butt AM (2003) Evidence that perinatal and adult NG2-glia are not conventional oligodendrocyte progenitors and do not depend on axons for their survival. *Mol Cell Neurosci* 23: 544-558.
42. Matthias K, Kirchhoff F, Seifert G, Hüttmann K, Matyash M, et al. (2003) Segregated expression of AMPA-type glutamate receptors and glutamate transporters defines distinct astrocyte populations in the mouse hippocampus. *J Neurosci* 23: 1750-1758.
43. Berry M, Hubbard P, Butt AM (2002) Cytology and lineage of NG2-positive glia. *J Neurocytol* 31: 457-467.
44. Barres BA, Raff MC (1993) Proliferation of oligodendrocyte precursor cells depends on electrical activity in axons. *Nature* 361: 258-260.
45. David S, Miller RH, Patel R, Raff MC (1984) Effects of neonatal transection on glial cell development in the rat optic nerve: evidence that the oligodendrocyte-type 2 astrocyte cell lineage depends on axons for its survival. *J Neurocytol* 13: 961-974.
46. Reynolds R, Cenci di Bello I, Dawson M, Levine J (2001) The response of adult oligodendrocyte progenitors to demyelination in EAE. *Prog Brain Res* 132: 165-174.
47. Chang A, Nishiyama A, Peterson J, Prineas J, Trapp BD (2000) NG2-positive oligodendrocyte progenitor cells in adult human brain and multiple sclerosis lesions. *J Neurosci* 20: 6404-6412.
48. Butt AM (2006) Neurotransmitter-mediated calcium signalling in oligodendrocyte physiology and pathology. *Glia* 54: 666-675.
49. Lin SC, Bergles DE (2002) Physiological characteristics of NG2-expressing glial cells. *J Neurocytol* 31: 537-549.
50. Kukley M, Capetillo-Zarate E, Dietrich D (2007) Vesicular glutamate release from axons in white matter. *Nat Neurosci* 10: 311-320.
51. Hamilton N, Hubbard PS, Butt AM (2009) Effects of glutamate receptor activation on NG2-glia in the rat optic nerve. *J Anat* 214: 208-218.
52. Butt AM, Hamilton N, Hubbard P, Pugh M, Ibrahim M (2005) Synantocytes: the fifth element. *J Anat* 207: 695-706.
53. Haberlandt C, Derouiche A, Wyczynski A, Haseleu J, Pohle J, et al. (2011) Gray matter NG2 cells display multiple Ca^{2+} -signaling pathways and highly motile processes. *PLoS One* 6: e17575.
54. Mangin JM, Gallo V (2011) The curious case of NG2 cells: transient trend or game changer? *ASN Neuro* 3: e00052.
55. Matute C, Domercq M, Sánchez-Gómez MV (2006) Glutamate-mediated glial injury: mechanisms and clinical importance. *Glia* 53: 212-224.
56. Matute C (1998) Characteristics of acute and chronic kainate excitotoxic damage to the optic nerve. *Proc Natl Acad Sci U S A* 95: 10229-10234.
57. Matute C, Sánchez-Gómez MV, Martínez-Millán L, Miledi R (1997) Glutamate receptor-mediated toxicity in optic nerve oligodendrocytes. *Proc Natl Acad Sci U S A* 94: 8830-8835.
58. Kozanoglu I, Boga C, Ozdogu H, Sozer O, Maytalman E, et al. (2009) Human bone marrow mesenchymal cells express NG2: possible increase in discriminative ability of flow cytometry during mesenchymal stromal cell identification. *Cytotherapy* 11: 527-533.
59. Yamanishi H, Fujiwara S, Soma T (2012) Perivascular localization of dermal stem cells in human scalp. *Exp Dermatol* 21: 78-80.
60. Covas DT, Panepucci RA, Fontes AM, Silva WA Jr, Orellana MD, et al. (2008) Multipotent mesenchymal stromal cells obtained from diverse human tissues share functional properties and gene-expression profile with CD146+ perivascular cells and fibroblasts. *Exp Hematol* 36: 642-654.
