

EFFECT OF DICLOFENAC SODIUM AT LOW CONCENTRATION LEVEL ON THE RATE OF ORTHODONTIC TOOTH MOVEMENT IN RAT

¹ Kurunji Kumaran N	¹ Reader
² Rajasigamani K	² Professor and Head
³ Sethupathy S	³ Professor
⁴ Madhavan Nirmal S	⁴ Professor
⁵ Venkataramana	⁵ Reader

^{1,2,5} Department Of Orthodontics and Dentofacial Orthopedics, ³Department Of Biochemistry, ⁴Department Of Oral Pathology, Rajah Muthiah Dental College and Hospital, Annamalai University, Annamalai Nagar, Tamilnadu, India 608402.

ABSTRACT: The purpose was to evaluate the influence of a NSAID, Diclofenac Sodium (DFS) on the tissue reaction related to orthodontic tooth movement (OTM). **Methods:** 27 adult male wistar rats were divided into 3 Groups of 9 each. Orthodontic closed coil spring was placed between rat incisor and molar to produce a 50 gm of force. The experimental Groups 1 and 2 received orthodontic force and either DFS of 0.0025mg/0.05ml or 0.05ml of saline, Group 3 received only orthodontic force and served as a control group. At the end of day 5, 10 and 15 the animals were sacrificed and histological examination was performed. **Results:** Student's t test showed a statistically significant difference between the control Groups and experimental Groups. Group 1 showed a statistically significant reduction in osteoclastic cell count at day 5, 10 and 15 when compared to Group 2 and Group 3. **Conclusion:** The results indicate that the Diclofenac Sodium even at low concentration level diminishes the number of osteoclasts probably by inhibiting the secretion of prostaglandins there by reducing the OTM.

KEYWORDS: NSAID, Diclofenac Sodium, Orthodontic Tooth Movement (OTM).

INTRODUCTION

Even though the biochemical mediators that initiate orthodontic tooth movement were not fully understood¹, the role of prostaglandins and other mediators like cyclic amp, vitamin D, thromboxanes, cytokines, interleukins, leucotrienes have been extensively studied by various authors²⁻⁹. As a milestone in 1984 Yamasaki et al¹⁰ administered prostaglandin₁ locally and showed an increase in rate of tooth movement in human subjects.

In 1991 vane and coworkers¹¹ made the landmark observation that Aspirin and some NSAIDS blocked prostaglandins synthesis. Chumbley and Tuncay¹² showed a significant reduction in tooth movement in mongrel cats' canine retraction experiment using Indomethacin (an Aspirin like drug). The hypothesis that those drugs which suppress production of prostaglandins should reduce the rate of tooth movement was well proved by previous studies using NSAIDS like aspirin, Acetaminophen, Meloxicam, Celecoxib, Parecoxib, Rofecoxib, Indomethacin, Ibuprofen and Flurbiprofen in miniature pigs, rabbits, beagles and rats.¹³⁻¹⁹

Pain killers especially commonly available NSAIDS have been prescribed by orthodontists to those patients receiving orthodontic treatment. Acetaminophen which acts centrally to block pain sensation rather than those work peripherally remains the only choice of pain relieving medication during orthodontic treatment for decades^{20,14}. In one study done by Felix de Carlos et al²¹ showed a significant reduction in rate of tooth movement for 50 and 100gms of force using 10mg/kg of body weight of Diclofenac Sodium (DFS). The present study was designed to test the efficacy of local injection of Diclofenac Sodium (DFS) in altering the tissue reaction relation to orthodontic tooth movement in rats at low concentration level

Materials and Methods

Twenty seven male adult albino wistar rats each weighing 250 to 350 gms were received and maintained with 12hr light/dark cycle in central animal house, Annamalai University. The animals were randomly divided into 3 Groups of 9 animals each. They were further divided into 3 sub groups of 3 animals each.

Prior to appliance insertion all animals were color coded according to their groups. The animals body weight were cautiously monitored prior to appliance insertion and then daily till they are sacrificed. The animals were anesthetized using ketamine (44mg/kg bodyweight) and xylazine (2mg/kg bodyweight) ³. The appliance design of this study follows that used by Leiker etal²². A closed coil nickel-titanium spring (Sentalloy, GAC, clr Islip, NY) calibrated to produce a force of 50gms was ligated to the maxillary first molar and incisors (Fig.1). To minimize the appliance dislodgement, all animals were fed with finely ground rat lab pellet ad libitum.

