

Adipose Tissue Hypoxia: Effects on Metabolism and Insulin Sensitivity

Emily A Lounsbury¹, Feng-Qi Zhao² and Karen M Lounsbury^{1*}

¹Department of Pharmacology, University of Vermont, Burlington, VT 05405, USA

²Department Animal and Veterinary Sciences, University of Vermont, Burlington, VT 05405, USA

*Corresponding author: Karen M. Lounsbury, Professor of Pharmacology, 89 Beaumont Avenue, University of Vermont, Burlington, VT 05405, USA, Tel: 8026561319; E-mail: karen.lounsbury@uvm.edu

Received date: Feb 27, 2016; Accepted date: Mar 01, 2016; Published date: Mar 10, 2016

Copyright: © 2016 Lounsbury EA, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Obesity is associated with an increased risk of insulin resistance and type 2 diabetes. These risks have been correlated with a chronic inflammatory response of the adipose tissue that affects glucose and lipid metabolism. Hypoxia, stemming from inadequate adipose tissue blood flow in obesity, has been shown to be an important contributor to this inflammatory response. The response to hypoxia includes changes in gene expression that affect glucose uptake, glycolysis, lipid metabolism, inflammation, and angiogenesis. This review describes the effects of obesity on chronic and intermittent hypoxia in white adipose tissue and highlights the resulting effects on gene expression and cell metabolism that can affect the onset of systemic insulin resistance and the development of chronic metabolic dysfunction in type 2 diabetes.

Keywords: Obesity; Glycolysis; Metabolism; Hypoxia; Insulin

Introduction

Adipose tissue is known to be an important endocrine organ that communicates metabolic information to the brain and other tissues. Peripheral insulin resistance and inflammation are evolutionarily conserved responses to preserve glucose supply and eliminate harmful stimuli during localized or systemic stress events. It is becoming increasingly clear that the connection between obesity and the development of diabetes includes signaling by the adipose tissue itself, particularly white adipose tissue. Several lines of evidence support the concept that the changes in adipose tissue signaling are linked to hypoxia-mediated changes in the expression of adipokines and inflammatory cytokines reviewed by Trayhurn [1]. These changes cause a disruption in normal glucose and lipid metabolism, and the expression of inflammatory cytokines stimulates macrophage infiltration, which promotes a chronic inflammatory condition within the tissue. The basis for these changes is discussed along with future possibilities of targeting the hypoxia-mediated inflammatory response as a strategy towards preventing the onset of obesity-related type 2 diabetes.

Adipose hypoxia and transcriptional responses

The accumulation of white adipose tissue that occurs due to obesity has been shown to result in chronic adipose tissue hypoxia. The adipose cell response to hypoxia parallels that seen for other cell types, however the response is magnified because obese adipose tissue exhibits on average a chronic 3-fold lower oxygen level than lean adipose tissue (pO₂ 15 vs. 45 mm Hg) [2,3]. As the adipose tissue expands, it is unable to compensate for the tissue oxygenation needs and becomes chronically hypoxic.

The adipose tissue response to hypoxia includes two major transcription factor pathways, hypoxia-inducible factor (HIF) and nuclear factor κB (NFκB). The HIF-1α (and less studied HIF-2α)

subunits act as oxygen sensors. In normoxia, the HIF-1α subunit is rapidly degraded due to oxygen-dependent proline hydroxylation. The hydroxyl group is bound by the E3 ubiquitin ligase, von Hippel-Lindau, which then targets the protein for ubiquitination and degradation by the proteasome. This fundamental regulation by degradation allows a rapid increase in HIF-α protein levels when oxygen levels decline, preventing hydroxylation [4]. Stabilized HIF-1α can then interact with HIF-1β in the nucleus to stimulate the expression of adipocyte genes which improves glucose utilization (i.e., GLUT1, lactate dehydrogenase), initiates hormone signals (i.e., leptin), and stimulates angiogenesis (i.e., vascular endothelial growth factor (VEGF)). Expression of some genes is inhibited by HIF-1, most notably the anti-inflammatory hormone adiponectin [5].