61. Caplan AI (2007) Adult mesenchymal stem cells for tissue engineering versus regenerative medicine. *J Cell Physiol* 213: 341-347.
62. Caplan AI, Dennis JE (2006) Mesenchymal stem cells as trophic mediators. *J Cell Biochem* 98: 1076-1084.
63. Di Nicola M, Carlo-Stella C, Magni M, Milanesi M, Longoni PD, et al. (2002) Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Blood* 99: 3838-3843.
64. Gerdoni E, Gallo B, Casazza S, Musio S, Bonanni I, et al. (2007) Mesenchymal stem cells effectively modulate pathogenic immune response in experimental autoimmune encephalomyelitis. *Ann Neurol* 61: 219-227.
65. Nauta AJ, Fibbe WE (2007) Immunomodulatory properties of mesenchymal stromal cells. *Blood* 110: 3499-3506.
66. Caplan AI (2008) All MSCs are pericytes? *Cell Stem Cell* 3: 229-230.
67. Zebardast N, Lickorish D, Davies JE (2010) Human umbilical cord perivascular cells (HUCPVC): A mesenchymal cell source for dermal wound healing. *Organogenesis* 6: 197-203.

68. Lennon DP, Caplan AI (2006) Isolation of human marrow-derived mesenchymal stem cells. *Exp Hematol* 34: 1604-1605.
69. Caplan AI, Correa D (2011) The MSC: an injury drugstore. *Cell Stem Cell* 9: 11-15.
70. Crisan M, Yap S, Casteilla L, Chen CW, Corselli M, et al. (2008) A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell* 3: 301-313.
71. Dörmecq M, Matute C (1999) Expression of glutamate transporters in the adult bovine corpus callosum. *Brain Res Mol Brain Res* 67: 296-302.
72. Hirschi KK, D'Amore PA (1996) Pericytes in the microvasculature. *Cardiovasc Res* 32: 687-698.
73. Choi YS, Jeong JA, Lim DS (2012) Mesenchymal stem cell-mediated immature dendritic cells induce regulatory T cell-based immunosuppressive effect. *Immunol Invest* 41: 214-229.
74. Kassiss I, Vaknin-Dembinsky A, Karussis D (2011) Bone marrow mesenchymal stem cells: agents of immunomodulation and neuroprotection. *Curr Stem Cell Res Ther* 6: 63-68.
75. Reading JL, Sabbah S, Busch S, Tree TJ (2013) Mesenchymal stromal cells as a means of controlling pathological T-cell responses in allogeneic islet transplantation. *Curr Opin Organ Transplant* 18: 59-64.
76. Bai L, Lennon DP, Eaton V, Maier K, Caplan AI, et al. (2009) Human bone marrow-derived mesenchymal stem cells induce Th2-polarized immune response and promote endogenous repair in animal models of multiple sclerosis. *Glia* 57: 1192-1203.
77. Traktuev DO, Merfeld-Clauss S, Li J, Kolonin M, Arap W, et al. (2008) A population of multipotent CD34-positive adipose stromal cells share pericyte and mesenchymal surface markers, reside in a periendothelial location, and stabilize endothelial networks. *Circ Res* 102: 77-85.
78. Diaz-Flores L Jr, Gutierrez R, Madrid JF, Varela H, Valladares F, et al. (2009) Adult stem cells and repair through granulation tissue. *Front Biosci* 14: 1433-1470.
79. Caplan AI (1991) Mesenchymal stem cells. *J Orthop Res* 9: 641-650.
80. Arboleda D, Forostyck S, Jendelova P, Marekova D, Amemori T, et al. (2011) Transplantation of predifferentiated adipose-derived stromal cells for the treatment of spinal cord injury. *Cell Mol Neurobiol* 31: 1113-1122.