The drugs were delivered locally mesial to the maxillary first molar, once daily at the same time point from the day of appliance incersion till they are sacrificed. Group 1(DFS+OF) received Diclofenac Sodium at the low concentration level of 0.0025mg/0.05ml, Group 2 (S+OF) received saline 0.05ml and Group 3 (OF)received only orthodontic force and no injection, served as control.

Table I explains the animal grouping, drug dosage and their sacrifice regime. All animals well tolerated the experimental procedure. There was no gross reduction in body weight and no appliance dislodgement during the experimental period; hence no animal was excluded from the study. At the end of the experiment all the animals were sacrificed by CO₂ inhalation

Histopathology

After appliance removal Premaxilla was dissected, the specimens placed in 10% formalin immediately. Decalcification was done using 9% formic acid. After wax block preparation the premaxilla were hemi sectioned at coronal, middle and apical third of the molar root level. Each section was again serially sectioned at 4 to 6 µm in the coronal plane. The sections were mounted on glass microscope slides and stained with hematoxylin and eosin. Multinuclear osteoclasts and osteoblasts on the stained sections were counted by two pathologists twice at different times using light microscope.

Table.1. Animal grouping, drug dosage and sacrifice regime.

Groups	Drug dosage	Number of animals sacrificed			Total
		Day5	Day10	Day15	
G1- Diclofenac Sodium and Orthodontic force* (DFS+OF)	0.0025mg/0.05ml	3	3	3	9
G2 -Saline and Orthodontic force* (S+OF)	0.05ml	3	3	3	9
G3-Orthodontic force* (OF)	-----	3	3	3	9
Total					27

(*orthodontic force has been standardized for 50gms)

Table .2. The mean and standard deviation of histological osteoclastic cell count at the three levels coronal, middle, apical root surface for three Groups at three time intervals (5,10,15 days) of animal sacrifice

Groups	5 days			10 days			15 days		
	Coronal psi	Middle psi	Apical psi	Coronal psi	Middle psi	Apical psi	Coronal psi	Middle psi	Apical psi
Group 1	1.71±0.68	1.32±0.48	1.61±0.07	2.70±0.33	1.34±0.55	1.57±0.04	1.89±0.53	1.22±0.21	2.33±0.41
Group 2	3.30±0.74	4.10±0.78	2.20±0.02	4.20±0.12	5.20±0.22	5.03±0.36	6.30±0.11	7.10±0.22	5.22±0.85
Group 3	3.60±0.79	4.40±0.65	2.20±0.33	2.10±0.13	3.90±0.44	2.54±0.05	5.20±0.17	4.33±0.92	2.50±0.23

(Mean ± Standard Deviation)

Results

When the rat molar was moved mesial by the orthodontic force the remodeling phase of alveolar bone was markedly changed. The compression of periodontal ligament and appearance of osteoclasts on the mesial side of the molar root in the experimental Group 1(DFS+OF) at day 5 is shown in **Fig. 2**. The appearance of osteoblasts on the distal side of the molar root in the control Group G2(S+OF) is shown in **Fig 3**.

Table III, Table IV and Table V shows statistically significant reduction in osteoclastic cell count in Group 1(DFS+OF) than Group 2(S+OF) and Group 3 (OF) at day 5, 10, 15 except only at the apical level at day 15 between Group1 (DFS+OF) and Group 3 (OF). But Intra group comparison of Group 1(DFS+OF) coronal, middle and apical root surfaces at day 15 revealed no statistical significance between coronal and apical. Also there was no statistical significant difference between Group 2(S+OF) and Group 3(OF) at day 5. However Group 2(S+OF) showed a significant increase in osteoclastic cell count when compared to Group 3(of) at day 10 and 15. In overall Group 1(DFS+OF) showed statistically significant reduction in cell count throughout the study when compared to Group 2(S+OF) and Group 3(OF).

Statistical method

All the data were subjected to intergroup and intragroup comparison using descriptive statistical analysis and paired student's t- test (SPSS/win-10). Based on the mean value we are able to find out which group is higher. The difference is statistically proved if the t- test is significant. Suppose the "t" value is not significant that indicate there is no difference among groups. ($p < 0.05$). The mean and standard deviation of histological osteoclastic cell count at the three levels that is coronal middle and apical for three groups at three time intervals (5,10,15 days) of animal sacrifice is shown in **Table II**. Bar chart representation of the above results were given in **Graph I**. The data was then subjected to statistical analysis.