The NFκB transcription factor pathway is also central to the hypoxia response in adipose tissue. NFκB is held inactive in the cytoplasm by binding the inhibitory protein IκB. NFκB becomes active through several stress-mediated pathways that lead to the degradation of IκB. Several theories exist for the mechanism by which hypoxia activates NFκB, but they all include eventual activation of the IκB kinase that leads to degradation of IκB [6]. In cultured adipocytes, hypoxia leads to NFκB activation and an increase in the expression of pro-inflammatory cytokines including tumor necrosis factor-α (TNF-α), interleukin-6 (IL-6) and monocyte chemo-attractant protein-1 (MCP-1) [7]. These inflammatory signals lead to localized changes in extracellular matrix, vascular permeability, and recruitment of inflammatory cells, especially macrophages. This type of response is seen in many chronic inflammatory pathologies including fibrotic diseases and cancer.

In addition to chronic adipose tissue hypoxia, obese patients are also more likely to suffer from intermittent hypoxia due to sleep apnea, and it is often difficult to determine whether the link to metabolic disorders is due to the commonality of obesity, or due to the intermittent hypoxia. Specific effects of intermittent hypoxia on adipose tissue gene expression in animal models have shown significant changes in metabolic and oxidative stress pathways [8]. Unlike chronic hypoxia,

the overall effect of intermittent hypoxia on gene expression is predominated by oxidative stress and NFκB-regulated responses rather than HIF-mediated gene expression, thus inflammatory cytokines are preferentially produced rather than proteins required for metabolic adaptation.

Effects of adipose hypoxia on angiogenesis and inflammation

With obesity, the white adipose tissue hypoxia leads to transcriptional responses that result in changes in the expression and secretion of adipokines as well as angiogenic and inflammatory mediators. As described above, hypoxia promotes the expression and release of leptin, VEGF, IL-6, and MCP-1, whereas adiponectin production is reduced. Expression and secretion of VEGF affects endothelial cells within local vascular beds resulting in increased permeability and migration. In the context of other vascular growth factors such as platelet-derived growth factor (PDGF) and basic fibroblast growth factor (bFGF), the appropriate VEGF response leads to functional angiogenesis that improves the oxygen supply [4]. Unfortunately, adipose hypoxia in obesity and in many pathologies such as peripheral vascular disease and cancer, the VEGF response leads to leaky vessels and exacerbates local inflammation rather than improving oxygenation [9]. This combination along with the production of the inflammatory cytokine IL-6 and the monocyte-attractant MCP-1 results in edema and a buildup of immune cells, including macrophages, T-lymphocytes, B lymphocytes, natural killer cells, and neutrophils to the localized site [10].

An important question is whether the dysfunctional angiogenesis and inflammation have a causal relationship with insulin resistance. Although obese animal models exhibit reduced adipose tissue blood flow, non-obese insulin resistant animals show increased vascularization, suggesting insulin resistance itself does not directly reduce adipose tissue blood flow [11,12]. Instead, the evidence suggests that obesity leads to chronic hypoxia and inadequate angiogenesis that results in inflammation and contributes to the development of insulin resistance.

This model is supported by recent studies of adipose hypoxia in obese patients. Obese patients exhibit lower adipose tissue oxygenation than lean subjects, but the level of hypoxia is not affected by their insulin resistance status [13]. Patients with insulin resistance also exhibit higher plasma levels of adiponectin and increased HIF-1α and VEGF levels in adipose tissue biopsies, suggesting that insulin resistance itself does not exacerbate the hypoxia-mediated inflammatory response but that adipose tissue dysfunction may promote metabolic disease. Furthermore, in adipocytes isolated from type 2 diabetic patients, administration of an NFκB decoy molecule prevents excess NFκB activity and reduced insulin resistance, suggesting that decreasing the inflammatory response that results from adipose hypoxia could provide therapeutic benefit to the insulin resistance that results from obesity [14].