81. Daquinag AC, Zhang Y, Amaya-Manzanares F, Simmons PJ, Kolonin MG (2011) An isoform of decorin is a resistin receptor on the surface of adipose progenitor cells. *Cell Stem Cell* 9: 74-86.
82. Jones BJ, McTaggart SJ (2008) Immunosuppression by mesenchymal stromal cells: from culture to clinic. *Exp Hematol* 36: 733-741.
83. Le Blanc K, Tammik L, Sundberg B, Haynesworth SE, Ringdén O (2003) Mesenchymal stem cells inhibit and stimulate mixed lymphocyte cultures and mitogenic responses independently of the major histocompatibility complex. *Scand J Immunol* 57: 11-20.
84. Ezquer F, Ezquer M, Contador D, Ricca M, Simon V, et al. (2012) The antidiabetic effect of mesenchymal stem cells is unrelated to their transdifferentiation potential but to their capability to restore Th1/Th2 balance and to modify the pancreatic microenvironment. *Stem Cells* 30: 1664-1674.
85. Joyce N, Annett G, Wirthlin L, Olson S, Bauer G, et al. (2010) Mesenchymal stem cells for the treatment of neurodegenerative disease. *Regen Med* 5: 933-946.
86. Shabbir A, Zisa D, Suzuki G, Lee T (2009) Heart failure therapy mediated by the trophic activities of bone marrow mesenchymal stem cells: a noninvasive therapeutic regimen. *Am J Physiol Heart Circ Physiol* 296: H1888-1897.
87. Sorrell JM, Baber MA, Caplan AI (2009) Influence of adult mesenchymal stem cells on in vitro vascular formation. *Tissue Eng Part A* 15: 1751-1761.
88. Rehman MS, Student RI (2012) Dietary saturated fat intake, is there really an association with coronary heart disease? *J Pak Med Assoc* 62: 411.
89. Wagner J, Kean T, Young R, Dennis JE, Caplan AI (2009) Optimizing mesenchymal stem cell-based therapeutics. *Curr Opin Biotechnol* 20: 531-536.
90. Bai L, Lennon DP, Caplan AI, DeChant A, Hecker J, et al. (2012) Hepatocyte growth factor mediates mesenchymal stem cell-induced recovery in multiple sclerosis models. *Nat Neurosci* 15: 862-870.
91. Dellavalle A, Sampaolesi M, Tonlorenzi R, Tagliafico E, Sacchetti B, et al. (2007) Pericytes of human skeletal muscle are myogenic precursors distinct from satellite cells. *Nat Cell Biol* 9: 255-267.
92. Giordano A, Galderisi U, Marino IR (2007) From the laboratory bench to the patient's bedside: an update on clinical trials with mesenchymal stem cells. *J Cell Physiol* 211: 27-35.
93. Mackay-Lyons M, Thornton M, Macdonald A (2011) Cardiovascular fitness training for a patient in the early stages of recovery post stroke. *Physiother Can* 63: 377-382.
94. Pittenger MF (2008) Mesenchymal stem cells from adult bone marrow. *Methods Mol Biol* 449: 27-44.
95. Prockop DJ (1997) Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 276: 71-74.
96. Corselli M, Chen CW, Sun B, Yap S, Rubin JP, et al. (2012) The tunica adventitia of human arteries and veins as a source of mesenchymal stem cells. *Stem Cells Dev* 21: 1299-1308.
97. Caplan AI, Correa D (2011) PDGF in bone formation and regeneration: new insights into a novel mechanism involving MSCs. *J Orthop Res* 29: 1795-1803.
98. Hoch M, Fischer P, Stapel B, Missol-Kolka E, Sekkali B, et al. (2011) Erythropoietin preserves the endothelial differentiation capacity of cardiac progenitor cells and reduces heart failure during anticancer therapies. *Cell Stem Cell* 9: 131-143.