Discussion

Orthodontic tooth movement requires remodeling of the alveolar bone²³. A patient's bone resorption potential and timely orthodontic treatment outcome depends on recruitment of mature oestoclast and its precursors, oestoclast differentiation and number of functional oestoclast at the bone, periodontal ligament interfaces²⁴. Interpretation of Tsay et al²⁵ study results indicates that during prolonged periods of tooth movement the replenishment of osteoclasts in the periodontal membrane depends on the viability of hemopoietic organs

Interestingly early experiments done by Kenichi Yamasaki et al^{26,1} suggests a strong local role played by inflammation mediator prostaglandin as a mediator of bone resorption during orthodontic tooth movement,

revealing to a possible hypothesis that local administration of active prostaglandin increasing rate of tooth movement and suppression of local prostaglandin by use of such drugs (NSAID) either a reduction in rate of tooth movement or reduced number of osteoclasts at the experimental sites during experimental tooth movement done in animals^{2,9,27-30}.



Fig.1 After appliance insertion



Fig.2 . Appearance of osteoclast on the compression side

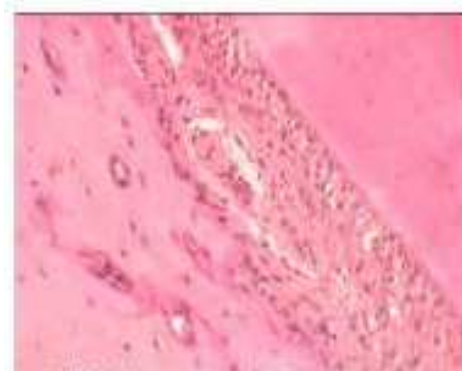


Fig.3 Appearance of osteoblasts on tension side

Table.3 Inter Group comparison for three Groups at day 5

Groups at day 5	Coronal psi		Middle psi		Apical psi	
	Mean	SD	Mean	SD	Mean	SD
Group 1	1.71	0.68	1.32	0.48	1.61	0.07
Group 2	3.30	0.74	4.10	0.78	2.20	0.02
t-value	43.69		7.10		17.30	
P value	0.001 ^S		0.019 ^S		0.003 ^S	
Group 1	1.71	0.68	1.32	0.48	1.61	0.07
Group 3	3.60	0.79	4.40	0.65	2.20	0.33
t-value	4.43		9.08		3.85	
P value	0.047 ^S		0.012 ^S		0.051 ^S	
Group 2	3.30	0.74	4.10	0.78	2.20	0.02
Group 3	3.60	0.79	4.40	0.65	2.20	0.33
t-value	0.68		0.42		0.00	
P value	0.567 ^{NS}		0.716 ^{NS}		1.000 ^{NS}	

S – Significant (p<0.05) NS – Not Significant

The histopathological results of the present study were subjected to statistical analysis using descriptive statistical analysis and paired student's t- test (SPSS-10) revealed that experimental rats which received daily injection of diclofenac sodium and orthodontic force Group 1 (DFS+OF) showed statistically significant reduction in osteoclastic cells at all three levels that is coronal, middle and apical at three time points (5,10,15 days) when compared to those rats received saline and orthodontic force Group2 (S+OF) and only orthodontic force Group 3(OF).

Diclofenac sodium, a member of aryl acetic acid group of NSAIDs is a potent anti-inflammatory drug routinely given for dental treatments and systemic reasons. It interferes with cyclooxygenase pathway and blocks both cox1 and cox2 metabolites. In one study Felix de Carlos²¹ used Diclofenac Sodium 10mg/kg with 50grms or 100grms of force and found significantly inhibited orthodontic tooth

Table.4 Inter Group comparison for three Groups at day 10

Groups at day 10	Coronal psi		Middle psi		Apical psi	
	Mean	SD	Mean	SD	Mean	SD
Group 1	2.70	0.33	1.34	0.55	1.57	0.04
Group 2	4.20	0.12	5.20	0.22	5.03	0.36
t-value	12.14		20.20		16.33	
P value	0.007 ^S		0.002 ^S		0.004 ^S	
Group 1	2.70	0.33	1.34	0.55	1.57	0.04
Group 3	2.10	0.13	3.90	0.44	2.54	0.05
t-value	5.14		40.68		211.74	
P value	0.036 ^S		0.001 ^S		0.001 ^S	
Group 2	4.20	0.12	5.20	0.22	5.03	0.36
Group 3	2.10	0.13	3.90	0.44	2.54	0.05
t-value	303.11		10.14		11.64	
P value	0.001 ^S		0.010 ^S		0.007 ^S	