Effects of adipose hypoxia on glucose metabolism and insulin sensitivity

The development of hypoxia in white adipose tissue results in physiological adaptation in adipocytes, including changes in glucose metabolism. Increasing glucose uptake is the first line of defense against hypoxia in most of cells [4,15], including adipocytes. Hypoxia induced glucose uptake has been observed in human subcutaneous preadipocytes [16] and 3T3-L1 adipocytes [17] and can be blocked by

cytochalasin B, an inhibitor of the facilitative glucose transporters (GLUT) [16]. This increase is primarily mediated by the increase in expression of GLUT1 [16,18,19], a ubiquitous GLUT responsible for basal glucose uptake by most of cells, but not GLUT4 and GLUT8 whose expression is reduced by prolonged hypoxia in contrast to GLUT1 [18,20]. There is also a possibility that hypoxia may induce GLUT1 subcellular translocation to the plasma membrane to enhance glucose uptake in adipocytes [17].

In addition to the increase in glucose uptake, another rapid cell response to hypoxia is to induce a metabolic switch from aerobic to anaerobic metabolism. Increased expression of genes encoding enzymes involved in glycolysis, including hexokinase 1 and 2 (HK1 and HK2), glucose-6-phosphate isomerase (GPI), phosphofructokinase (PFK), and aldolase C (ALDOC), has been shown in hypoxic adipocytes by both microarray and proteomic analyses [19,21,22]. More importantly, the production of the major end product of anaerobic glycolysis, lactate, is increased in adipocytes under hypoxia [23,24]. Consistently, the white adipose tissue of obese animals has higher levels of lactate than lean animals *in vivo* [25]. The increase in lactate production may have physiological significance because emerging evidence suggests that lactate is a signaling molecule in many cells. For instance, lactate can stimulate inflammation in macrophages and inhibit lipolysis in adipocytes [26,27]. Thus, the increased lactate under hypoxia may contribute to adipose inflammation and adipose tissue mass increase and obesity.

The metabolic switch toward anaerobic glycolysis leads to less glucose being oxidized through the citric acid cycle to produce ATP in adipocytes during hypoxia. Indeed, microarray analysis has shown a down regulation of a number of genes expressing proteins involved in mitochondrial metabolism and oxidative phosphorylation, such as cytochrome b (CTYB), cytochrome c oxidase subunit Va (COX5A), and ATP synthase, in human adipocytes in response to hypoxia [19]. In addition, reduced ATP production, mitochondrial membrane potential, and NADH dehydrogenase activity have been seen in 3T3-L1 adipocytes after exposed to hypoxia or incubated with hypoxia mimetics CoCl₂ [28].

Many of above changes in glucose metabolism in adipocytes under hypoxia are probably primarily mediated by HIFs as many genes involved in glucose metabolism, such as GLUT1, HKs, lactate dehydrogenase A (LDHA), and pyruvate dehydrogenase kinase 1 (PDK1), are well known direct targets of HIF-1 transcriptional regulation [4,15]. However, hypoxia induced changes in insulin and leptin signaling may also be involved.

Insulin is the primary hormone in regulating glucose homeostasis. In adipocytes, in addition to its role to stimulate glucose uptake, insulin stimulates glucose utilization by stimulating glucose oxidation and synthesis of glycerol-phosphate from glucose for triacylglycerol synthesis [29-32]. Increasing evidence has shown that adipose tissue hypoxia inhibits insulin signaling and blocks insulin-stimulated glucose uptake in adipocytes. The adipose tissue of obese mice has decreased insulin receptor-β and insulin receptor substrate-1, consistently with the changes observed in 3T3-L1 adipocytes after hypoxia treatment [33]. In addition, hypoxia reduces phosphorylation of insulin receptor, Akt, and AS160 in 3T3-L1 adipocytes in coupled with blocking insulin-stimulated glucose transport [34,35]. These changes are dependent on the HIF expression because overexpression or down regulation of HIF-1α or HIF-2α can mimic or inhibit the effects of hypoxia, respectively [34]. This evidence suggests that

hypoxia induces insulin resistance in adipose cells, contributing to the metabolic changes in these cells.