99. Mokry J, Ehrmann J, Karbanová J, Cizková D, Soukup T, et al. (2008) Expression of intermediate filament nestin in blood vessels of neural and non-neural tissues. *Acta Medica (Hradec Kralove)* 51: 173-179.
100. Ozerdem U, Grako KA, Dahlin-Huppe K, Monosov E, Stallcup WB (2001) NG2 proteoglycan is expressed exclusively by mural cells during vascular morphogenesis. *Dev Dyn* 222: 218-227.
101. Xiong JW (2008) Molecular and developmental biology of the hemangioblast. *Dev Dyn* 237: 1218-1231.
102. Xu Q (2006) The impact of progenitor cells in atherosclerosis. *Nat Clin Pract Cardiovasc Med* 3: 94-101.
103. Aicher A, Zeiher AM, Dimmeler S (2005) Mobilizing endothelial progenitor cells. *Hypertension* 45: 321-325.
104. Torsney E, Xu Q (2011) Resident vascular progenitor cells. *J Mol Cell Cardiol* 50: 304-311.
105. Pasquinelli G, Pacilli A, Alviano F, Foroni L, Ricci F, et al. (2010) Multidistrict human mesenchymal vascular cells: pluripotency and stemness characteristics. *Cytotherapy* 12: 275-287.
106. Wilkinson FL, Liu Y, Rucka AK, Jeziorska M, Hoyland JA, et al. (2007) Contribution of VCAF-positive cells to neovascularization and calcification in atherosclerotic plaque development. *J Pathol* 211: 362-369.
107. Johnson RC, Leopold JA, Loscalzo J (2006) Vascular calcification: pathobiological mechanisms and clinical implications. *Circ Res* 99: 1044-1059.
108. Wigley R, Butt AM (2009) Integration of NG2-glia (synantocytes) into the neuroglial network. *Neuron Glia Biol* 5: 21-28.
109. Iwasaki K, Komaki M, Yokoyama N, Tanaka Y, Taki A, et al. (2012) Periodontal Ligament Stem Cells Possess the Characteristics of Pericytes. *J Periodontol* .
110. O'Brien JE Jr, Shi Y, Fard A, Bauer T, Zalewski A, et al. (1997) Wound healing around and within saphenous vein bypass grafts. *J Thorac Cardiovasc Surg* 114: 38-45.
111. Chen CW, Okada M, Proto JD, Gao X, Sekiya N, et al. (2013) Human pericytes for ischemic heart repair. *Stem Cells* 31: 305-316.
112. Andreeva ER, Pugach IM, Gordon D, Orekhov AN (1998) Continuous subendothelial network formed by pericyte-like cells in human vascular bed. *Tissue Cell* 30: 127-135.
113. Tárnok A, Ulrich H, Bocsi J (2010) Phenotypes of stem cells from diverse origin. *Cytometry A* 77: 6-10.
114. Fisher M (2009) Pericyte signaling in the neurovascular unit. *Stroke* 40: S13-15.
115. Sims DE (2000) Diversity within pericytes. *Clin Exp Pharmacol Physiol* 27: 842-846.

116. Díaz-Flores L, Gutiérrez R, Varela H, Rancel N, Valladares F (1991) Microvascular pericytes: a review of their morphological and functional characteristics. *Histol Histopathol* 6: 269-286.
117. Howson KM, Aplin AC, Gelati M, Alessandri G, Parati EA, et al. (2005) The postnatal rat aorta contains pericyte progenitor cells that form spheroidal colonies in suspension culture. *Am J Physiol Cell Physiol* 289: C1396-407.
118. Canfield AE, Doherty MJ, Wood AC, Farrington C, Ashton B, et al. (2000) Role of pericytes in vascular calcification: a review. *Z Kardiol* 89 Suppl 2: 20-27.
119. Bollini S, Smart N, Riley PR (2011) Resident cardiac progenitor cells: at the heart of regeneration. *J Mol Cell Cardiol* 50: 296-303.