S – Significant (p<0.05) NS – Not Significant

movement in rats. In the present study we used local injection of (Group 1) . Diclofenac sodium at 0.0025mg/0.05ml with 50gms of force, which is comparatively very low dose to the previous study, also potently reduced the appearance of osteoclasts. Group 2(S+OF) showed a gradual increase in cell count on 5, 10, 15 days. This might be probably due to the daily injection regime which might have altered the local inflammatory environment. But Group 3(OF) which received only orthodontic force showed an increased cell count at day 5 followed by a slight drop in cell count at day 10 then a sudden or gradual increase in cell count which represents the classical 3 phase orthodontic tooth movement described by Burstone.C³¹ in 1962. That is initial phase [displacement of tooth in the periodontal membrane space], lag phase [no or relatively low rate of tooth movement], post lag phase [rate of movement gradually or suddenly increased].

Table.5. Inter Group comparison for three Groups at day 15

Groups at day 15	Coronal psi		Middle psi		Apical psi	
	Mean	SD	Mean	SD	Mean	SD
Group 1	1.89	0.53	1.22	0.21	2.33	0.41
Group 2	6.30	0.11	7.10	0.22	5.22	0.85
t-value	18.38		45.53		6.85	
P value	0.003 ^S		0.001 ^S		0.021 ^S	
Group 1	1.89	0.53	1.22	0.21	2.33	0.41
Group 3	5.20	0.17	4.33	0.92	2.50	0.23
t-value	16.13		4.76		0.84	
P value	0.004 ^S		0.041 ^S		0.491 ^{NS}	
Group 2	6.30	0.11	7.10	0.22	5.22	0.85
Group 3	5.20	0.17	4.33	0.92	2.50	0.23
t-value	31.75		4.56		4.82	
P value	0.001 ^S		0.045 ^S		0.040 ^S	

S – Significant (p<0.05) NS – Not Significant

In overall Diclofenac Sodium is capable of reducing the number of oestoclast at the compression side even at the low concentration level [0.0025mg/ml] when injected locally. In the present study we were not able to achieve the complete elimination of oestoclast, this further validate that not only the cyclooxygenase metabolites play a vital role in OTM, lipooxygenase pathway also contributes a Role for OTM. Since multiple mediators and signaling molecules are involved during inflammatory reactions evoked by orthodontic force, blocking of all mediators and signaling molecules at molecular level is questionable.