Leptin is a hormone primarily produced by adipose tissue and plays an important role in the regulation of body energy balance, mainly food intake and energy expenditure [36,37]. It plays a critical role in the regulation of glucose homeostasis, as demonstrated by the hyperglycemia phenotype in leptin gene knockout mice (ob/ob and db/db) [36]. The specific action of leptin on adipocytes is likely mediated by its inhibition of insulin action, as observed in white adipose tissue [37]. In adipocytes, hypoxia induces leptin expression [18,23]. This increase of leptin signaling may play a role in the above mentioned inhibition of insulin signaling by hypoxia.

The increase of leptin signaling and inhibition of insulin signaling and insulin-stimulated glucose uptake by hypoxia in adipocytes imply that hypoxia plays a role in developing type 2 diabetes in obese subjects. Further effects of hypoxia on insulin sensitivity in adipose tissue also include: 1) the production of several insulin-sensitizing adipokines is reduced under hypoxia [38,39], and 2) expression of GLUT4 is markedly reduced by hypoxia in adipose cells [18]. Consistently, knockout of HIF1 α in adipose tissue reduces fat formation and ameliorates insulin resistance in mice [40]. In humans, the more severe the obesity, the greater the relative risk of developing type 2 diabetes. Thus, efforts to control adipose tissue hypoxia or the resulting transcriptional response could provide preventative or treatment options for obesity-related pathologies such as type 2 diabetes.

Effects of hypoxia on lipid metabolism

In normal white adipose tissue, lipolysis is stimulated when the insulin level is low and epinephrine levels are high. Epinephrine binds to cell membrane beta adrenergic receptors activating adenyl cyclase and generating cAMP. The subsequent activation of protein kinase A leads to phosphorylation of hormone sensitive lipase which facilitates its interaction with the lipid droplet and the formation of free fatty acids and monoacylglycerol [41]. Due to the accumulation of free fatty acids, glucose utilization decreases because free fatty acids block the signal for more glucose uptake [42].

This normal regulation of lipid metabolism in adipocytes is disrupted in obesity due to the effects of hypoxia. In obese mice with measurably significant adipose hypoxia (40-50% reduced PO₂), free fatty acid uptake is reduced and lipolysis is increased. These effects on lipid metabolism are also mimicked by exposure of cultured adipose cells to a hypoxic environment [35]. Many of these changes are attributed to NF κ B-mediated TNF- α production that leads to down regulation of the transcription factor peroxisome proliferator-activated receptor- γ (PPAR- γ). PPAR- γ is necessary for the induction of genes that control fatty acid synthesis and sequestration of triglycerides in lipid droplets including phosphoenolpyruvate carboxykinase, fatty acid synthase, and perilipin among others [43]. The hypoxia response of adipocytes also leads to insulin resistance which prevents its inhibitory effect on lipolysis and an increase in free fatty acid levels in the blood stream. The combination of tissue insulin resistance, increased lipolysis and excess leptin production contribute to an increased risk of systemic insulin resistance, lipid-mediated pathologies and the development of type 2 diabetes.

Directions of research and potential for therapy

It is clear that adipose tissue hypoxia results in multiple changes in lipid and glucose metabolism that can contribute to metabolic dysfunction that promotes or exacerbates type 2 diabetes (Figure 1). The findings related to hypoxia-mediated inflammation and metabolic disruption in adipose tissue have moved the research field towards efforts to correct these fundamental changes through possible therapeutic intervention.