120. Leri A, Kajstura J, Anversa P (2005) Cardiac stem cells and mechanisms of myocardial regeneration. *Physiol Rev* 85: 1373-1416.
121. Ballard VL, Edelberg JM (2007) Stem cells and the regeneration of the aging cardiovascular system. *Circ Res* 100: 1116-1127.
122. Rosmorduc O, Housset C (2010) Hypoxia: a link between fibrogenesis, angiogenesis, and carcinogenesis in liver disease. *Semin Liver Dis* 30: 258-270.
123. Chernousov AF, Khorobrykh TV, Karpova RV, Nekrasova TP (2013) Regeneration of cirrhotic liver in rabbits after intrahepatic injection of cryoprecipitate. *Bull Exp Biol Med* 154: 396-398.
124. Li Y, Wang J, Asahina K (2013) Mesothelial cells give rise to hepatic stellate cells and myofibroblasts via mesothelial-mesenchymal transition in liver injury. *Proc Natl Acad Sci U S A* 110: 2324-2329.
125. Yokoi Y, Namiyama T, Kuroda H, Komatsu I, Miyazaki A, et al. (1984) Immunocytochemical detection of desmin in fat-storing cells (Ito cells). *Hepatology* 4: 709-714.
126. Zhu NL, Asahina K, Wang J, Ueno A, Lazaro R, et al. (2012) Hepatic stellate cell-derived delta-like homolog 1 (DLK1) protein in liver regeneration. *J Biol Chem* 287: 10355-10367.
127. Schotanus BA, van den Ingh TS, Penning LC, Rothuizen J, Roskams TA, et al. (2009) Cross-species immunohistochemical investigation of the activation of the liver progenitor cell niche in different types of liver disease. *Liver Int* 29: 1241-1252.
128. Kordes C, Sawitz A, Müller-Marbach A, Ale-Agha N, Keitel V, et al. (2007) CD133+ hepatic stellate cells are progenitor cells. *Biochem Biophys Res Commun* 352: 410-417.
129. Berg T, DeLanghe S, Al Alam D, Utley S, Estrada J, et al. (2010) β -catenin regulates mesenchymal progenitor cell differentiation during hepatogenesis. *J Surg Res* 164: 276-285.
130. Castilho-Fernandes A, de Almeida DC, Fontes AM, Melo FU, Picanço-Castro V, et al. (2011) Human hepatic stellate cell line (LX-2) exhibits characteristics of bone marrow-derived mesenchymal stem cells. *Exp Mol Pathol* 91: 664-672.
131. Fausto N (2000) Liver regeneration. *J Hepatol* 32: 19-31.
132. Michalopoulos GK, DeFrances MC (1997) Liver regeneration. *Science* 276: 60-66.
133. Ross MA, Sander CM, Kleeb TB, Watkins SC, Stolz DB (2001) Spatiotemporal expression of angiogenesis growth factor receptors during the revascularization of regenerating rat liver. *Hepatology* 34: 1135-1148.
134. Stolz DB, Ross MA, Salem HM, Mars WM, Michalopoulos GK, et al. (1999) Cationic colloidal silica membrane perturbation as a means of examining changes at the sinusoidal surface during liver regeneration. *Am J Pathol* 155: 1487-1498.
135. Wack KE, Ross MA, Zegar V, Sysko LR, Watkins SC, et al. (2001) Sinusoidal ultrastructure evaluated during the revascularization of regenerating rat liver. *Hepatology* 33: 363-378.
136. Issa R, Zhou X, Constandinou CM, Fallowfield J, Millward-Sadler H, et al. (2004) Spontaneous recovery from micronodular cirrhosis: evidence for incomplete resolution associated with matrix cross-linking. *Gastroenterology* 126: 1795-1808.
137. Forbes SJ, Russo FP, Rey V, Burra P, Rugge M, et al. (2004) A significant proportion of myofibroblasts are of bone marrow origin in human liver fibrosis. *Gastroenterology* 126: 955-963.