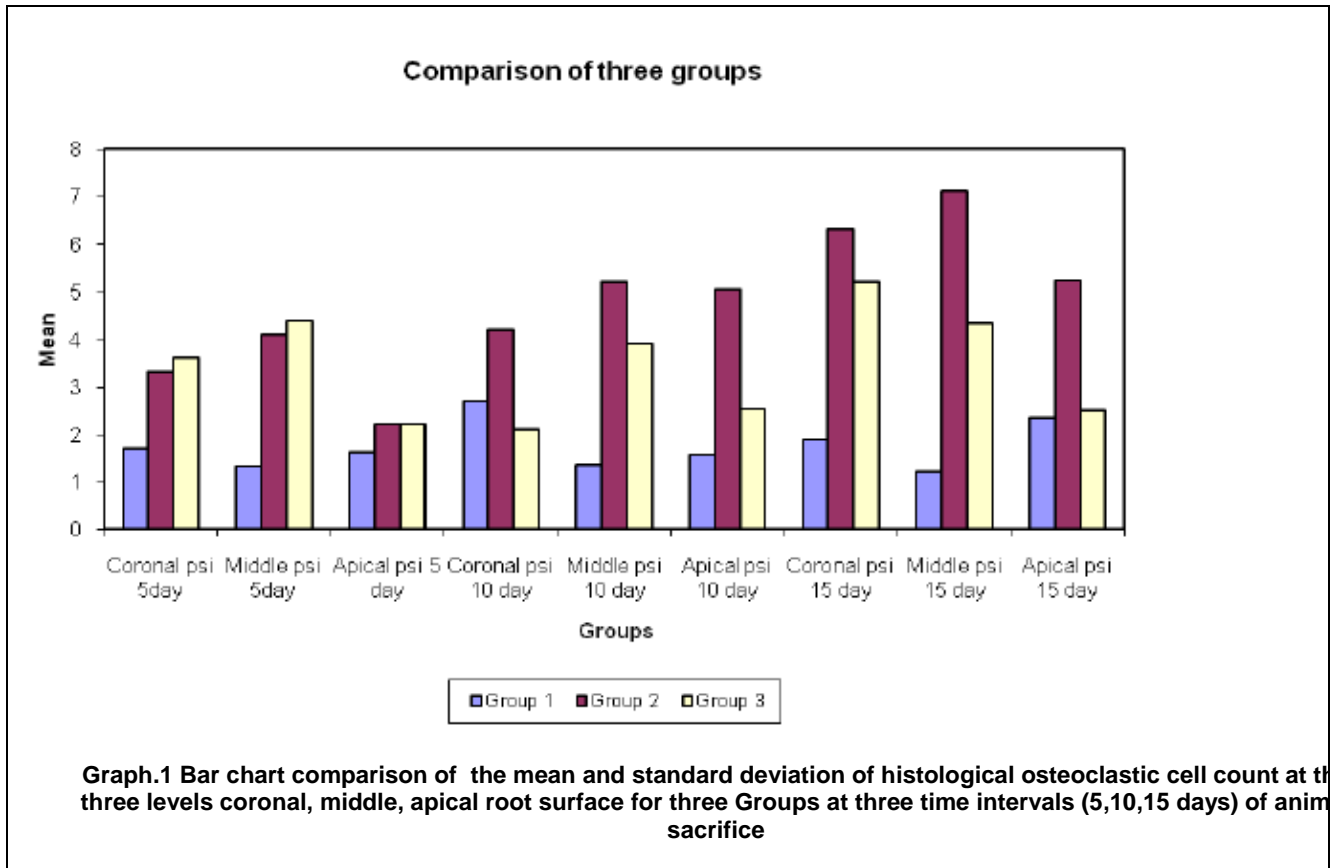
CONCLUSION

The ability of Diclofenac Sodium to interrupt orthodontic tooth movement and reduce the rate of tooth movement was proved in the present study by the expression of reduction in osteoclastic cell numbers .Although systemic

administration of these NSAIDS are contraindicated in the sense of impairing the entire orthodontic treatment process by reducing the rate of tooth movement^{32,33,34}, the daily local injection of Diclofenac Sodium to a particular tooth, especially to the posterior teeth might enhance the anchorage potential. However the present results need to be confirmed in other species including humans. To overcome the daily injection in humans, sustained release transmucosal adhesive strips carrying these drug or the 'IntelliDrug' device³⁵ representing a revolutionary method for delivering drugs for long-term through the buccal mucosa, according to the patient needs, in periods lasting days, weeks or months will be a promising solution in near future.

References

1. Yamasaki K. The role of cyclic amp, calcium, and prostaglandins in the induction of osteoclastic bone resorption associated with experimental tooth movement. J.Dent.Res, August 1983; 62(8);[877-881]. PMID:6306082
<http://dx.doi.org/10.1177/00220345830620080501>
2. Arif Umit Gurton, Erol Akin, Deniz Sagdic, and Huseyin Olmez. Effects Of Pgi2and Txa2 Analogs And Inhibitors In Orthodontic Tooth Movement. Angle Orthod, 2004: 74(4); 526-532.
3. Abbas H Mohammed, Dimitris N Tatakis and Rosemary Dziak. Leukotrienes in orthodontic tooth movement. Am J Orthod Dentofacial Orthop, 1989; Mar 95; 231-237.
4. Laura R Iwasaki, Larry D Crouch, Albert Tutor, Scott Gibson, Navin Hukmani, David B Marx, Jeffrey C Nickel. Tooth movement and cytokines in gingival crevicular fluid and whole blood in growing and adult subjects. Am J Orthod Dentofacial Orthop, October 2005;128(4);483-491.
5. Kee-Joon Lee, Young-Chel Park, Hyung-Seog Yu, Seong-Ho Choi, Yun-Jung Yoo. Effects of continuous and interrupted orthodontic force on interleukin-1 β and prostaglandin E2 production in gingival crevicular fluid. Am J Orthod Dentofacial Orthop, 2004,125(2);168-177.
6. Monte K. Collins and Peter M. Sinclair. The local use of vitamin D to increase the rate of orthodontic tooth movement. Am J Orthod Dentofacial Orthop, 1988 Oct94;278-284. PMID:1506515
7. Takano T-Yamamoto, Kawakami M., Yamashiro T. Effect of age on the rate of tooth movement in combination with local use Of 1, 25 (Oh) 2d 3 and mechanical forces in the rat. J.Dent.Res, August 1992;71(8);1487-1492.
<http://dx.doi.org/10.1177/00220345920710080501>
8. Burcu Balo Tuncer, Nurdan A-zmeriAŞ, Cumhuri Tuncer, dil Teoman, Burcu A-ak Ic, Ay egA¼l YA¼cel, Reha Alpar, and KA¼ksal Balo .2005: Levels of interleukin-8 during tooth movement. AngleOrthod,75(4);631-636].
9. William G. Grieve, Georgia K Johnson, Robert N Moore, Richard A Reinhardt, Linda M DuBois.