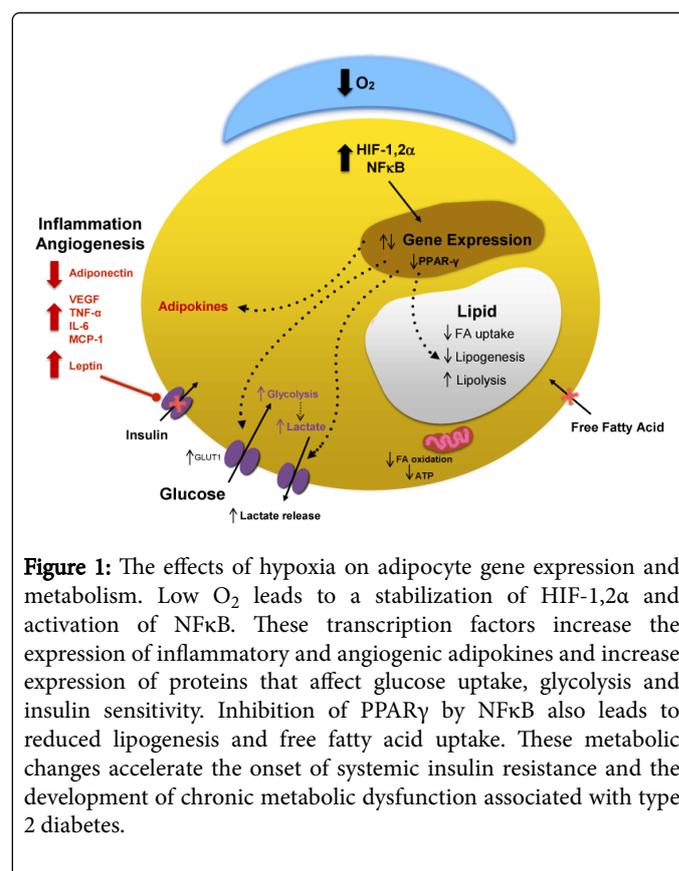


Figure 1: The effects of hypoxia on adipocyte gene expression and metabolism. Low O₂ leads to a stabilization of HIF-1,2 α and activation of NF κ B. These transcription factors increase the expression of inflammatory and angiogenic adipokines and increase expression of proteins that affect glucose uptake, glycolysis and insulin sensitivity. Inhibition of PPAR γ by NF κ B also leads to reduced lipogenesis and free fatty acid uptake. These metabolic changes accelerate the onset of systemic insulin resistance and the development of chronic metabolic dysfunction associated with type 2 diabetes.

Many questions remain including the benefit of accelerating vascularization of adipose tissue or perhaps preventing the hypoxic response. Studies using transgenic mouse models have shown conflicting evidence which leads to even more questions. For example, overexpression of VEGF in adipose tissue of animals fed a high-fat diet promotes an increase in vascularization, a decrease in hypoxia and normalization of metabolic indicators [44], and knockdown of HIF-2 α in adipose tissue worsens the hypoxia and inflammation. These findings suggest that increasing expression of angiogenic factors to stimulate vascularization will reduce the inflammatory effects of adipose hypoxia. Alternatively, knocking down HIF-1 α in a mouse model improves the insulin sensitivity and decreases adiposity in obese mice [45]. Thus, preventing the overall hypoxic transcriptional response may be more important for reducing the inflammation and growth response of adipose tissue. Realistically, there are no current therapies that directly target HIF-1 α , and anti-VEGF therapies would likely make the hypoxic situation worse.

Efforts to target the resulting inflammatory response to hypoxia have also been explored. These approaches include targeting NF κ B (salicylates, salsalate), TNF- α (entanercept, infliximab, adalimumab),

IL-1 β (anakinra, canakinumab) and IL-6 (tocilizumab) [46]. Using agents to modulate NF κ B, such as resveratrol have also been proposed [47]. These therapies have shown promise in animal models and have successfully reduced inflammatory markers in patient studies; however, none of the anti-inflammatory therapies have had significant benefit towards improving insulin sensitivity and glucose homeostasis in type 2 diabetic patients [48]. Although there remain many questions related to the appropriate therapeutic target in treating adipose tissue inflammation in obesity, a better understanding of the complex interactions between angiogenic and inflammatory signaling pathways may improve the efforts to reduce chronic adipose hypoxia and prevent the resulting effects on insulin sensitivity and the development of type 2 diabetes.

References

1. Trayhurn P (2014) Hypoxia and adipocyte physiology: implications for adipose tissue dysfunction in obesity. *Annu Rev Nutr* 34: 207-236.
2. Rausch ME, Weisberg S, Vardhana P, Tortoriello DV (2008) Obesity in C57BL/6J mice is characterized by adipose tissue hypoxia and cytotoxic T-cell infiltration. *Int J Obes (Lond)* 32: 451-463.
3. Sun K, Halberg N, Khan M, Magalang UJ, Scherer PE (2013) Selective inhibition of hypoxia-inducible factor 1 α ameliorates adipose tissue dysfunction. *Mol Cell Biol* 33: 904-917.
4. Cassavaugh J, Lounsbury KM (2011) Hypoxia-mediated biological control. *J Cell Biochem* 112: 735-744.
5. Wang B, Wood IS, Trayhurn P (2007) Dysregulation of the expression and secretion of inflammation-related adipokines by hypoxia in human adipocytes. *Euro J Physiol* 455: 479-492.
6. Melvin A, Mudie S, Rocha S (2011) Mechanism of hypoxia-induced NF κ B. *Cell Cycle* 10: 879-882.
7. Ye J, Gao Z, Yin J, He Q (2007b) Hypoxia is a potential risk factor for chronic inflammation and adiponectin reduction in adipose tissue of ob/ob and dietary obese mice. *Am J Physiol Endocrinol Metab* 293: E1118-E1128.
8. Gharib SA, Khalyfa A, Abdelkarim A, Ramesh V, Buazza M, et al. (2011) Intermittent hypoxia activates temporally coordinated transcriptional programs in visceral adipose tissue. *J Mol Med* 90: 435-445.
9. Carmeliet P, Jain RK (2011) Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat Rev Drug Discov* 10: 417-427.
10. Lee J (2013) Adipose tissue macrophages in the development of obesity-induced inflammation, insulin resistance and type 2 Diabetes. *Archives of Pharmacol Research* 36: 208-222.
11. Kampf C, Bodin B, Kallskog O, Carlsson C, Jansson L (2005) Marked increase in white adipose tissue blood perfusion in the type 2 diabetic GK rat. *Diabetes* 54: 2620-2627.
12. Ye J (2011) Adipose tissue vascularization: its role in chronic inflammation. *Curr Diab Rep* 11: 203-210.
13. Lawler HM, Underkofler CM, Kern PA, Erickson C, Bredbeck B, et al. (2016) Adipose Tissue Hypoxia, Inflammation and Fibrosis in Obese Insulin Sensitive and Obese Insulin Resistant Subjects. *J Clin Endocrinol Metab*, 0: 2015-4125.
14. Zhong L, Luo Y, Huang C, Liu L (2011) Effect of NF-kappaB decoy on insulin resistance of adipocytes from patients with type 2 diabetes mellitus. *Diabetes Metab* 37: 520-526.
15. Shao Y, Zhao FQ (2014) Emerging evidence of the physiological role of hypoxia in mammary development and lactation. *J Anim Sci Biotechnol* 5: 9.
16. Wood IS, Wang B, Lorente-Cebrian S, Trayhurn P (2007) Hypoxia increases expression of selective facilitative glucose transporters (GLUT) and 2-deoxy-D-glucose uptake in human adipocytes. *Biochem Biophys Res Commun* 361: 468-473.