10. Prostaglandin E (PGE) and interleukin-1 β (IL-1 β) levels in gingival crevicular fluid during human orthodontic tooth movement. *Am J Orthod Dentofacial Orthop*, April 1994; 105(4):369-374.
11. Kenichi Yamasaki, Yasunori Shibata, Satoshi Imai, Yuji Tani, Yoshinobu Shibasaki and Tatsuo Fukuhara. Clinical application of prostaglandin E1 (PGE1) upon orthodontic tooth movement. *Am J Orthod Dentofacial Orthop*, 1984 Jun; 508-518.
12. K.D.Tripathi, Essentials of medical pharmacology. JP Brothers Medical Publishers P.Limit.New Delhi. 5th Edition,(P-168)
13. Brent Chumbley A. and Orhan C. Tuncay. The effect of indomethacin (an aspirin-like drug) on the rate of orthodontic tooth movement. *Am J Orthod Dentofacial Orthop* 1986; 89; 312-314.
14. Emel SarÄ Comparison of some effects of acetylsalicylic acid and rofecoxib during orthodontic tooth movement. *Am J Orthod Dentofacial Orthop*, March 2004; 125(3):310-315.
15. John.J.Roche, George Cisneros, George Acs, The effect of acetaminophen on tooth movement in rabbits. *Angle Orthod*, 1997;67(3):231-36 PMID:19537869.
16. Daniela Giunta, Johnny Keller, Frank FarsÄfÄ, Nielsen, BirteMelsen. Influence of indomethacin on bone turnover related to orthodontic tooth movement in miniature pigs. *Am J Orthod Dentofacial Orthop*, October 1995; 108(4):361-366. PMID:17878187 [http://dx.doi.org/10.1016/S0889-5406\(95\)70033-1](http://dx.doi.org/10.1016/S0889-5406(95)70033-1)
17. Carmen Gonzales, Hitoshi Hotokezaka, Ken-Ichiro Matsuo, Tatsunori Shibasaki, Joseph H. Yozgatian, M. Ali Darendeliler, Noriaki Yoshida. Effects of Steroidal and Nonsteroidal Drugs on Tooth Movement and Root Resorption in the Rat Molar. *Angle Orthod*, 2009;79(4);715-726. <http://dx.doi.org/10.2319/072108-381.1>
18. Felix de Carlos, Juan Cobo, Carmen Perillan, Miguel A. Garcia, Juan Arguelles, Manuel Vijande and Marina Costales. Orthodontic tooth movement after different coxib therapies. *European J Orthod*, 2007;29(6):596-599. <http://dx.doi.org/10.1093/ejo/cjm072>
19. Oscar R. Arias, Maria C. Marquez-Orozco. Aspirin, acetaminophen, and ibuprofen: Their effects on orthodontic tooth movement. *Am J Orthod Dentofacial Orthop*, September 2006; 130(3):364-370.
20. Williams R. C, Jeffcoat, M. K, Howell T. H, Hall C. M, Johnson, H. G, Wechter W. J and Goldhaber P. Indomethacin or flurbiprofen treatment of periodontitis in beagles: Comparison of effect on bone loss. *J. Periodont. Res*, 1987;22:403-407.
21. Michael J Kehoe, Steven M Cohen, Kourosh Zarrinnia, Alan Cowan. The effect of acetaminophen, ibuprofen, and misoprostol on prostaglandin E2 synthesis and the degree and rate of orthodontic tooth movement. *Angle Orthod*, 1996;66(5):339-350.
22. Felix de Carlos, Juan Cobo, Belen DÄfÄ-az-Esnal, Juan Arguelles, Manuel Vijande, Marina Costales. Orthodontic tooth movement after inhibition of