17. Bashan N, Burdett E, Gumà A, Sargeant R, Tumiati L, et al. (1993) Mechanisms of adaptation of glucose transporters to changes in the oxidative chain of muscle and fat cells. *Am J Physiol* 264: C430-440.
18. Grosfeld A, Zilberfarb V, Turban S, André J, Guerre-Millo M, et al. (2002) Hypoxia increases leptin expression in human PAZ6 adipose cells. *Diabetologia* 45: 527-530.
19. Mazzatti D, Lim FL, O'Hara A, Wood IS, Trayhurn P (2012) A microarray analysis of the hypoxia-induced modulation of gene expression in human adipocytes. *Arch Physiol Biochem* 118: 112-120.
20. Scheepers A, Doege H, Joost HG, Schürmann A (2001) Mouse GLUT8: genomic organization and regulation of expression in 3T3-L1 adipocytes by glucose. *Biochem Biophys Res Commun* 288: 969-974.
21. Choi S, Cho K, Kim J, Yea K, Park G, et al. (2009) Comparative proteome analysis using amine-reactive isobaric tagging reagents coupled with 2D LC/MS/MS in 3T3-L1 adipocytes following hypoxia or normoxia. *Biochem Biophys Res Commun* 383: 135-140.
22. Geiger K, Leiherer A, Muendlein A, Stark N, Geller-Rhomberg S, et al. (2011) Identification of hypoxia-induced genes in human SGBS adipocytes by microarray analysis. *PLoS One* 6: e26465.
23. Lolmède K, Durand de Saint Front V, Galitzky J, Lafontan M, Bouloumié A (2003) Effects of hypoxia on the expression of proangiogenic factors in differentiated 3T3-F442A adipocytes. *Int J Obes Relat Metab Disord* 27: 1187-1195.
24. Pérez de Heredia F, Wood IS, Trayhurn P (2010) Hypoxia stimulates lactate release and modulates monocarboxylate transporter (MCT1, MCT2, and MCT4) expression in human adipocytes. *Pflugers Arch* 459: 509-518.
25. Hosogai N, Fukuhara A, Oshima K, Miyata Y, Tanaka S, et al. (2007) Adipose tissue hypoxia in obesity and its impact on adipocytokine dysregulation. *Diabetes* 56: 901-911.
26. Liu C, Wu J, Zhu J, Kuei C, Yu J, et al. (2009) Lactate inhibits lipolysis in fat cells through activation of an orphan G-protein-coupled receptor, GPR81. *J Biol Chem* 284: 2811-2822.
27. Samuvel DJ, Sundararaj KP, Nareika A, Lopes-Virella MF, Huang Y (2009) Lactate boosts TLR4 signaling and NF-kappaB pathway-mediated gene transcription in macrophages via monocarboxylate transporters and MD-2 up-regulation. *J Immunol* 182: 2476-2484.
28. Jang MK, Son Y, Jung MH (2013) ATF3 plays a role in adipocyte hypoxia-mediated mitochondria dysfunction in obesity. *Biochem Biophys Res Commun* 431: 421-427.
29. Baquer NZ, Sinclair M, Kunjara S, Yadav UC, McLean P (2003) Regulation of glucose utilization and lipogenesis in adipose tissue of diabetic and fat fed animals: effects of insulin and manganese. *J Biosci* 28: 215-221.
30. Ebner S, Burnol AF, Ferre P, de Saintaurin MA, Girard J (1987) Effects of insulin and norepinephrine on glucose transport and metabolism in rat brown adipocytes. Potentiation by insulin of norepinephrine-induced glucose oxidation. *Eur J Biochem* 170: 469-474.
31. Ferré P, Burnol AF, Leturque A, Terretaz J, Penicaud L, et al. (1986) Glucose utilization *in vivo* and insulin-sensitivity of rat brown adipose tissue in various physiological and pathological conditions. *Biochem J* 233: 249-252.
32. Saltiel AR, Sorbara-Cazan LR (1987) Inositol glycan mimics the action of insulin on glucose utilization in rat adipocytes. *Biochem Biophys Res Commun* 149: 1084-1092.
33. Yin J, Gao Z, He Q, Zhou D, Guo Z, et al. (2009) Role of hypoxia in obesity-induced disorders of glucose and lipid metabolism in adipose tissue. *Am J Physiol Endocrinol Metab* 296: E333-342.
34. Regazzetti C, Peraldi P, Grémeaux T, Najem-Lendom R, Ben-Sahra I, et al. (2009) Hypoxia decreases insulin signaling pathways in adipocytes. *Diabetes* 58: 95-103.
35. Yin J, Gao Z, He Q, Zhou D, Guo Z, et al. (2009) Role of hypoxia in obesity-induced disorders of glucose and lipid metabolism in adipose tissue. *Am J Physiol Endocrinol Metab* 296: E333-342.

36. Denroche HC, Huynh FK, Kieffer TJ (2012) The role of leptin in glucose homeostasis. *J Diabetes Investig* 3: 115-129.
37. Harris RB (2014) Direct and indirect effects of leptin on adipocyte metabolism. *Biochim Biophys Acta* 1842: 414-423.
38. Yamauchi T, Kamon J, Waki H, Terauchi Y, Kubota N, et al. (2001) The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nat Med* 7: 941-946.
39. Ye J, Gao Z, Yin J, He Q (2007) Hypoxia is a potential risk factor for chronic inflammation and adiponectin reduction in adipose tissue of ob/ob and dietary obese mice. *Am J Physiol Endocrinol Metab* 293: E1118-1128.
40. Jiang C, Qu A, Matsubara T, Chanturiya T, Jou W, et al. (2011) Disruption of hypoxia-inducible factor 1 in adipocytes improves insulin sensitivity and decreases adiposity in high-fat diet-fed mice. *Diabetes* 60: 2484-2495.
41. Morigny P, Houssier M, Mouisel E, Langin D (2015) Adipocyte lipolysis and insulin resistance. *Biochimie* .
42. White MF (2002) IRS proteins and the common path to diabetes. *Am J Physiol Endocrinol Metab* 283: E413-422.
43. Guilherme A, Virbasius JV, Puri V, Czech MP (2008) Adipocyte dysfunctions linking obesity to insulin resistance and type 2 diabetes. *Nat Rev Mol Cell Biol* 9: 367-377.
44. Sung HK, Doh KO, Son Joe E, Park Jin G, Bae Y, et al. (2013) Adipose Vascular Endothelial Growth Factor Regulates Metabolic Homeostasis through Angiogenesis. *Cell Metabolism* 17: 61-72.
45. Jiang C, Qu A, Matsubara T, Chanturiya T, Jou W, et al. (2011) Disruption of hypoxia-inducible factor 1 in adipocytes improves insulin sensitivity and decreases adiposity in high-fat diet-fed mice. *Diabetes* 60: 2484-2495.
46. Esser N, Paquot N, Scheen AJ (2015) Anti-inflammatory agents to treat or prevent type 2 diabetes, metabolic syndrome and cardiovascular disease. *Expert Opin Investig Drugs* 24: 283-307.
47. Carreras A, Zhang SXL, Almendros I, Wang Y, Peris E, et al. (2015) Resveratrol Attenuates Intermittent Hypoxia-Induced Macrophage Migration to Visceral White Adipose Tissue and Insulin Resistance in Male Mice. *Endocrinology* 156: 437-443.
48. Ye J, McGuinness OP (2013) Inflammation during obesity is not all bad: evidence from animal and human studies. *Am J Physiol Endocrinol Metab* 304: E466-477.