- cyclooxygenase-2. Am J Orthod Dentofacial Orthop, March 2006;129(3); 402-406. PMID:6932420
23. Bradley J, Liker B.J, Nanda R.S, currier G.F, Howes R I, Pramod K.Sinha. The effects of exogenous prostaglandins on orthodontic tooth movement in rats. Am J Orthod Dentofac Orthop, 1995; 108(4);380-88.
24. Elsdon Storey. The nature of tooth movement. Am J Orthod Dentofac Orthop. 1973;63(3);292-314
25. Richard S Masella, Malcolm Meister. Current concepts in the biology of orthodontic tooth movement. Am J orthod Dentofac Orthop,2006;129(4); 458-468.
26. Peter T T say, Min-Huey Chen, Ordean Oyen. Osteoclast activation and recruitment after application of orthodontic force. Am J orthod Dentofac Orthop, 1999; 115(3);323-330.
27. Kenichi Yamasaki, Fujio Miura, and Tatsuo Suda. Prostaglandin as a mediator of bone resorption induced by experimental tooth movement in rats. J.Dent.Res,October 1980; 59(10); 1635-1642.
<http://dx.doi.org/10.1177/00220345800590101301>
28. Selin Kale, Ilken Kocadereli, Pergin Atilla, Esin Asan. Comparison of the effects of 1,25 dihydroxycholecalciferol and prostaglandin E2 on orthodontic tooth movement. Am J orthod Dentofac Orthop, 2004; 125(5);607-614.
29. Daryl I. Boekenoogen, Pramod K. Sinha, Ram S. Nanda, Joydeep Ghosh, G FrÃfÃns Currier, Robert I Howes. The effects of exogenous prostaglandin E2 on root resorption in rats. Am J Orthod Dentofacial Orthop March 1996;109(3);277-286. PMID:19121496
30. Ali Reza sekhavat, Kazem Mousavizadeh, Hamid Reza Pakshir, Fatemeh Sari Asalani. Effect of misoprostol, a prostaglandin E1 analog on orthodontic tooth movement. Am J orthod Dentofacial Orthop, 2002; 122(5); 542-547.
31. Massoud Seifi, Behnam E Slami, Arash, Shoja, saffar. The effect of prostaglandin E2 and calcium gluconate on orthodontic tooth movement and root resorption in rats. Europ.J.Orthod.2003;25;199-204.
32. Burstone C.- The biomechanics of tooth movement. In: Kraus b. Vistas in orthodontics. Philadelphia, pa: lea & febiger 1962[197-213].
33. Theodosia Bartzela, Jens C. Turp, Edith Motschall, and Jaap C. Maltha. Medication effects on the rate of orthodontic tooth movement: A systematic literature review. Am J Orthod Dentofacial Orthop 2009;135(1);16-26.
<http://dx.doi.org/10.1016/j.ajodo.2008.08.016>
34. Gustavo Hauber, Gameiro, Joao Sarmiento Pereira-Neto, Maria Beatriz Borges De Araujo agnani, Darcy Flavio Nouer. The influence of Drugs and systemic factors on orthodontic tooth movement. JCO-2007. Vol XLI (2):73-78.
35. David L. Turpin. Medications weigh-in on tooth movement. Am J Orthod Dentofacial Orthop 2009;135: 139-140.
36. <http://pharmtech.findpharma.com/pharmtech/Manufacturing+and+Processing/Current-status-in-buccal-drug-delivery/ArticleStandard/Article/detail/514485>

Corresponding Author

Dr.K.Rajasigamani. M.D.S,
Professor , Head, Vice Principal,
Department Of Orthodontics and Dentofacial
Orthopedics,
Rajah Muthiah Dental College and Hospital,
Annamalai University, Annamalai Nagar,
Tamilnadu, India 608402.
Mobile: 9843177988
Email: rajasigamani.k_79@yahoo.com